

2010

Impaired M3 and Enhanced M2 Muscarinic Receptor Contractile Function in a Streptozotocin Model of Mouse Diabetic Urinary Bladder

K. J. Pak

University of California, Irvine

Rennolds S. Ostrom

Chapman University, rostrom@chapman.edu

M. Matsui

Tokyo-Nishi Tokushukai Hospital

F. J. Ehlert

University of California, Irvine

Follow this and additional works at: http://digitalcommons.chapman.edu/pharmacy_articles

 Part of the [Amino Acids, Peptides, and Proteins Commons](#), [Cell Biology Commons](#), and the [Other Pharmacy and Pharmaceutical Sciences Commons](#)

Recommended Citation

Kirk J. Pak, Rennolds S Ostrom, Minoru Matsui and Frederick J Ehlert. Impaired M3 and enhanced M2 muscarinic receptor contractile function in a streptozotocin model of mouse diabetic urinary bladder. *Naunyn-Schmiedebergs Arch Pharmacol*, 381(5):441-54, 2010.

This Article is brought to you for free and open access by the School of Pharmacy at Chapman University Digital Commons. It has been accepted for inclusion in Pharmacy Faculty Articles and Research by an authorized administrator of Chapman University Digital Commons. For more information, please contact laughtin@chapman.edu.

Impaired M3 and Enhanced M2 Muscarinic Receptor Contractile Function in a Streptozotocin Model of Mouse Diabetic Urinary Bladder

Comments

This article was originally published in *Naunyn-Schmiedeberg's Archives of Pharmacology*, volume 381, issue 5, in 2010. DOI: [10.1007/s00210-010-0509-6](https://doi.org/10.1007/s00210-010-0509-6)

Copyright

The authors

Impaired M_3 and enhanced M_2 muscarinic receptor contractile function in a streptozotocin model of mouse diabetic urinary bladder

K. J. Pak · R. S. Ostrom · M. Matsui · F. J. Ehlert

Received: 8 December 2009 / Accepted: 25 February 2010 / Published online: 27 March 2010
© The Author(s) 2010. This article is published with open access at Springerlink.com

Abstract We investigated the contractile roles of M_2 and M_3 muscarinic receptors in urinary bladder from streptozotocin-treated mice. Wild-type and M_2 muscarinic receptor knockout (M_2 KO) mice were given a single injection of vehicle or streptozotocin (125 mg kg^{-1}) 2–24 weeks prior to bladder assays. The effect of forskolin on contractions elicited to the muscarinic agonist, oxotremorine-M, was measured in isolated urinary bladder (intact or denuded of urothelium). Denuded urinary bladder from vehicle-treated wild-type and M_2 KO mice exhibited similar contractile responses to oxotremorine-M, when contraction was normalized relative to that elicited by KCl (50 mM). Eight to 9 weeks after streptozotocin treatment, the EC_{50} value of oxotremorine-M increased 3.1-fold in urinary bladder from the M_2 KO mouse ($N=5$) compared to wild type ($N=6$; $P<0.001$). Analogous changes were observed in intact bladder. In denuded urinary bladder from vehicle-treated mice, forskolin ($5 \mu\text{M}$) caused a much greater inhibition of contraction in M_2 KO bladder compared to wild type. Following streptozotocin treatment, this forskolin effect increased 1.6-fold ($P=0.032$). At the

20- to 24-week time point, the forskolin effect increased 1.7-fold for denuded as well as intact bladders ($P=0.036$, 0.01, respectively). Although streptozotocin treatment inhibits M_3 receptor-mediated contraction in denuded urinary bladder, muscarinic contractile function is maintained in wild-type bladder by enhanced M_2 contractile function. M_2 receptor activation opposes forskolin-induced relaxation of the urinary bladder, and this M_2 function is enhanced following streptozotocin treatment.

Keywords M_2 muscarinic receptor · Urinary bladder · Contraction · Streptozotocin · Diabetes · Smooth muscle

Introduction

Diabetes is associated with an alteration in the contractility of urinary bladder smooth muscle in addition to the well-known increase in urinary frequency (Longhurst and Belis 1986; Latifpour et al. 1989; Michel and Barendrecht 2008). The two main neurotransmitters eliciting contraction in human and rodent urinary bladder are adenosine triphosphate and acetylcholine, which act on a ligand-gated ion channel (P2X receptor) and two G-protein-coupled receptors (M_2 and M_3 muscarinic), respectively (de Groat and Yoshimura 2001). A common approach for studying how the functions of these receptors change in diabetes mellitus is to examine their contractile roles in isolated urinary bladder from animals treated with streptozotocin (STZ).

Treatment with STZ causes a hypertrophy of the rat urinary bladder (Longhurst and Belis 1986; Latifpour et al. 1989) and an upregulation in M_2 and M_3 muscarinic receptors in the urothelium and smooth muscle (Tong and Cheng 2002; Tong et al. 2006; Cheng et al. 2007). Not surprisingly, the contractile response to muscarinic agonists

K. J. Pak · F. J. Ehlert (✉)
Department of Pharmacology, School of Medicine,
University of California,
Irvine, CA 92697-4625, USA
e-mail: fjehlert@uci.edu

R. S. Ostrom
Department of Pharmacology,
University of Tennessee Health Science Center,
Memphis, TN 38163, USA

M. Matsui
Department of Clinical Research and General Medicine,
Tokyo-Nishi Tokushukai Hospital,
Tokyo, Japan

in whole urinary bladder increases following STZ treatment (Kolta et al. 1985; Nsabimana and Ojewole 2008). To control for the STZ-induced hypertrophy, several investigators have normalized contraction relative to the cross-sectional area of the muscle strip or have measured contractions in strips of urinary bladder having a similar cross-sectional area. These studies have yielded conflicting reports, however, including an increase (Latifpour et al. 1989), a decrease (Longhurst and Belis 1986), and no change (Lincoln et al. 1984) in muscarinic agonist-induced contraction. Whether there are specific changes in the contractile functions of M_2 and M_3 muscarinic receptors in urinary bladder from STZ-treated rodents is unknown.

In smooth muscle from the urinary bladder and several other organs, the M_2 receptor outnumbers the M_3 by a ratio that is often at least four to one, yet its function at the level of contraction is less than obvious (Ehlert 2003a, b). For example, it has long been known that subtype-selective muscarinic antagonists inhibit contraction of the urinary bladder with a profile consistent with that of an M_3 mechanism (Hegde and Eglén 1999). In addition, there is little loss of muscarinic agonist-induced contraction in isolated urinary bladder from the M_2 muscarinic receptor knockout out (M_2 KO) mouse (Matsui et al. 2000, 2002). In smooth muscle from the M_3 KO mouse, muscarinic agonists elicit contraction, and this response is lost in the M_2/M_3 double KO mouse, indicating that the M_2 receptor can mediate contraction. Although the potency of this response is high, its E_{\max} expressed relative to that of wild type is only 10% in urinary bladder and about 30% in ileum and the fundus of the stomach (Matsui et al. 2000; Stengel et al. 2002). Thus, experiments on the M_3 KO mouse have revealed a small role for the M_2 receptor in contraction. In guinea pig ileum, this high-potency M_2 mechanism is absent (Griffin et al. 2009).

Part of the reason why the M_2 receptor appears to have little role in the experiments described above can be explained by the nature of some of the G_i -signaling pathways of the M_2 receptor in smooth muscle. Muscarinic activation of smooth muscle elicits a Ca^{2+} -dependent nonselective cation conductance (I_{cat} ; Bolton 1979; Inoue 1991). This conductance is pertussis toxin-sensitive (Inoue and Isenberg 1990), and it is greatly inhibited in intestinal smooth muscle from either the M_2 KO or the M_3 KO mouse (Sakamoto et al. 2007). Muscarinic receptor activation indirectly stimulates a Ca^{2+} -activated K^+ (BK_{Ca}) channel in smooth muscle from guinea pig trachea (Wade and Sims 1993) and many other organs, presumably through mobilization of Ca^{2+} by the M_3 receptor. This conductance may represent an inhibitory-feedback mechanism to suppress excessive excitation of smooth muscle. Muscarinic receptor activation in canine colon

(Cole and Sanders 1989) and many other types of smooth muscle also directly inhibits the BK_{Ca} channel, and this response is pertussis toxin-sensitive (Kume et al. 1995; Nakamura et al. 2002), which suggests a role for the M_2 receptor. In rat urinary bladder, pertussis toxin-treatment and M_2 selective antagonists inhibit the direct muscarinic suppression of BK_{Ca} (Nakamura et al. 2002). Both the M_2 inhibition of the BK_{Ca} channel and the M_2 stimulation of I_{cat} are expected to be lost in the M_3 KO mouse, because they are contingent upon Ca^{2+} mobilization triggered by M_3 receptor activation. Perhaps this accounts for the near complete loss of muscarinic contractile function in urinary bladder from the M_3 KO mouse. Obviously, this M_2 – M_3 interaction mechanism cannot account for the residual M_2 receptor-mediated contraction in the ileum and fundus of the stomach from the M_3 KO mouse.

One might expect some decrease in muscarinic agonist-induced contraction in smooth muscle from the M_2 KO mouse or from pertussis toxin-treated animals because of the loss of M_2 conductance mechanisms just described. In most instances, however, this is not observed (Thomas and Ehlert 1994; Sawyer and Ehlert 1999; Ehlert 2003a). We have previously shown that the potency of the M_2 enhancement mechanism in intestinal (Sawyer and Ehlert 1999; Ehlert 2003a) and urinary bladder (Ehlert et al. 2005, 2007) smooth muscle is less than that of the M_3 contractile mechanism so that muscarinic contractions are already maximal at an agonist concentration that is too low to trigger M_2 mechanisms. One exception is the mouse uterus where muscarinic agonist-induced contraction is greatly inhibited in the M_2 KO mouse (Kitazawa et al. 2008).

In smooth muscle, cAMP mediates relaxation, and activation of the M_2 receptor inhibits adenylate cyclase and the relaxation elicited by forskolin and activation of β -adrenoceptors in gastrointestinal and urinary bladder smooth muscle (Ehlert 2003b). The potency of muscarinic agonists for eliciting this response in the guinea pig ileum is actually greater than that for eliciting a direct contraction through the M_3 receptor (Ostrom and Ehlert 1997). In bovine and guinea pig trachea, M_2 receptor activation inhibits forskolin- but not isoproterenol-mediated relaxation, suggesting that the β -adrenoceptor mediates relaxation through a non-cAMP mechanism (Ostrom and Ehlert 1998).

The M_3 receptor interacts with G_q to mediate phosphoinositide hydrolysis and direct contraction in smooth muscle (Ehlert 2003b). Both the effect on phosphoinositide hydrolysis and contraction are pertussis toxin-insensitive (Ehlert 2003b). While many G_q -linked receptors elicit contraction in smooth muscle, the source of Ca^{2+} for M_3 receptor-mediated contractions is mainly extracellular through voltage-sensitive Ca^{2+} channels (Bolger et al. 1983). How the M_3 receptor triggers an influx of Ca^{2+} is unclear. Most of the older electrophysiological literature

suggests that the initial depolarization occurs through I_{cat} followed by an influx of Ca^{2+} through voltage-sensitive channels, but neither the activation of I_{cat} nor the inhibition of BK_{Ca} channels can explain the potent muscarinic agonist-induced contraction of isolated smooth muscle because the latter is insensitive to pertussis toxin (Ehlert 2003b), whereas the former two ionic mechanisms are pertussis toxin-sensitive (see above).

Given the importance of the STZ model of diabetes, we undertook a study to determine how M_2 and M_3 receptor-mediated contractions are altered in isolated urinary bladder from STZ-treated mice. We observed an increase in M_2 and a decrease in M_3 receptor-mediated contractions in urinary bladder smooth muscle from STZ-treated mice. We also observed a similar phenomenon in urinary bladder with intact urothelium, although the results are more complicated because of changes in muscarinic function in the urothelium with STZ treatment.

Methods

Animals

All procedures were approved by the Institutional Laboratory Animal Care and Use Committee. M_2 muscarinic receptor knockout (M_2 KO) mice were generated as previously described (Matsui et al. 2000, 2002). Male C57BL/6 wild-type (Harlan Sprague Dawley, Inc., Indianapolis, IN, USA) and M_2 KO mice (2–3 months old, 18–30 g) were fasted for 4–6 h and then briefly anesthetized with isoflurane (Phoenix Pharmaceuticals, Inc., St. Joseph, MO, USA) before a single intraperitoneal injection with either vehicle (sodium citrate dihydrate, Fisher Scientific, Pittsburgh, PA, USA) or 125 mg kg^{-1} of STZ (Sigma Aldrich Chemical, St. Louis, MO, USA) 2 to 24 weeks prior to experiments on isolated bladder. Water containing 10% sucrose was given to animals 48 h after injection to prevent hypoglycemic shock. All mice were housed in a 12-h light/dark facility with free access to water and food. Blood glucose was measured using an Ascensia Contour glucometer (Bayer, Leverkusen, Germany), and diabetic ketoacidosis was assessed by Keto-Diastix (Bayer, Leverkusen, Germany). Heparinized saline was used to dilute and estimate blood glucose levels above the 600-mg/dL glucometer limit.

Isolated urinary bladder

Whole mouse urinary bladder was removed and cut in half sagittally. The urothelium was carefully dissected from one of the strips, while the other was left intact. Half-bladder strips were mounted in an organ bath in a longitudinal orientation and connected to a force-displacement trans-

ducer using silk thread. Tissues were immersed in a Krebs–Ringer bicarbonate buffer (124 mM NaCl, 5 mM KCl, 1.3 mM $MgSO_4$, 26 mM $NaHCO_3$, 1.2 mM KH_2PO_4 , 1.8 mM $CaCl_2$, and 10 mM glucose) at 37°C and gassed with O_2/CO_2 (19:1) as previously described (Ehlert et al. 2005). Resting tension was adjusted to a load of 1 g (9.8 mN) during an equilibration period of at least 1 h. Contractile responses were first generated by two test doses of KCl (50 mM). Bladder was washed and allowed to rest for 10 min after each test dose. The contractile response to KCl was calculated as the stable plateau value after a 3-min period minus the initial resting tension. Responses to other agents were either expressed in millinewton of force or normalized relative to the larger of the two responses to KCl, which were usually similar.

Contraction of the urinary bladder to the highly efficacious muscarinic agonist, oxotremorine-M, was measured using a cumulative technique. Each aliquot of the agonist was added to the bath, and the response was calculated as the stable plateau achieved within approximately 1 min minus the initial resting tension. Figure 1 shows a representative recording with the arrows indicating the final cumulative concentrations of agonist. After a complete concentration–response curve was measured, tissues were washed five to eight times over a period of 35–45 min before measurement of another curve. This initial concentration–response curve was not utilized but enabled the bladder to equilibrate and to achieve a maximal contractile state. Subsequent responses were recorded in a similar manner and are described under “Results” section.

Analysis of oxotremorine-M concentration–response curves

The maximal response (E_{max}) and concentration of agonist eliciting half-maximal response (EC_{50}) were estimated from the concentration–response curve using nonlinear regression

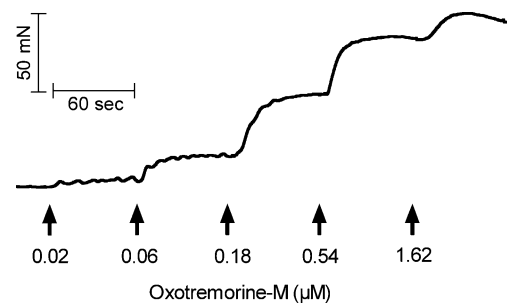


Fig. 1 Contractile response of denuded urinary bladder to oxotremorine-M measured using a cumulative technique. The urinary bladder was from a wild-type mouse that had been treated with streptozotocin approximately 20 weeks prior. Force of contraction of this representative experiment is plotted against time. The arrows indicate when aliquots of oxotremorine-M were added to the tissue bath. The final, cumulative concentration of oxotremorine-M is indicated beneath each arrow

analysis with Prism software (GraphPad Software, Inc., San Diego, CA, USA).

To estimate the potential loss of function in the M₂ KO mouse bladder relative to wild type, we fitted the concentration–response curves from wild-type and M₂ KO mice to Eqs. 1 and 2, respectively:

$$y = \frac{X^n E_{\max}}{X^n + EC_{50}^n} \quad (1)$$

$$y = \frac{X^n (E_{\max} + \delta)}{X^n + \gamma EC_{50}^n} \quad (2)$$

In both of these equations, EC₅₀ and E_{max} represent the corresponding estimates for wild-type tissue, *y* represents the measured contraction, *X* represents the concentration of oxotremorine-M, and *n* represents the Hill slope. In Eq. 2, δ represents the change in E_{max}, and γ represents the scalar change in EC₅₀ between wild type and M₂ KO. For each treatment (i.e., vehicle or STZ), the concentration–response curves were fitted simultaneously to Eqs. 1 and 2 sharing the estimate of E_{max} and EC₅₀ between wild type and M₂ KO and obtaining unique estimates of *n* for both wild type and M₂ KO and unique estimates of δ and γ for the M₂KO. To test for differences between bladders from vehicle- and STZ-treated animals, we determined the significance of the increase in residual variance when the estimate of δ or γ was shared between the vehicle- and STZ-treated groups as described by Motulsky and Christopoulos (2003).

We used the operational model (Black and Leff 1983) to quantify the effect of forskolin on the concentration–response curve to oxotremorine-M:

$$y = \frac{X^m M_{\text{sys}}}{X^m + \frac{(X+K)^m}{\tau^m}} \quad (3)$$

In this equation, *y* represents the measured contraction, *X* represents the concentration of oxotremorine-M, M_{sys} represents the maximum response of the system, *K* represents the observed dissociation constant, *m* represents the transducer slope factor, and τ represents a parameter related to coupling efficiency. Specifically, the value of τ is proportional to the intrinsic efficacy of the agonist (ε), the amount of potentially active receptors (*R_T*), and the sensitivity of the signaling cascade (1/*K_E*). In our experiments, forskolin either caused a shift to the right or both a shift to the right and a decrease in the E_{max} of the concentration–response curve. Both situations were adequately described by a reduction in the τ parameter of the operational model. The effects of forskolin on the two disparate parameters, EC₅₀ and E_{max}, could be adequately described, therefore, by a decrease in the value of a single parameter (τ) that accurately accounts for the

inhibitory effect of cAMP on the contractile elements (i.e., decrease in 1/*K_E*).

To quantify this effect in bladder from the wild-type mouse, we analyzed the control concentration–response curve using Eq. 3 and that measured in the presence of forskolin using an identical equation, but with the parameter τ replaced with the product, $\alpha\tau$:

$$y = \frac{X^m M_{\text{sys}}}{X^m + \frac{(X+K)^m}{\alpha\tau^m}} \quad (4)$$

The parameter α , therefore, represents the factor by which forskolin reduces the sensitivity of the signaling pathway for contraction.

To investigate how the M₂ receptor opposes forskolin-induced relaxation, we compared the relaxant effect of forskolin in urinary bladder from wild-type and M₂ KO mice. The control concentration–response curve in the M₂ KO mouse was analyzed using Eq. 3 and that in the presence of forskolin was analyzed using an identical equation, but with the parameter τ replaced with the product, $\alpha\beta\tau$:

$$y = \frac{X^m M_{\text{sys}}}{X^m + \frac{(X+K)^m}{\alpha\beta\tau^m}} \quad (5)$$

Here, β represents the relative difference, expressed as a factor, between the effect of forskolin in wild-type (α) and M₂ KO mice ($\alpha\beta$). This parameter, therefore, is a measure of the role of the M₂ receptor in opposing forskolin-induced relaxation—the smaller the value of β (or alternatively, the larger the value of $-\log \beta$), the greater the role of the M₂ receptor.

To estimate β , the concentration–response curves in the absence (control) and presence of forskolin and in both wild-type and M₂ KO mice were fitted simultaneously to Eqs. 3, 4, and 5 to obtain unique estimates of M_{sys}, *m*, α , and β (global nonlinear regression analysis). The estimate of τ was shared among the curves and that of *K* was shared between those of control and forskolin conditions. Figure 2 shows a pictorial representation of our analysis, and the legend describes additional details.

The statistical significance of differences between parameter estimates measured in urinary bladders from vehicle- and STZ-treated mice was determined as described above in connection with Eqs. 1 and 2.

Results

Effects of STZ on body weight and blood glucose

Wild-type and M₂ KO mice injected with a single dose (125 mg kg⁻¹) of STZ exhibited significantly higher fasting

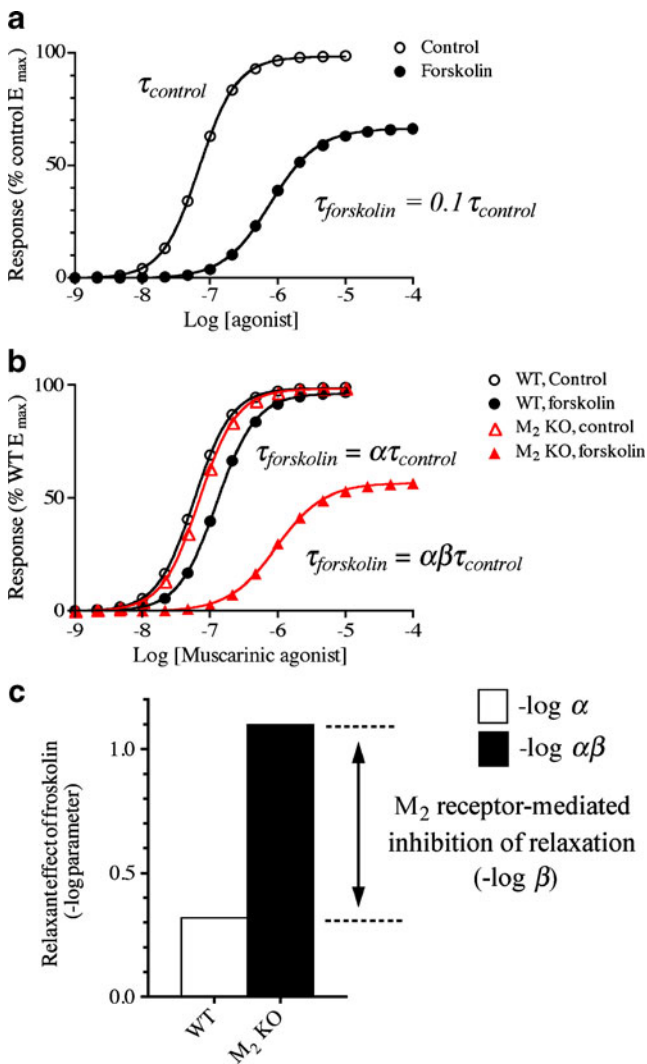


Fig. 2 Determination of the role of the M_2 receptor in opposing forskolin-induced relaxation. **a** The plot shows a simulation of a concentration–response curve to a muscarinic agonist in the absence (control) and presence of forskolin. The operational model (Eq. 3) was used to simulate the data. All of the parameters were the same for the two curves except that the parameter τ for the curve measured in the presence of forskolin was reduced to one tenth that of the control curve. The values of the parameters in the simulation were $M_{\text{sys}}=100$, $m=1.65$, $K=10^6$, and $\tau=15$. **b** The simulation illustrates the effect of forskolin on the concentration–response curve to a muscarinic agonist in smooth muscle from wild-type and M_2 KO animals. For the wild-type curves, the effect of forskolin has been simulated by reducing the value of τ by the factor α . In other words, Eqs. 3 and 4 were used to simulate the control and forskolin conditions, respectively, with the value of α set to 0.48 in Eq. 4 and the values of all of the other parameters the same for the two equations. For the M_2 KO curves, the effect of forskolin has been simulated by reducing τ by the factor $\alpha\beta$, with $\alpha=0.48$ as in wild type and $\beta=0.16$. It follows that the log difference in the effect of forskolin in wild type ($\log \alpha$) and M_2 KO ($\log \alpha\beta$) is equivalent to $\log \beta$. The values of the parameters for the simulations were, wild-type control, $M_{\text{sys}}=100$, $m=1.65$, $K=10^6$, and $\tau=18$; wild-type forskolin, same as control with $\alpha=0.48$; M_2 KO control, same as wild type but with $\log \tau=15$; M_2 KO forskolin, same as M_2 KO but with $\alpha\beta=0.078$. **c** The plot shows that the role of the M_2 receptor in opposing forskolin-induced relaxation ($\log \beta$) is equivalent to the difference between the relaxant effect of forskolin in wild type ($\log \alpha$) and M_2 KO ($\log \alpha\beta$). The values of α and β are the same as those used in the simulation in (b)

blood glucose levels compared to control animals (Table 1). Body weight decreased significantly more in STZ-treated mice compared to those treated with vehicle with the exception of wild-type mice at the 2- to 3-week time point (Table 1). Although STZ significantly decreased the weight of M_2 KO compared to wild-type mice at 8 to 9 weeks (Table 1), denuded bladder tissue from these animals did not exhibit a difference in KCl-induced contraction (data not shown).

Effects of STZ on denuded urinary bladder

Muscarinic agonist-induced contractions Two to 3 weeks after treatment with STZ, wild-type urinary bladder lacking urothelium (denuded) displayed a slight decrease in contractile responsiveness to oxotremorine-M as compared to tissue from vehicle-treated mice (Fig. 3a). At both the 8–9- and 20–24-week time points after STZ treatment, however, the E_{max} values of oxotremorine-M in denuded

bladders were markedly greater than those from vehicle-treated animals (Fig. 3d, g). These effects were manifest mainly as significant 58% and 61% increases in E_{max} when expressed in units of force (millinewton; Table 2). These data illustrate a time-dependent increase of contractile response in tissues from STZ-treated mice.

When normalized relative to the contraction elicited by KCl (50 mM), however, the concentration–response curves in bladder from wild-type mice showed little difference between vehicle and STZ groups at all time points (Fig. 3b, e, h and Table 2), suggesting that STZ treatment might cause smooth muscle hypertrophy or a general increase in contractility over time. In experiments on urinary bladder from M_2 KO mice, there was little difference between vehicle and STZ groups at the early (2–3 weeks) and late (20–24 weeks) time points (Fig. 3c, i). At the intermediate time point (8–9 weeks), STZ treatment caused a significant loss of contractile function (Fig. 3f), which was manifest as a significant 2.3-fold increase in EC_{50} (Table 2).

To quantify the role of the M_2 receptor, we compared the loss of contractile function in the M_2 KO mouse relative to wild type. In vehicle-treated mice, there was little difference in the contractile activity of oxotremorine-M in urinary bladder from wild-type and M_2 KO mice. Similarly, there was little difference in contractile activity between wild-type and M_2 KO mice at the early (2–3 weeks) and late (20–24 weeks) time points after STZ treatment. At the intermediate time point (8–9 weeks), however, STZ

Table 1 Body weight percent change and fasting blood glucose levels 2–24 weeks after injection

		Percent body weight change		Fasting blood glucose (mgdL ⁻¹)	
		Wild type	M ₂ KO	Wild type	M ₂ KO
2–3 weeks after treatment	Vehicle	5.9±2.7 (7)	1.2±1.2 (3)	111.1±14.7 (7)	113.7±18.5 (3)
	STZ	1.4±8.5 (3)	-9.2±1.0 (3) ^a	298.3±59.2 (3) ^a	436.0±39.7 (3) ^a
8–9 weeks after treatment	Vehicle	27.0±2.4 (6)	21.1±3.5 (4)	115.0±11.2 (6)	140.8±8.9 (4)
	STZ	14.3±2.4 (6) ^a	4.36±1.4 (5) ^{a, b}	431.3±61.3 (6) ^c	470.6±37.9 (5) ^c
20–24 weeks after treatment	Vehicle	48.2±6.7 (5)	41.2±1.8 (4)	133.0±5.3 (5)	127.5±6.3 (4)
	STZ	14.4±8.3 (3) ^a	8.9±4.2 (4) ^c	458.7±113.3 (3) ^a	513.7±36.3 (4) ^c

Vehicle (sodium citrate-buffered saline) and streptozotocin (STZ) were injected intraperitoneally (125 mg kg⁻¹). Data are means ± SE (number of animals)

^a Statistically significant difference from vehicle ($P < 0.05$)

^b Significantly different from STZ-treated wild-type mice ($P = 0.011$)

^c Statistically significant difference from vehicle ($P < 0.001$)

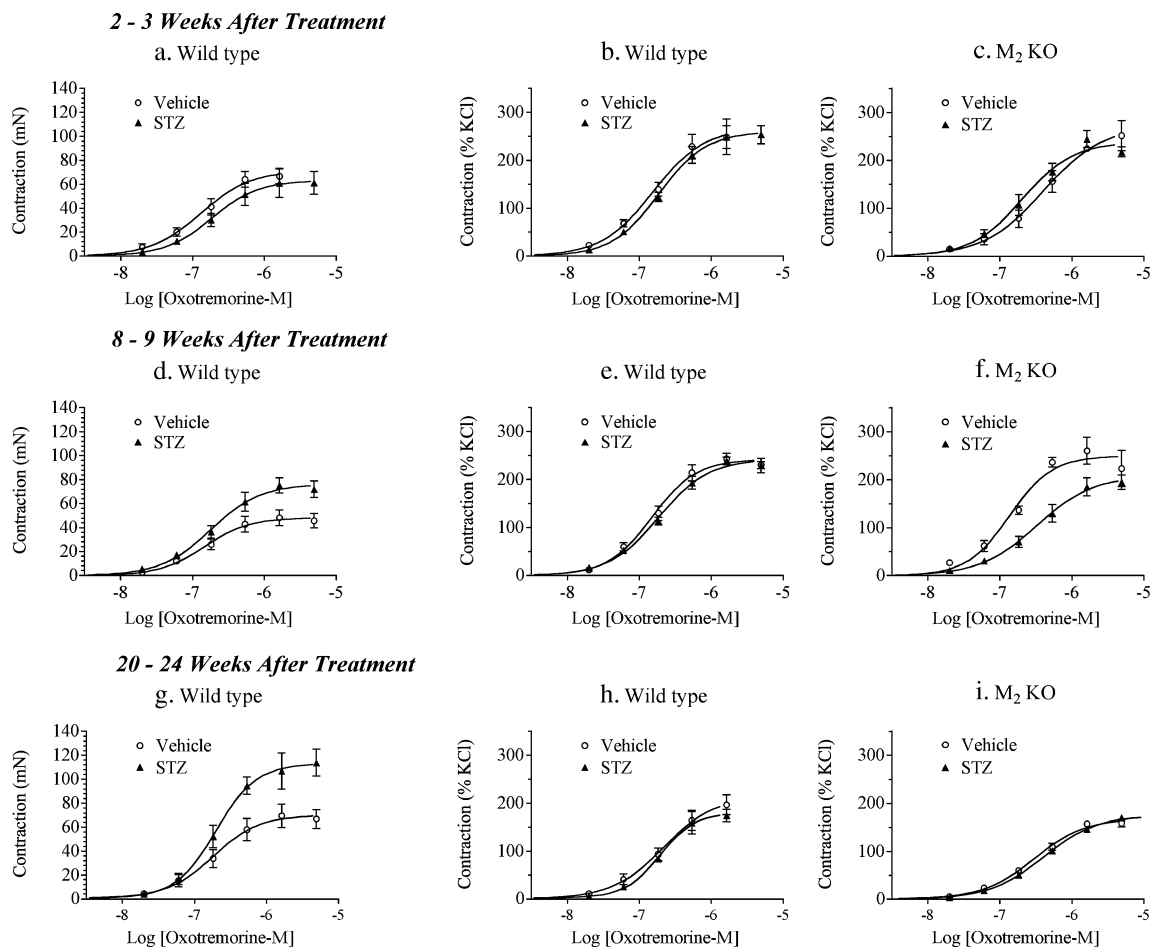


Fig. 3 Effect of streptozotocin (STZ) treatment on oxotremorine-M-mediated contractions in urinary bladder lacking urothelium from wild-type and M₂ KO mice. Concentration–response curves were measured 2 to 3 weeks (a–c), 8 to 9 weeks (d–f), and 20 to 24 weeks (g–i) after STZ treatment in urinary bladder from wild-type (a, b, d, e,

g, h) and M₂ KO (c, f, i) mice. Contractions are expressed in units of force (millinewton; a, d, g) or relative to the contraction elicited by KCl (50 mM; b, c, e, f, h, i). Circles denote the vehicle-treated group; triangles denote the STZ-treated group. Mean ± SEM values from three to seven experiments are shown, each done on a separate bladder

Table 2 E_{\max} and pEC_{50} values measured in denuded bladders from mice 2–24 weeks after injection

Denuded bladder	2–3 weeks			8–9 weeks			20–24 weeks		
	E_{\max} (mN)	E_{\max} (% KCl)	pEC_{50}	E_{\max} (mN)	E_{\max} (% KCl)	pEC_{50}	E_{\max} (mN)	E_{\max} (% KCl)	pEC_{50}
Vehicle	WT	268.10±30.43	6.81±0.14	48.17±3.76	242.20±8.82	6.83±0.052	70.27±6.15	215.60±26.79	6.66±0.14
	M ₂ KO	272.60±26.26	6.39±0.13	–	249.50±15.35	6.86±0.088	–	168.30±6.51	6.53±0.051
STZ	WT	63.12±6.05	6.73±0.064	76.28±4.94 ^b	242.60±8.93	6.73±0.052	113.30±7.12 ^b	181.70±12.84	6.72±0.071
	M ₂ KO	–	6.69±0.087	–	208.60±17.21	6.49±0.11 ^a	–	175.30±3.08	6.40±0.027

Data are means ± SE (number of animals)

^a Statistically significant difference from M₂ KO vehicle ($P<0.01$)^b Significantly different from wild-type vehicle ($P<0.001$)

induced a greater loss of function in urinary bladder from the M₂ KO mouse (Fig. 3f) compared to that from wild type (Fig. 3e), which was manifest as a 3.1-fold increase in EC_{50} (log shift in EC_{50} = 0.49 ± 0.055 ; $F=21.30$, $P<0.001$). Since muscarinic contractile function is absent in urinary bladder from M₂/M₃ double KO mice (Matsui et al. 2002), these data indicate that STZ treatment inhibits M₃ receptor-mediated contraction and that this response is restored by the M₂ receptor.

M₂ muscarinic receptor-mediated inhibition of forskolin-induced relaxation We investigated the effect of prior treatment with STZ on forskolin-induced inhibition of contraction to oxotremorine-M in denuded urinary bladder from wild-type and M₂ KO mice. Forskolin (5 μ M) had a small inhibitory effect on the concentration–response curve to oxotremorine-M in wild-type bladder from vehicle- and STZ-treated mice at all time points (Fig. 4). A much greater effect was observed in denuded bladder from M₂ KO mice, which is consistent with the role of the M₂ receptor in opposing adenylate cyclase-mediated inhibition of contraction.

We used the operational model (see “Methods” section) to estimate the relaxant effect of forskolin in urinary bladder from wild-type ($-\log \alpha$) and M₂ KO ($-\log \alpha/\beta$) mice. The difference between these two estimates ($-\log \beta$) is a measure of the function of the M₂ receptor in opposing forskolin-induced relaxation. The $-\log \beta$ values were estimated in urinary bladder from vehicle- and STZ-treated mice at the three time points, and the results are summarized in Fig. 5a. At the 8–9- and 20–24-week time points, STZ treatment caused a significant increase in the role of the M₂ receptor in opposing the relaxant effect of forskolin.

Effects of STZ on intact urinary bladder

Muscarinic agonist-induced contractions The presence of urothelium caused a modest change in the contractile activity of oxotremorine-M in the urinary bladder (Fig. 6). The overall average E_{\max} value of oxotremorine-M in intact, wild-type urinary bladder from all of the vehicle-treated groups ($201.9\pm8.9\%$, KCl-normalized contraction, Fig. 6b, e, h) was significantly smaller than the corresponding estimate for denuded urinary bladder ($243.4\pm11.6\%$, KCl-normalized contraction, Fig. 3b, e, h; $F=8.82$; $P=0.0033$). There was little or no difference in the contractile activity of oxotremorine-M in denuded bladder from vehicle-treated wild-type and M₂ KO mice at all time points (Fig. 3). In contrast, the overall average EC_{50} value of oxotremorine-M in intact, M₂ KO urinary bladder from all of the vehicle-treated groups ($pEC_{50}=6.38\pm0.13$) was

Fig. 4 Effect of streptozotocin (STZ) treatment on M_2 receptor-mediated inhibition of relaxation in urinary bladder lacking urothelium. The effect of forskolin (5 μ M) on oxotremorine-M-mediated contractile responses was measured in wild-type and M_2 KO bladders 2 to 3 weeks (a, b), 8 to 9 weeks (c–d), and 20 to 24 weeks (e–f) after STZ treatment. *Open circles* denote wild type, control; *closed circles* denote wild type in the presence of forskolin; *open triangles* denote M_2 KO, control; *closed triangles* denote M_2 KO in the presence of forskolin. Mean \pm SEM values from three to seven experiments are indicated, each done on a separate bladder

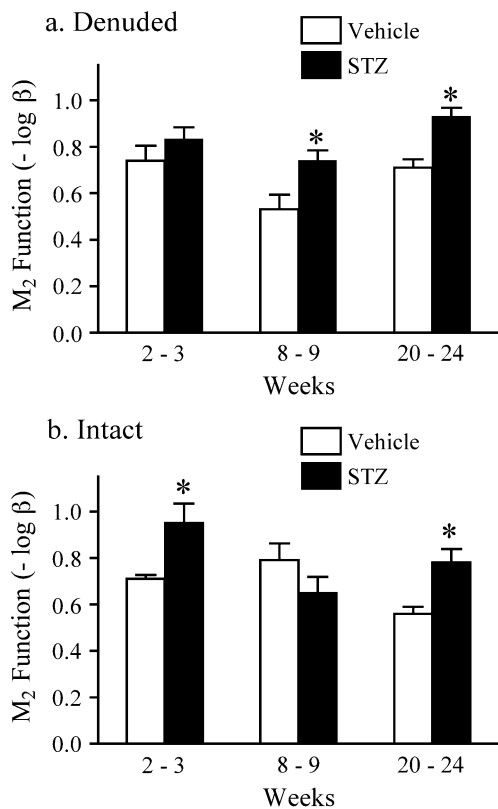
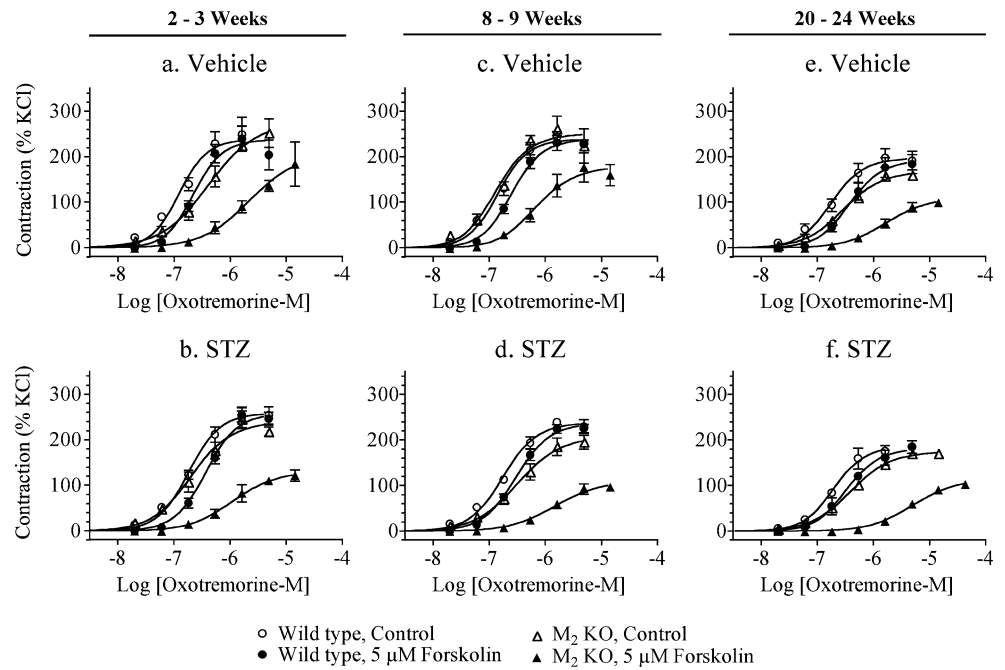


Fig. 5 Effect of streptozotocin treatment on the role of the M_2 receptor in opposing forskolin-induced relaxation in denuded (a) and intact (b) urinary bladder. The function of the M_2 receptor is quantified as the parameter $-\log \beta$, which is defined as the difference between the effect of forskolin in urinary bladders from wild-type ($-\log \alpha$) and M_2 KO ($-\log \alpha\beta$) mice as described under “Methods” section. The parameters were estimated from the data in Figs. 4 and 7. *Significantly different from vehicle, $P < 0.05$

significantly greater (lower potency) than the corresponding estimate for wild-type bladder ($pEC_{50} = 6.84 \pm 0.055$; Fig. 6; $F = 16.029$, $P < 0.001$).

Eight to 9 weeks after STZ treatment, the E_{max} value of oxotremorine-M, expressed in millinewton, was significantly greater by 51% in intact urinary bladder from STZ-treated wild-type mice compared to intact bladder from vehicle-treated wild-type animals (Fig. 6d and Table 3). When contraction was normalized relative to that elicited by KCl (50 mM; Fig. 6e), this difference persisted (Table 3). STZ treatment (8 to 9 weeks prior) had no effect on the contractile action of oxotremorine-M in intact urinary bladder from the M_2 KO mouse (Fig. 6f). Nonetheless, when the activity of oxotremorine-M in the intact M_2 KO bladder is compared to that of wild type, there is a greater reduction in E_{max} following STZ treatment ($F = 5.11$, $P = 0.026$).

These changes were not significant 2 to 3 weeks (Fig. 6a–c) or 20 to 24 weeks (Fig. 6g–i) after STZ treatment.

M_2 muscarinic receptor-mediated inhibition of forskolin-induced relaxation We also investigated M_2 receptor-mediated inhibition of relaxation in urinary bladder with intact urothelium (Fig. 7). As observed in denuded bladder, the relaxant effect of forskolin on oxotremorine-M-induced contractions was greater in intact urinary bladder from the M_2 KO mouse compared to that from wild type, illustrating the role of the M_2 receptor in opposing the relaxant effect of forskolin. This action of the M_2 receptor was significantly enhanced 2 to 3 weeks and 20 to 24 weeks after STZ treatment (Fig. 5b).

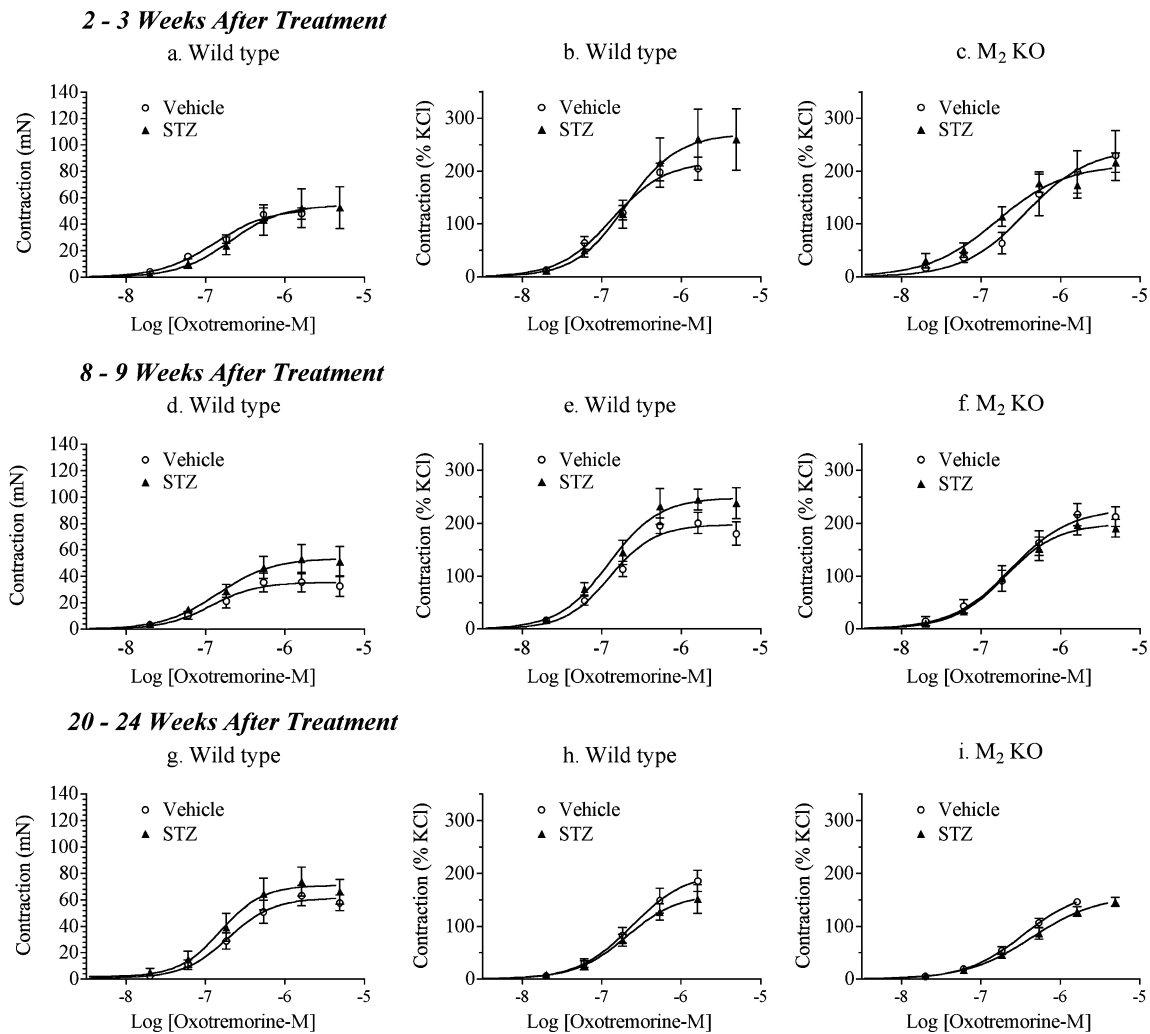


Fig. 6 Effect of streptozotocin (STZ) treatment on oxotremorine-M-mediated contractions in urinary bladder with intact urothelium from wild-type and M_2 KO mice. Concentration–response curves were measured 2 to 3 weeks (a–c), 8 to 9 weeks (d–f), and 20 to 24 weeks (g–i) after STZ treatment in urinary bladder from wild-type (a, b, d, e,

g, h) and M_2 KO (c, f, i) mice. Contractions are expressed in units of force (millinewton; a, d, g) or relative to the contraction elicited by KCl (50 mM; b, c, e, f, h, i). Circles denote the vehicle-treated group; triangles denote the STZ-treated group. Mean \pm SEM values from three to seven experiments are shown, each done on a separate bladder

Discussion and conclusions

In our study, we used a single injection of STZ (125 mg kg⁻¹) to induce diabetes. This dose was determined to yield maximal levels of hyperglycemia with minimal mortality (data not shown). High doses of STZ (≥ 200 mg kg⁻¹) promote nonspecific cytotoxicity and acute renal tubular damage (Tay et al. 2005). When moderate doses (100–125 mg kg⁻¹) are used on two consecutive days, nonspecific and acute renal cytotoxicity is markedly reduced, and mice develop renal injury biochemically and histologically similar (albeit less severe) to human nephropathy (Tesch and Nikolic-Paterson 2006; Tesch and Allen 2007), which can develop as a late diabetic complication after 10–20 years (Molitch et al. 2004). Our use of a single injection of STZ (125 mg kg⁻¹) probably caused even less STZ-induced

nonspecific and acute cytotoxicity. Also, it is unlikely that our mice suffered from diabetic ketoacidosis as urinalysis test strips showed zero, or very rarely, trace (5 mg dL⁻¹) amounts of acetoacetic acid in animals treated with STZ at all time points (data not shown). We found that M_3 KO mice were very sensitive to STZ, and none survived long enough for experiments. M_3 KO mice of a C57BL/6 genetic background exhibit impaired glucose tolerance (Gautam et al. 2006), which may contribute to their high sensitivity to STZ.

We observed that STZ inhibits M_2 KO but not wild-type bladder contraction 8 to 9 weeks after treatment (Fig. 3e, f), suggesting that the M_2 receptor rescues impaired M_3 receptor-mediated contraction in denuded diabetic bladder. Mimata et al. (1995) observed a 30% increase in carbachol-stimulated phosphoinositide hydro-

Table 3 E_{\max} and pEC_{50} values measured in intact bladders from mice 2–24 weeks after injection

Intact bladder	2–3 weeks			8–9 weeks			20–24 weeks		
	E_{\max} (mN)	E_{\max} (% KCl)	pEC_{50}	E_{\max} (mN)	E_{\max} (% KCl)	pEC_{50}	E_{\max} (mN)	E_{\max} (% KCl)	pEC_{50}
Vehicle	WT	218.50±4.28	218.50±18.04	6.87±0.099	35.53±3.96	197.20±10.90	6.90±0.080	201.20±26.81	6.62±0.14
	M_2 KO	–	242.20±40.89	6.44±0.22	–	227.20±18.14	6.63±0.11	168.80±15.63	6.47±0.096
STZ	WT	54.48±8.48	269.70±31.94	6.69±0.16	53.63±6.14 ^a	248.60±16.69 ^a	6.91±0.10	161.40±21.21	6.69±0.14
	M_2 KO	–	212.30±19.14	6.80±0.14	–	199.40±14.93	6.70±0.10	157.90±11.02	6.36±0.091

Data are means ± SE (number of animals)

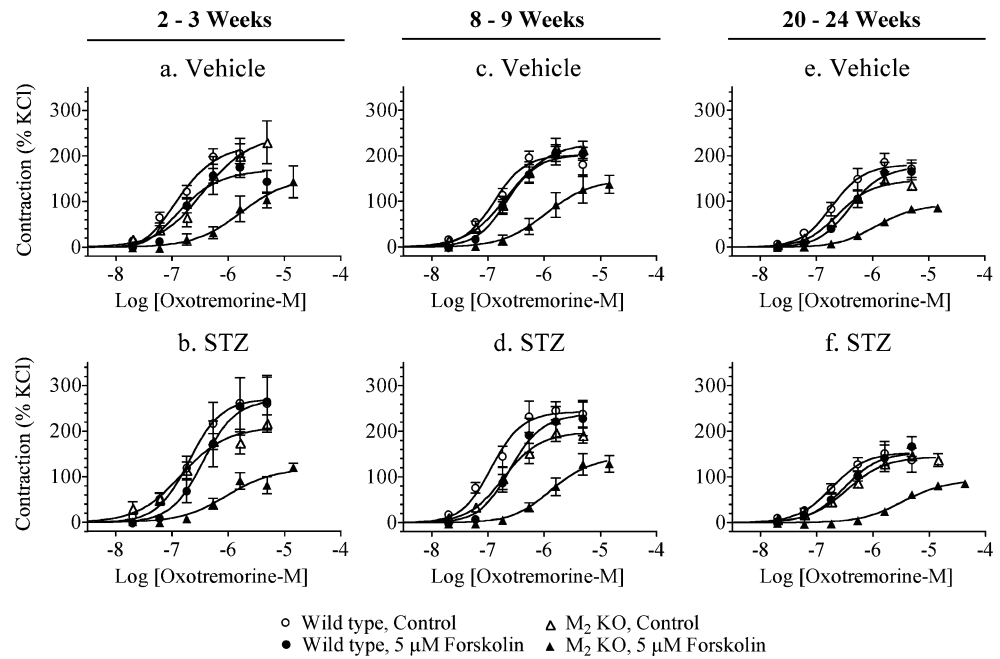
^a Statistically significant difference from wild-type vehicle ($P<0.01$)

lysis in [³H]inositol-labeled diabetic rat bladder when this response was expressed as the conversion of labeled [³H] phosphoinositides into [³H]inositol phosphates. Since the M_3 receptor is known to mediate phosphoinositide hydrolysis in mouse urinary bladder (Tran et al. 2006), the results of Mimata et al. (1995) suggest an increase in M_3 function in diabetic bladder. Perhaps this increase might contribute to the overall increase in muscarinic receptor-induced contractile force (millinewton) that we observed in wild-type mouse urinary bladder. Most of this increase in contraction can be attributed to the M_2 receptor, however, which does not signal through phosphoinositide hydrolysis in mouse urinary bladder (Tran et al. 2006). Also, the increase in muscarinic contractile force was absent when contractions were normalized relative to that elicited by KCl. Alternatively, there may be differences in the effects of STZ on urinary bladder from rats and mice.

Previous work from our lab has shown that the M_2 receptor enhances contractions elicited by the M_3 receptor in colon (Sawyer and Ehlert 1999) and urinary bladder (Ehlert et al. 2005) from normal guinea pig and mouse, respectively. This M_2 enhancement mechanism acts at a lower potency than that required for M_3 -mediated contractions, and oxotremorine-M elicits a maximal contraction through the M_3 receptor at a concentration that is too low to activate the M_2 enhancement function (Sawyer and Ehlert 1999; Ehlert et al. 2005). When a portion of the population of M_3 receptors is pharmacologically inactivated, oxotremorine-M is still able to elicit maximal contraction, albeit at a 20-fold lower potency. In these bladders, inhibition of M_2 receptor signaling by pertussis toxin (Sawyer and Ehlert 1999) or by gene deletion (Ehlert et al. 2005) induces a substantial loss of function in urinary bladder, revealing M_2 enhancement of contractions elicited by the M_3 receptor. Since the M_3 receptor can elicit a maximal contraction on its own, the normal physiological significance for the M_2 enhancement mechanism is unclear, but the observation in the current study that this mechanism rescues M_3 receptor-mediated contractions, which have been attenuated by STZ treatment, may suggest that the M_2 receptor has a crucial role in sustaining function in disease states such as diabetes.

A similar upregulation in M_2 receptor function occurs with denervation of the urinary bladder or obstruction of its outlet. In the rat model of these conditions, there is an upregulation of M_2 muscarinic receptor expression and mRNA, an increase in the potency of muscarinic agonists for eliciting contraction, and a change in the potency of antagonists for inhibiting contraction that is consistent with a greater role for the M_2 receptor (Braverman et al. 1998, 1999). This increase in M_2 function is caused by hypertrophy of the bladder because the increase in M_2 receptor

Fig. 7 Effect of streptozotocin (STZ)-treatment on M_2 receptor-mediated inhibition of relaxation in urinary bladder with intact urothelium. The effect of forskolin (5 μ M) on oxotremorine-M-mediated contractile responses was measured in wild-type and M_2 KO bladders 2 to 3 weeks (a, b), 8 to 9 weeks (c, d), and 20 to 24 weeks (e, f) after STZ treatment. Open circles denote wild type, control; closed circles denote wild type in the presence of forskolin; open triangles denote M_2 KO, control; closed triangles denote M_2 KO in the presence of forskolin. Mean \pm SEM values from three to seven experiments are indicated, each done on a separate bladder



expression is prevented in rats that have had their ureters sutured to the colon to divert the flow of urine to the GI tract and prevent the denervation-induced distension and hypertrophy of the bladder (Braverman and Ruggieri 2003). An analogous increase in M_2 function has been observed in human urinary bladder with reduced innervation as a result of spinal injury (Pontari et al. 2004; Ruggieri and Braverman 2006).

Another role of the M_2 receptor involves inhibition of cAMP-mediated relaxation in smooth muscle from mouse (Matsui et al. 2003; Ehlert et al. 2005, 2007), guinea pig (Noronha-Blob et al. 1989; Thomas et al. 1993), and rat (Hegde et al. 1997). In the current study, we investigated how STZ treatment affects this M_2 receptor-mediated disinhibition of contraction by comparing the degree of forskolin-induced inhibition of muscarinic contractions in M_2 KO versus wild-type bladders. Forskolin had a greater effect on the concentration–response curve to oxotremorine-M (i.e., rightward shift) in bladder from M_2 KO mice compared to that from wild-type animals, illustrating the ability of the M_2 receptor to oppose the relaxant effect of forskolin. This M_2 disinhibition of contraction was significantly greater at the intermediate (8–9 weeks) and long (20–24 weeks) time points after STZ treatment in denuded urinary bladders (Fig. 5a). These data are consistent with the observation that STZ treatment increases M_2 receptor expression (Tong et al. 1999, 2002a, b, 2006; Stevens et al. 2006) in bladder smooth muscle.

Urothelium is known to release an inhibitory factor upon muscarinic stimulation (Hawthorn et al. 2000; Kosan et al.

2005). Our results are consistent with this phenomenon, because we found that oxotremorine-M exhibited a significantly greater E_{max} value for contraction in denuded wild type compared to intact wild type urinary bladder. We also found that while there was no difference in the contractile activity of oxotremorine-M in denuded bladder from wild-type and M_2 KO mice, the agonist did exhibit 2.9-fold lower potency in intact M_2 KO relative to intact wild-type urinary bladder. Previously, we reported a similar difference in intact bladder (Matsui et al. 2003). These results might suggest that urothelial M_2 receptors mediate an inhibition of the release of the inhibitory factor, whose initial release is stimulated by a muscarinic subtype other than M_2 . The human urothelium expresses mRNA for all muscarinic subtypes, with M_2 mRNA greatly exceeding that of the other subtypes (Bschleipfer et al. 2007). Urothelial M_2 (Tong et al. 2006) and M_3 (Cheng et al. 2007) receptor expression is upregulated with STZ treatment. The urothelium is now known to be more involved in bladder function beyond its traditional role as a passive barrier to underlying smooth muscle (Birder and de Groat 2007).

The inhibitory effect of the urothelium on muscarinic contractility is abolished by STZ treatment in rats, suggesting a loss in release of the inhibitory factor (Kosan et al. 2005). We also confirmed that STZ treatment enhanced contractions in intact wild-type mouse urinary bladder (Fig. 6e), but had no effect on denuded wild-type bladder when contraction was normalized relative to KCl (Fig. 3e).

In denuded M_2 KO bladder (Fig. 3f), we found that STZ treatment inhibited muscarinic contractile potency, whereas in intact M_2 KO bladder (Fig. 6f), STZ treatment had no effect.

This seemingly conflicting result can be explained by the opposing effects of STZ treatment on muscarinic contractility in intact M₂ KO urinary bladder—an enhancement from the loss of urothelial inhibition and an inhibition from a loss of M₃ function. The loss of M₃ function was manifest in intact urinary bladder as a significant reduction in the E_{\max} of oxotremorine-M in M₂ KO relative to wild type 8 to 9 weeks after STZ treatment. This loss of function was significantly greater than that observed in intact bladder from vehicle-treated animals. Thus, our hypothesis that STZ treatment inhibits M₃ receptor contractile function in smooth muscle and that the smooth muscle M₂ receptor rescues contractile function is supported by our findings on both denuded and intact bladder 8 to 9 weeks after STZ treatment. This effect seems to be short-lived, however, because it is lost 20 to 24 weeks after STZ treatment.

Our experiments on the role of the M₂ receptor in opposing forskolin-induced relaxation in intact bladders showed an increase in this function following STZ treatment. We observed a significant increase in M₂ function at the early (2–3 weeks) and late (20–24 weeks) time points after STZ treatment, but not at the intermediate time point (8–9 weeks). In denuded bladder, significant differences were measured at the intermediate and late time points. We have no explanation for this difference, but experimental variation and changes in the effect of forskolin on the release of mediators from the urothelium after STZ treatment (Birder et al. 2002) may confound our results.

This temporal progression of differential changes in M₂ and M₃ contractile function in urinary bladder is likely caused by gradual STZ-induced alterations. Hyperglycemia-induced neuronal dysfunction and subsequent damage to the neuromuscular junction may alter contractile function in smooth muscle. Additionally, the rich innervation of the urothelium (Wakabayashi et al. 1995) and its high level of glucose metabolism may make urothelial tissue gradually susceptible to diabetic conditions. Our results provide a clearer understanding of the differential roles of M₂ and M₃ muscarinic receptors in STZ-induced diabetes and how the M₂ receptor maintains contractile function in this disease model.

Acknowledgements This study was supported by National Institutes of General Medical Sciences (NIGMS) RO1 GM069829, University of Tennessee, federal flow through funds from the National Heart, Lung and Blood Institute (NHLBI) UTN-37775, National Institute of Diabetes and Digestive and Kidney Diseases (NIDDK) Individual National Research Service Award Fellowship F30 DK081289, Achievement Rewards for College Scientists (ARCS) Foundation grant, and the UC Irvine Medical Scientist Training Program (MSTP). The content is solely the responsibility of the authors and does not necessarily represent the official views of the aforementioned funding sources. This work was also presented at the British Pharmacological Society Third Focused Meeting on Cell Signaling on April 21, 2009, at the University of Leicester, Leicester, UK.

Open Access This article is distributed under the terms of the Creative Commons Attribution Noncommercial License which permits any noncommercial use, distribution, and reproduction in any medium, provided the original author(s) and source are credited.

References

- Birder LA, de Groat WC (2007) Mechanisms of disease: involvement of the urothelium in bladder dysfunction. *Nat Clin Pract Urol* 4:46–54
- Birder LA, Nealen ML, Kiss S, de Groat WC, Caterina MJ, Wang E, Apodaca G, Kanai AJ (2002) Beta-adrenoceptor agonists stimulate endothelial nitric oxide synthase in rat urinary bladder urothelial cells. *J Neurosci* 22:8063–8070
- Black JW, Leff P (1983) Operational models of pharmacological agonism. *Proc R Soc Lond B Biol Sci* 220:141–162
- Bolger GT, Gengo P, Klockowski R, Luchowski E, Siegel H, Janis RA, Triggle AM, Triggle DJ (1983) Characterization of binding of the Ca⁺⁺ channel antagonist, [3H]nitrendipine, to guinea-pig ileal smooth muscle. *J Pharmacol Exp Ther* 225:291–309
- Bolton TB (1979) Mechanisms of action of transmitters and other substances on smooth muscle. *Physiol Rev* 59:606–718
- Braverman AS, Ruggieri MR Sr (2003) Hypertrophy changes the muscarinic receptor subtype mediating bladder contraction from M3 toward M2. *Am J Physiol Regul Integr Comp Physiol* 285: R701–R708
- Braverman AS, Luthin GR, Ruggieri MR (1998) M2 muscarinic receptor contributes to contraction of the denervated rat urinary bladder. *Am J Physiol* 275:R1654–R1660
- Braverman A, Legos J, Young W, Luthin G, Ruggieri M (1999) M2 receptors in genito-urinary smooth muscle pathology. *Life Sci* 64:429–436
- Bschleipfer T, Schukowski K, Weidner W, Grando SA, Schwantes U, Kummer W, Lips KS (2007) Expression and distribution of cholinergic receptors in the human urothelium. *Life Sci* 80:2303–2307
- Cheng J-T, Yu B-C, Tong Y-C (2007) Changes of M3-muscarinic receptor protein and mRNA expressions in the bladder urothelium and muscle layer of streptozotocin-induced diabetic rats. *Neurosci Lett* 423:1–5
- Cole WC, Sanders KM (1989) G proteins mediate suppression of Ca²⁺ +-activated K current by acetylcholine in smooth muscle cells. *Am J Physiol* 257:C596–C600
- de Groat WC, Yoshimura N (2001) Pharmacology of the lower urinary tract. *Annu Rev Pharmacol Toxicol* 41:691–721
- Ehlert FJ (2003a) Pharmacological analysis of the contractile role of M2 and M3 muscarinic receptors in smooth muscle. *Receptors Channels* 9:261–277
- Ehlert FJ (2003b) Contractile role of M2 and M3 muscarinic receptors in gastrointestinal, airway and urinary bladder smooth muscle. *Life Sci* 74:355–366
- Ehlert FJ, Griffin MT, Abe DM, Vo TH, Taketo MM, Manabe T, Matsui M (2005) The M2 muscarinic receptor mediates contraction through indirect mechanisms in mouse urinary bladder. *J Pharmacol Exp Ther* 313:368–378
- Ehlert FJ, Ahn S, Pak KJ, Park GJ, Sangnil MS, Tran JA, Matsui M (2007) Neuronally released acetylcholine acts on the m2 muscarinic receptor to oppose the relaxant effect of isoproterenol on cholinergic contractions in mouse urinary bladder. *J Pharmacol Exp Ther* 322:631–637
- Gautam D, Han S-J, Hamdan FF, Jeon J, Li B, Li JH, Cui Y, Mears D, Lu H, Deng C, Heard T, Wess J (2006) A critical role for [beta] cell M3 muscarinic acetylcholine receptors in regulating insulin

- release and blood glucose homeostasis in vivo. *Cell Metabolism* 3:449–461
- Griffin MT, Matsui M, Ostrom RS, Ehlert FJ (2009) The guinea pig ileum lacks the direct, high-potency, M(2)-muscarinic, contractile mechanism characteristic of the mouse ileum. *Naunyn Schmiedeberg Arch Pharmacol* 380:327–335
- Hawthorn MH, Chapple CR, Cock M, Chess-Williams R (2000) Urothelium-derived inhibitory factor(s) influences on detrusor muscle contractility in vitro. *Br J Pharmacol* 129:416–419
- Hegde SS, Eglen RM (1999) Muscarinic receptor subtypes modulating smooth muscle contractility in the urinary bladder. *Life Sci* 64:419–428
- Hegde SS, Choppin A, Bonhaus D, Briaud S, Loeb M, Moy TM, Loury D, Eglen RM (1997) Functional role of M2 and M3 muscarinic receptors in the urinary bladder of rats in vitro and in vivo. *Br J Pharmacol* 120:1409–1418
- Inoue R (1991) Ion channels involved in responses to muscarinic receptor activation in smooth muscle. In: Sperelakis HKN (ed) *Ion channels of vascular smooth muscle cells and endothelial cells*. Elsevier Science Publishing Co., Inc, New York, pp 81–91
- Inoue R, Isenberg G (1990) Acetylcholine activates nonselective cation channels in guinea pig ileum through a G protein. *Am J Physiol* 258:C1173–C1178
- Kitazawa T, Hiram R, Masunaga K, Nakamura T, Asakawa K, Cao J, Teraoka H, Unno T, Komori S, Yamada M, Wess J, Taneike T (2008) Muscarinic receptor subtypes involved in carbachol-induced contraction of mouse uterine smooth muscle. *Naunyn Schmiedeberg Arch Pharmacol* 377:503–513
- Kolta MG, Wallace LJ, Gerald MC (1985) Streptozocin-induced diabetes affects rat urinary bladder response to autonomic agents. *Diabetes* 34:917–921
- Kosan M, Hafez G, Ozturk B, Ozgunes O, Gur S, Cetinkaya M (2005) Effect of urothelium on bladder contractility in diabetic rats. *Int J Urol* 12:677–682
- Kume H, Mikawa K, Takagi K, Kotlikoff MI (1995) Role of G proteins and KCa channels in the muscarinic and beta-adrenergic regulation of airway smooth muscle. *Am J Physiol* 268:L221–L229
- Latifpour J, Gousse A, Kondo S, Morita T, Weiss RM (1989) Effects of experimental diabetes on biochemical and functional characteristics of bladder muscarinic receptors. *J Pharmacol Exp Ther* 248:81–88
- Lincoln J, Crockett M, Haven AJ, Burnstock G (1984) Rat bladder in the early stages of streptozotocin-induced diabetes: adrenergic and cholinergic innervation. *Diabetologia* 26:81–87
- Longhurst PA, Belis JA (1986) Abnormalities of rat bladder contractility in streptozotocin-induced diabetes mellitus. *J Pharmacol Exp Ther* 238:773–777
- Matsui M, Motomura D, Karasawa H, Fujikawa T, Jiang J, Komiya Y, Takahashi S, Taketo MM (2000) Multiple functional defects in peripheral autonomic organs in mice lacking muscarinic acetylcholine receptor gene for the M3 subtype. *Proc Natl Acad Sci USA* 97:9579–9584
- Matsui M, Motomura D, Fujikawa T, Jiang J, Takahashi S, Manabe T, Taketo MM (2002) Mice lacking M2 and M3 muscarinic acetylcholine receptors are devoid of cholinergic smooth muscle contractions but still viable. *J Neurosci* 22:10627–10632
- Matsui M, Griffin MT, Shehnaz D, Taketo MM, Ehlert FJ (2003) Increased relaxant action of forskolin and isoproterenol against muscarinic agonist-induced contractions in smooth muscle from M2 receptor knockout mice. *J Pharmacol Exp Ther* 305:106–113
- Michel MC, Barendrecht MM (2008) Physiological and pathological regulation of the autonomic control of urinary bladder contractility. *Pharmacol Ther* 117:297–312
- Mimata H, Wheeler MA, Fukumoto Y, Takigawa H, Nishimoto T, Weiss RM, Latifpour J (1995) Enhancement of muscarinic receptor-coupled phosphatidylinositol hydrolysis in diabetic bladder. *Mol Cell Biochem* 152:71–76
- Molitch ME, DeFronzo RA, Franz MJ, Keane WF, Mogensen CE, Parving HH, Steffes MW (2004) Nephropathy in diabetes. *Diabetes Care* 27(Suppl 1):S79–S83
- Motulsky H, Christopoulos A (2003) *Fitting models to biological data using linear and nonlinear regression*. GraphPad Software, Inc, San Diego
- Nakamura T, Kimura J, Yamaguchi O (2002) Muscarinic M2 receptors inhibit Ca²⁺-activated K⁺ channels in rat bladder smooth muscle. *Int J Urol* 9:689–696
- Noronha-Blob L, Lowe V, Patton A, Canning B, Costello D, Kinnier WJ (1989) Muscarinic receptors: relationships among phosphoinositide breakdown, adenylate cyclase inhibition, in vitro detrusor muscle contractions and in vivo cystometrograms studies in guinea pig bladder. *J Pharmacol Exp Ther* 249:843–851
- Nsabimana AM, Ojewole JA (2008) Diabetes mellitus and responses of the urinary bladder to acetylcholine: an in vitro study. *J Smooth Muscle Res* 44:143–150
- Ostrom RS, Ehlert FJ (1997) M2 muscarinic receptor inhibition of agonist-induced cyclic adenosine monophosphate accumulation and relaxation in the guinea pig ileum. *J Pharmacol Exp Ther* 280:189–199
- Ostrom RS, Ehlert FJ (1998) M2 muscarinic receptors inhibit forskolin-but not isoproterenol-mediated relaxation in bovine tracheal smooth muscle. *J Pharmacol Exp Ther* 286:234–242
- Pontari MA, Braverman AS, Ruggieri MR Sr (2004) The M2 muscarinic receptor mediates in vitro bladder contractions from patients with neurogenic bladder dysfunction. *Am J Physiol Regul Integr Comp Physiol* 286:R874–R880
- Ruggieri MR Sr, Braverman AS (2006) Regulation of bladder muscarinic receptor subtypes by experimental pathologies. *Auton Autacoid Pharmacol* 26:311–325
- Sakamoto T, Unno T, Kitazawa T, Taneike T, Yamada M, Wess J, Nishimura M, Komori S (2007) Three distinct muscarinic signalling pathways for cationic channel activation in mouse gut smooth muscle cells. *J Physiol* 582:41–61
- Sawyer GW, Ehlert FJ (1999) Muscarinic M3 receptor inactivation reveals a pertussis toxin-sensitive contractile response in the guinea pig colon: evidence for M2/M3 receptor interactions. *J Pharmacol Exp Ther* 289:464–476
- Stengel PW, Yamada M, Wess J, Cohen ML (2002) M(3)-receptor knockout mice: muscarinic receptor function in atria, stomach fundus, urinary bladder, and trachea. *Am J Physiol Regul Integr Comp Physiol* 282:R1443–R1449
- Stevens LA, Sellers DJ, McKay NG, Chapple CR, Chess-Williams R (2006) Muscarinic receptor function, density and G-protein coupling in the overactive diabetic rat bladder. *Auton Autacoid Pharmacol* 26:303–309
- Tay YC, Wang Y, Kairaitis L, Rangan GK, Zhang C, Harris DC (2005) Can murine diabetic nephropathy be separated from superimposed acute renal failure? *Kidney Int* 68:391–398
- Tesch GH, Allen TJ (2007) Rodent models of streptozotocin-induced diabetic nephropathy. *Nephrology (Carlton)* 12:261–266
- Tesch GH, Nikolic-Paterson DJ (2006) Recent insights into experimental mouse models of diabetic nephropathy. *Nephron Exp Nephrol* 104:e57–e62
- Thomas EA, Ehlert FJ (1994) Pertussis toxin blocks M2 muscarinic receptor-mediated effects on contraction and cyclic AMP in the guinea pig ileum, but not M3-mediated contractions and phosphoinositide hydrolysis. *J Pharmacol Exp Ther* 271:1042–1050
- Thomas EA, Baker SA, Ehlert FJ (1993) Functional role for the M2 muscarinic receptor in smooth muscle of guinea pig ileum. *Mol Pharmacol* 44:102–110
- Tong YC, Cheng JT (2002) Alteration of M(3) subtype muscarinic receptors in the diabetic rat urinary bladder. *Pharmacology* 64:148–151

- Tong YC, Chin WT, Cheng JT (1999) Alterations in urinary bladder M2-muscarinic receptor protein and mRNA in 2-week streptozotocin-induced diabetic rats. *Neurosci Lett* 277:173–176
- Tong YC, Cheng JT, Wan WC (2002a) Effects of Ba-Wei-Die-Huang-Wan on the cholinergic function and protein expression of M2 muscarinic receptor of the urinary bladder in diabetic rats. *Neurosci Lett* 330:21–24
- Tong YC, Chin WT, Cheng JT (2002b) Role of sorbitol in the up-regulation of urinary bladder M(2) muscarinic receptors in streptozotocin-induced diabetic rats. *Neurourol Urodyn* 21:154–159
- Tong Y-C, Cheng J-T, Hsu C-T (2006) Alterations of M2-muscarinic receptor protein and mRNA expression in the urothelium and muscle layer of the streptozotocin-induced diabetic rat urinary bladder. *Neurosci Lett* 406:216–221
- Tran JA, Matsui M, Ehlert FJ (2006) Differential coupling of muscarinic M1, M2, and M3 receptors to phosphoinositide hydrolysis in urinary bladder and longitudinal muscle of the ileum of the mouse. *J Pharmacol Exp Ther* 318:649–656
- Wade GR, Sims SM (1993) Muscarinic stimulation of tracheal smooth muscle cells activates large-conductance Ca(2+)-dependent K+ channel. *Am J Physiol* 265:C658–C665
- Wakabayashi Y, Kojima Y, Makiura Y, Tomoyoshi T, Maeda T (1995) Acetylcholinesterase-positive afferent axons in mucosa of urinary bladder of adult cats: retrograde tracing and degeneration studies. *Histol Histopathol* 10:523–530