Tigecycline Induction of Phenol-Soluble Modulins by Invasive Methicillin-Resistant *Staphylococcus aureus* Strains

Jason Yamaki  
*Chapman University*, yamaki@chapman.edu

Timothy Synold  
*City of Hope Medical Center*

Annie Wong-Beringer  
*University of Southern California*

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Tigecycline Induction of Phenol-Soluble Modulins by Invasive Methicillin-Resistant *Staphylococcus aureus* Strains

**Comments**
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We examined the effects of tigecycline on three types of exoproteins, α-type phenol-soluble modulins (PSMα1 to PSMα4), α-hemolysin, and protein A, in 13 methicillin-resistant Staphylococcus aureus isolates compared to those of clindamycin and linezolid. Paradoxical increases in PSMα4 occurred in 77% of the isolates with tigecycline at 1/4 and 1/8 MICs and clindamycin at 1/8 MIC compared to only 23% of the isolates with linezolid at 1/8 MIC. Induction was specific to PSMα4 as protein A and α-hemolysin production was decreased under the same conditions by all of the antibiotics used.

Methicillin-resistant Staphylococcus aureus (MRSA) virulence in pneumonia and bacteremia has been attributed to exoproteins, specifically, α-hemolysin (Hla) and α-type phenol-soluble modulins (PSMα1 to PSMα4), which are produced by nearly all S. aureus strains and in excess by community-acquired MRSA strains (1). These exoproteins not only cause direct damage to target host cells but also exacerbate the host inflammatory response, contributing to acute lung injury. Additionally, recent reports have demonstrated the importance of protein A (Spa) in invasive diseases such as pneumonia (2, 3).

In light of the impressive arsenal of virulence factors contributing to the success of MRSA as a pathogen, it is of keen interest to determine if anti-MRSA agents belonging to the antibiotic class of protein synthesis inhibitors provide the added antivirulence benefit of exoprotein inhibition. In the present study, we investigated the antivirulence potential of tigecycline, linezolid, and clindamycin which encode the Panton-Valentine leukocidin.

Specifically, we tested the effects of the above three antibiotics at subinhibitory concentrations on formylated PSMα4, produced at the baseline as very low to low (<1 μg/ml), low (1 to 5 μg/ml), medium (6 to 15 μg/ml), or high (>15 μg/ml).

Eleven invasive MRSA isolates were tested under the following conditions. A modified CLSI broth macrodilution assay was used to determine MICs after 24 h of incubation at 37°C and shaking at 250 rpm in tryptic soy broth. Supernatants were then analyzed by liquid chromatography-tandem mass spectrometry, and Hla and Spa were analyzed by Western blotting as previously described (4, 5). Measured exotoxin values were normalized to the cell optical density at 600 nm (OD600) at the time the supernatant was harvested. PSMα concentrations under various test conditions were compared by analysis of variance with Dunn’s correction using GraphPad Prism version 5.0 software (GraphPad, San Diego, CA).

Table 1 depicts the SCCmec type, PVL status, and baseline PSMα production characteristics of the 13 isolates studied (11 clinical isolates and two control strains). The PSMα1 to PSMα4 peptides have been shown to cause concentration-dependent neutrophil lysis (6, 7). A PSMα4 concentration of 5 μg/ml has been shown to lyse nearly 50% of human polymorphonuclear neutrophils (PMNs), while 10 μg/ml of PSMα3 and PSMα4 can cause 60% and 10% PMN lysis, respectively (8, 9). Thus, isolates were grouped on the basis of the amount of the most potent peptide, PSMα4, produced at the baseline as very low to low (≤5 μg/ml), medium (6 to 15 μg/ml), or high (>15 μg/ml) producers.

Bacterial growth at 1/2 MICs of all three antibiotics was altered in half of the isolates tested by as much as 50% of the final OD compared to a no-antibiotic control, while no significant difference in the growth of any isolates was observed at 1/4 and 1/8 MICs. Thus, subsequent discussions will focus on the effect of antibiotics at the latter subinhibitory concentrations that did not affect growth. Measured PSMα values were normalized to the OD600 at the time the supernatant was harvested.
Tigecycline appears to have the least inhibitory potential overall, while linezolid has the greatest (Fig. 1a to c). Increased production of all four PSM\textsubscript{1} to PSM\textsubscript{4} peptides was the primary response observed in the presence of tigecycline at 1/4 and 1/8 MICs, while with clindamycin this occurred only at 1/8 MIC (Fig. 1b and c). Notably, compared to clindamycin at 1/8 MIC, which significantly induced PSM\textsubscript{1} production in seven isolates, linezolid at 1/8 MIC modestly induced PSM\textsubscript{1} production in only three isolates (Fig. 1a).

Strain-specific responses to the presence of subinhibitory antibiotic concentrations in PSM\textsubscript{1} to PSM\textsubscript{4} production were observed. Drug concentrations that induced PSM\textsubscript{1} to PSM\textsubscript{4} production in some isolates did not do so in others, independently of the baseline production level. Specifically, with tigecycline at both 1/4 and 1/8 MICs (Fig. 1d), PSM\textsubscript{1} was induced at least 1.5 times above the baseline in 77% (11/13) of the isolates, while induction did not occur in two isolates at any of the concentrations tested. Similar results were observed with clindamycin, where 54% (7/13) of the isolates were induced by greater than 150%; however, this was observed only at 1/8 MIC. In those isolates, PSM\textsubscript{1} production was induced to greater than 10 \(\mu\)g/ml, which has been shown to cause significant PMN lysis (9). Linezolid at the same concentration resulted in increases in PSM\textsubscript{1} in only 23% (3/13) of the isolates tested, was inhibitory in 5 isolates, and had no significant effects on the remaining isolates. Of additional interest is the observation that nonproducers at the baseline produced PSM\textsubscript{1} peptides in the presence of subinhibitory concentrations of both linezolid and tigecycline. However, it is noteworthy that PSM\textsubscript{1} production that was induced in those two isolates did not exceed 4 \(\mu\)g/ml and thus would not be expected to have a significant impact on host PMNs.

To our knowledge, we are the first to investigate the effect of tigecycline on MRSA PSM\textsubscript{1} to PSM\textsubscript{4} peptide production. We also included clindamycin and linezolid for comparison and tested their antivirulence potential against two other key exotoxins, Hla and Spa. We found that the production of both Hla and Spa did not increase under any condition and was inhibited in a dose-dependent manner at the concentrations of all three antibiotics tested regardless of whether PSM\textsubscript{1} peptides were induced or suppressed in those isolates (Fig. 2).

FIG 1 Overall effects of linezolid, clindamycin, and tigecycline on PSM\textsubscript{1} to PSM\textsubscript{4} production. (a to c) Effects of antibiotics on the production of PSM\textsubscript{1} to PSM\textsubscript{4} (a\textsubscript{1}, a\textsubscript{2}, a\textsubscript{3}, and a\textsubscript{4}, respectively). All measured PSM values (\(\mu\)g/ml) were normalized to OD\textsubscript{600} and then calculated as percentages relative to the baseline. Isolates (n = 10) included only those that produced measureable PSM\textsubscript{1} peptides at the baseline (***, P < 0.0001; **, P < 0.001; *, P < 0.05). (d) Strain-to-strain variability of PSM\textsubscript{1} production at 1/8 MIC of tigecycline (TYG). Strains were grouped according to their baseline PSM\textsubscript{1} production as very low (<1 \(\mu\)g/ml), low (1 to 5 \(\mu\)g/ml), medium (6 to 15 \(\mu\)g/ml), and high (>15 \(\mu\)g/ml) producers.
pneumonia, respectively, which are below the 0.5-µg/ml FDA MIC breakpoint of *S. aureus* susceptibility (10–12).

Low concentrations (1/4 and 1/8 MICs) of the antibiotics tested were more likely to induce PSMx production overall, with tigecycline having the greatest induction potential, clindamycin having less, and linezolid even less than that. Furthermore, the magnitudes of induction and inhibition differed among isolates, where the same drug could induce PSMx production in one isolate while inhibiting production in another. The observation of PSMx induction by protein synthesis-inhibiting antibiotics (e.g., linezolid and clindamycin) at sub-MICs is contrary to published literature on other exotoxins, where studies have consistently found toxin suppression under this class of antibiotics (13–16). The production of both Hla and Spa was suppressed by antibiotics tested at subinhibitory concentrations among the study isolates, independently of whether PSMx was induced in the same isolates. Hence, our results indicate that the induction effects may be specific for PSMx peptides. In contrast to our findings where significant inhibition was observed, a recent article showed tigecycline to be minimally inhibitory of Hla and Spa production (17); however, this difference may be due to the different strains and testing methods used.

Our study has several limitations. First, while we accounted for changes in cell counts by normalizing the amount of toxin measured to CFU counts, other factors, such as initial slowing of growth, could also affect toxin production. In addition, our observations are strictly *in vitro* and may not reflect how MRSA strains express virulence in the host environment, where the effects of toxin induction may be mitigated by host defenses that are present.

Further investigation is needed to examine if any other major toxins are induced, as PSMx peptides did when exposed to subinhibitory concentrations of different antibiotics. Elucidating the mechanism that underlies the induction of PSMx peptides is of great interest and has implications for the future development of new antibiotics. In light of the recent evidence implicating tigecycline in excess deaths and treatment failure in severe infections (18), the enhanced toxin production observed in this study may have clinical relevance and deserves further study. For now, our data caution against the general assumption by clinicians that all protein synthesis inhibitors possess inhibitory potential against all of the exotoxins produced by *S. aureus* and affirm the importance of adequate dosing.

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FIG 2 Effects of tigecycline on Hla and Spa production. (a) Representative images of Western blot assays of Hla and Spa, respectively. (b and c) Production of Hla and Spa in the presence of 1/8 and 1/4 MICs of tigecycline relative to the baseline. In no case was Hla or Spa production increased by subinhibitory concentrations of any of the antibiotics tested.

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