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Carriers for the Tunable Release of Therapeutics: Etymological Classification and Examples

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Abstract

Introduction—Physiological processes at the molecular level take place at precise spatiotemporal scales, which vary from tissue to tissue and from one patient to another, implying the need for the carriers that enable tunable release of therapeutics.

Areas Covered—Classification of all drug release to intrinsic and extrinsic is proposed, followed by the etymological clarification of the term “tunable” and its distinction from the term “tailorable”. Tunability is defined as analogous to tuning a guitar string or a radio receiver to the right frequency using a single knob. It implies changing a structural parameter along a continuous quantitative scale and correlating it numerically with the release kinetics. Examples of tunable, tailorable and environmentally responsive carriers are given, along with the parameters used to achieve these levels of control.

Expert Opinion—Interdependence of multiple variables defining the carrier microstructure obstructs the attempts to elucidate parameters that allow for the independent tuning of release kinetics. Learning from the tunability of nanostructured materials and superstructured metamaterials can be a fruitful source of inspiration in the quest for the new generation of tunable release carriers. The greater intersection of traditional materials sciences and pharmacokinetic perspectives could foster the development of more sophisticated mechanisms for tunable release.

Keywords

Drug Delivery; Release Kinetics; Tailorable Release; Tunable Release

1. Introduction

The increasing incidence of chronic diseases coupled with the incentives to improve the quality of life has attributed to the development of advanced biomaterials[1][2][3]. Over the past several decades, metals, polymers, ceramics and bio-derived materials have been extensively studied and used as medical implants. Such materials can be designed for a wide
range of biomedical applications, e.g., controlled drug delivery devices, tissue engineering constructs, vascular grafts and multiple other therapeutic, regenerative and/or diagnostic platforms. However, the ability of materials scientists to fine-tune the structure of materials to desired mechanical or electromagnetic properties has surpassed the ability to fine-tune them to exhibit complex kinetics of release of therapeutic molecules.

Overall, there is a need for the development of ever more sensitive and smarter controlled release carriers in pharmaceutics. To terminate the drug release when the therapeutic goal has been achieved and to trigger the release from the carrier only in the presence of a certain disease marker are only some of the demands for the new generation of drug release carriers. At a finer level, the drug release kinetic profiles could be designed for precise temporal matching of the targeted physiological processes that facilitate healing on the molecular scale. This would prevent systemic or local toxicity, ensuring patient safety while also increasing the efficacy with which the device delivers its therapeutic payload.

Many biological processes take place at precise timescales. For example, the reaction of the complement system to a foreign entity involves a cascade of processes, each of which is preceded and antecedent by others at precise timescales. Sophisticated targeting of individual processes in such a cascade necessitates the initiation of the therapeutic action through the controlled release of the right molecular agent within a very specific time window. Another example comes from wound healing (Fig. 1a), a physical response to injury that similarly involves a multitude of processes, each of which occur within precise time windows. Interfering with these processes with the intended therapeutic effect means that the timescale of this interference must be precisely set to the right time windows. Bone remodeling is yet another biological process divisible to individual stages (Fig. 1b), implying that an ideal therapeutic interference with this process would also require a sequential, precisely temporally tuned delivery of the right therapeutic agents. Targeted drug delivery in terms of precise 3D location of the released drug would be greatly enriched if one such targeting effect in the fourth, temporal dimension is achieved too.

Types of compounds traditionally delivered with the use of sustained release platforms since the early days of controlled drug delivery devices are listed in Table 1. However, preclinical optimizations of ideal release kinetics to individual pathological features on the molecular scale have ceded place to “one size fits all” approach, where a single type of implant, microstructurally speaking, is being supplied to the clinic, ignoring the fact that every clinical case is unique and different from any other. Additionally, utilizing the therapeutic role of small biomolecules and proteins on tissue regeneration may demand their controlled release at precisely tunable timescales. Signaling molecules, including hormones, cytokines and growth factors are included on this long list of compounds whose finely tuned delivery from the kinetic standpoint would greatly facilitate the wound healing and tissue regeneration processes.

The need for tunable release profiles is illustratable by the dichotomy between cortical and cancellous bone. Namely, with the cortical bone being more compact and having a lower remodeling rate than the more porous and vascularized cancellous bone, the ideal release profiles should be faster and more intense in the treatment of cancellous bone. As one shifts
from the surface of long bones to their marrow, one simultaneously moves in the direction of decreasing compactness and an increasing soft tissue nature. The clearance rate of the released drug would consequently decrease with the distance from the central axis of long bones; the need for the tuning of the release rate to the right implant location naturally follows from acknowledging this functionally gradient nature of bone. Sustained delivery of pain medication from sutures and wound dressings has, likewise, demonstrated that no single release profile fits all patients. Rather, in some cases the medication has to be supplemented orally because of the insufficient dose delivered, whereas in other cases the oral administration of opioid antagonists is necessary because of the adverse effects of the systemic overdose[19]. These clinical circumstances imply that tunable release agents are necessary to match the wide variety of analgesic needs of patients.

Therefore, the variability of structure and properties of a single tissue depending on the location as well as the variability of structure and properties of identical tissues in different patients imply the demand for carriers whose release rates could be tuned by varying a single parameter that defines the corresponding internal structure of the material across a continuous range of values. In such a manner, carriers would be designed to deliver drugs to tissues in a tunable fashion and accommodate directly to the ideal projected drug delivery profiles for the given tissues and the patient. In theory, rapid prototyping of implants in direct feedback with computerized axial tomography scans could thus be complemented with setting of the properties of the implant material to achieve the exactly desired release profile without resorting to qualitative modifications of the material. This qualitative constancy, notwithstanding the kinetic versatility, would also minimize the number of regulatory hurdles standing on the translational path. The applicative benefits of devising tunable drug carriers are thus clearly numerous enough to justify the furthering of the research on them.

2.1. Elementary classification: intrinsic vs. extrinsic control of the release

Kinetically controlled and tunable release of therapeutics from a carrier is attainable by multiple means. Still, all of them could be roughly divided to two categories: intrinsic and extrinsic. As shown in Fig. 2, in case of the intrinsic controlled release, the structure of the carrier becomes modified to vary the release rates and amounts, whereas in case of the extrinsic controlled release, either the environmental factors or the external, manual or automatized control triggers the release and occasionally enables the tuning of its rate to the desired kinetics. In what follows we will briefly mention the examples that belong to both, albeit for the most part limiting the scope of this review to the intrinsically tunable carriers.

2.2. Etymological clarification: tailorable vs. tunable

Confusion exists in the literature as to what the term “tunable” exactly means. Most occurrences of this attribute are incorrect, given that they describe the mere ability of a carrier to produce two or more distinct release profiles following the tweaking of its structure or introducing a variation in the physicochemical environment surrounding it. However, tunability, in the truer sense of the word, should be analogous to tuning a radio receiver along a continuous range of frequencies to a desired radio station or tuning a guitar string to the desired pitch. In other words, it should imply the correspondence between a
continuous range of values of a specific structural parameter of the carrier and a similarly continuous range of values of properties determining the release of the drug from the given carrier.

For the sake of convention, we will ascribe the attribute “tailorable” to the process of controlling the release kinetics by means of introducing qualitative changes in the carrier. We will ascribe the attribute “tunable” to the process of controlling the release kinetics by varying the quantitative value of a parameter that defines the structure of the carrier on a continuous scale. For example, if the size of particles loaded with a drug is reduced and such a reduction entails an increased rate of release of the physisorbed drug as the result of increased surface-to-volume ratio, one such effect could receive the attribute of “tunable”, provided that the control over the particle size is possible within a more or less continuous range and that a numerical correlation could be established between the particle size and the release rate. Likewise, when the stirring rate is varied to tune the nanoparticle size of coordination-metal polymeric particles within a wide range of values[20], such an effect, albeit somewhat trivial, is tunable so long as this range is continuous and broad enough.

Changing the release profile by changing the chemical identity of the ligand, on the other hand, does not constitute a tunable effect, even though it is frequently reported in the literature as such. Since tailoring denotes the act of cutting and pasting different patches of clothes, controlling the release by the presence or absence of certain particle components can be seen as analogous to the act of tailoring.

Fig. 2 schematically depicts this difference between tailoring and tuning. In the case of tunable release, a particular compositional or structural parameter is tuned to give a desired release profile, whereas in the case of tailorable release, adding or subtracting different carrier components, analogous to copying and pasting different patches of clothes, is used to modify the release profile. For example, the yielding of distinct release profiles upon the qualitative modification of the phase composition of calcium phosphate nanoparticles as carriers could be considered tailorable (Fig. 3a) [21]. In contrast, the tuning of the release of vancomycin and ciprofloxacin to anywhere between zero hours and two weeks by changing the weight ratio between two components of a hydroxyapatite cement, differing only in the rate of transformation into apatite from the amorphous primary precipitate, presents an authentic instance of the use of calcium phosphate nanoparticles as tunable release carriers (Fig. 5a–b) [22].

Another class of carriers which are often inappropriately named “tunable” are those whose release kinetics takes different forms depending on a specific physicochemical parameter that defines the conditions of their environment. Unlike the authentically tunable release carriers, in which tunability originates from the intrinsic control over a specific structural, compositional or synthesis parameter, in the case of such “smart” carriers, which release their payload only upon receiving a specific environmental stimuli in the form of a disease marker, the release kinetics is controlled extrinsically, as it is being caused by the indiscrete variation of an environmental parameter. One could include in this category carriers whose release is controlled by external effects, such as heat, light, magnetic field, ultrasound, mechanical force, and electric current (Fig. 2).


2.3. Examples of tailorable carriers

A study in which various material/drug combinations for wound healing were tested came to conclusion that incorporation of diclofenac into four fibrous wound dressing materials, namely alginate, sodium carboxymethyl cellulose (Na-CMC), viscose rayon and poly(ethylene terephthalate) (PET) renders kinetically distinct release profiles that could be tailored for both chronic and acute wound treatments [23]. While alginate and Na-CMC exhibited prolonged release and were found to be applicable for the treatment of chronic wounds, viscose rayon and PET were suitable for acute wounds. Next, Fig. 3b illustrates a difference in the release of vancomycin depending on the qualitative composition of cements composed of poly(D, L-lactide-co-glycolide) (PLGA) and calcium phosphate nanoparticles. While sole calcium phosphate caused the drug to be rapidly released, the addition of mannitol or PLGA, be it as empty or loaded microspheres, extended it to various degrees. Then, the release of fusion protein SH3-IGF-1 can be tailored by binding different peptides to a hydrogel composed of hyaluronan and methylcellulose (HAMC) as the carrier. As can be seen in Fig. 3c, the protein release is reduced in the presence of a strong binding peptide (SBP) as compared to unbound HAMC and HAMC with a weak binding peptide (WBP). Another example of tailorable release comes from the threefold acceleration of the release of diltiazem hydrochloride from chitosan nanoparticles following the conjugation of chitosan with L-leucine[24]. Adjusting the release of doxorubicin or vincristine to a desired rate by modifying silk films as the carrier with carboxyl or sulfonate groups also belongs to the category of tailored release (Fig. 3d).

2.4. Examples of devices with environmentally sensitive, stimuli-responsive release

2.4.1. Carriers stimulated by the chemical conditions of their microenvironment—Carriers capable of releasing their therapeutic payload only in the presence of a particular disease marker present the first step toward the development of theranostic drug delivery platforms. One example of “smart”, environmentally triggered release is shown in Fig. 4a–b: the release rate of the anti-inflammatory drug, piroxicam (PX), and of the negatively charged fluorescent sodium salt (FL) from polypyrrole (PPy) nanoparticles is directly proportional to pH (Fig. 4a), whereas that of the positively charged rhodamine 6G (R6G) is inversely proportional to pH (Fig. 4b). In Fig. 4c, gentamicin release is shown to be triggered in the event of infection and the associated increase in the acidity of the environment around the carrier. Such systems are attractive because of their potential to obliterate the need for the prophylactic use of antibiotics and at the same time minimize the systemic side effects. Also, the intracellular environment has a markedly higher redox potential than the extracellular milieu, with the concentration of glutathionate inside the cell (~ 10 mM) being three orders of magnitude higher than that outside the cell (~ 10 μM). The cytosolic cleavage of disulfide or thioester bonds can be used as a trigger for the controlled release of therapeutics, and one example is shown in Fig. 4d: as the concentration of dithiothreitol, which cleaves the disulfide bond, is increased, so does the rate of release of SN-38 from 2,6-bis(hydroxymethyl)aniline based polymeric nanoparticles increase too. The concentration of an enzyme in the environment surrounding the carrier is another parameter that could control the drug release rate: an example comes from the concentration of β-D-galactosidase being directly proportional to the rate of release of nitric oxide from the
enzymatically cleaved O₂-(β-D-galactopyranosyl)1-(N-isopropylamino)diazen-1-ium-1,2-diolate (Gal-IPA/NO) prodrug[28]

2.4.2. Carriers stimulated by the external physical signal—A special class of materials capable of having the release triggered by a change in a specific environmental parameter are those in which such a change is being implemented not spontaneously, by a biological process, but by the clinical operator. In such a case, the material responds to an optical, magnetic, thermal or acoustic signal from an external device. PPy scaffolds, for example, can exhibit tunable release by controlling the magnitude of electrical stimulation applied to the polymer, which causes an increase in the interplanar spacing in the inverse opal structure of PPy and the release of the intercalated drug [32]. Calcium alginate hydrogels also enable the tunability of the drug release by controlling the electric field strength. In one study, the release of folic acid increased from 13 to 18 to 25 to 31 % after the first 30 min of immersion as the electric field strength increased from 0 to 0.5 to 1 to 5 V[33]. The release of Texas-red-conjugated bovine serum albumin (BSA) from gelatinous microspheres incorporating thermoresponsive poly(N-isopropylacrylamide-co-acrylamide) (PNIPAm-co-AAm) can be tuned by controlling the degree of deswelling, which itself is controllable by temperature: from 20 to 100 % after 30 min as the temperature increased from 22 to 42 °C[34]. The drug release with thermoresponsive polymers need not always increase with temperature, as exemplified by the release of fluorescent 1-anilinonaphthalene-8-sulfonic acid from polystyrene-block-poly(N-isopropylacrylamide)-block-polystyrene (PS-b-PNIPAm-bPS) nanoparticles decreasing from 10 – 15 % after 3.6 h at 25 °C to 4 – 8 % after the same time at 45 °C, i.e., above the cloud point temperature of the PNIPAm[35].

In a nanomachine domain, the concentration of Nile blue was made precisely tunable by varying the rotation speed of nanoparticles in a nanomotor composed of a plasmonic, tri-layered Au/Ni/Au nanorod as a rotor, a 200 nm thick magnetic film of Au/Ni/Cr as the bearing and microelectrodes as the stator [36]. Specifically, the release rate of Nile blue bound to the surface of plasmonic nanorods by incubation in a solution monotonically increased with the rotation speed of the nanorods. This speed was controllable using the electric voltage applied to the quadruple microelectrodes in the device.

Then, a continuous increase in the magnetic field intensity in the 0 – 250 mT range linearly increased the amount of released docetaxel from a battery-less MEMS device composed of a drug reservoir wrapped up in a magnetic membrane made up of iron oxide nanoparticles dispersed in a polydimethylsiloxane (PDMS) matrix[37]. Ultrasound irradiation has not been controlled in intensity so as to produce tunable release profiles, but the use of on/off modulation of the field to control dosing was noted in the literature: for example, the release of rhodamine B from polystyrene-PDMS block co-polymeric nanofibers would be halted without the field and induced at a constant rate when the field is switched on[38]. Similar on/off modulations of photothermal, continuous-wave near-infrared laser irradiation allowed a precise control of the amount of released doxorubicin from mesoporous copper sulfide nanoparticles capped with hyaluronic acid to be achieved[39].
2.5. Examples of intrinsically tunable carriers

A limited number of intrinsic structural parameters has been used to control drug release rates in a tunable fashion. For example, porosity can be tuned to control the drug release; one example comes from a study in which an increase in the pore size from 15 to 43 to 95 nm in an oxidized porous silicon carrier gradually increased the release rate of daunorubicin (Fig. 5c) [40]. Similarly, the increase of the pore size of hollow silicon microspheres from 15 to 25 to 50 nm entailed a directly proportional increase in the release rate of quantum dots from these microspheres [41]. Control over the current density and anodization time in the 40 – 60 mA/cm$^2$ and 5 – 10 s range allowed for tuning the porosity of mesoporous silicon from 53 to 60 to 72 % and the corresponding release of α-chymotrypsinogen from 3 to 45 to 85 % after 24 h [42].

The crosslinking ratio in a polymer, typically controlled with the molar ratio between the monomer and the cross-linker in the synthesis stage, has been frequently used to control the diffusion coefficient of the drug entrapped within the polymer matrix and the rate of its release into the environment. As for the tunable effect, increasing the crosslinking ratio of calcium alginate in the 0.3 – 1.3 range proportionally decreased the amount of released folic acid [33]. Then, increasing the crosslinking ratio of gelatin in starch/gelatin microspheres from 0 to 12.5 to 25 % using glutaraldehyde decreased the released amount of methylene blue from 99 to 70 to 68 % after 24 h [43]. Also, controlling the concentration of the cross-linker, 3,3’-ethyldiene-bis(N-vinyl-2-pyrrolidone) (EBVP), in the 0.15 – 0.20 % range allowed the release of niacin to be made tunable to anywhere between 5 and 20 h [44]. Interestingly, no changes in crosslinking outside this range had an effect on the drug release, illustrating the limitations of this type of drug release control. In fact, it appears that crosslinking degree ranges in which the tunable release effect is achievable are often narrow and different depending on whether the release is short- or long-term. For example, increasing the crosslinking degree in gelatin scaffolds from 0 to 23.7 % had no long-term (7 – 21 days) effect on the release of mitomycin C, even though it did have a threefold effect on the decrease of the released amount after 24 h [45]. Finally, although the crosslinking degree usually inversely relates to the water ingestion and the drug diffusion and release rates, opposite effects are frequently found in the literature. Thus, because the concentration of active norbornene and thiol sites that bind the drug dropped with crosslinking, an increase in the molecular weight of a poly(ethylene glycol) (PEG) hydrogel led to an increase in the release rate of BSA: 40 and 60 % after 7 days for 2.5 and 4 kDa, respectively [46]. The same effect explains why an increase in the crosslinking density of both chain- and step-grown PEG caused a monotonous increase in the rate of release of BSA [47].

Crosslinking can be the key parameter for tuning the release rate of drugs not only from hydrogels, but from vitreous and elastomeric polymers too. For example, an increased acrylation of trimethylene carbonate (TMC) elastomer and the corresponding decrease in the ratio between TMC and D,L-lactide (DLLA) in their copolymer from 100:0 to 50:50, indicating a more efficient crosslinking reaction, gradually increased the release of BSA from 0 to 13 to 20 to 35 % after 40 days of the release time [48]. In this case, the fundamental understanding of the process is technically easier because the rate-limiting step is usually diffusion and occasionally osmosis, given that the effects of polymer swelling and...
erosion have a lesser effect on the release kinetics than in hydrogels. Diffusion of the solute through the polymeric matrix is, of course, dependent on a plenty of other parameters, many of which affect or are indirectly affected by crosslinking, including porosity, tortuosity, pore connectivity, the hydrodynamic radius of the solute, hydrophobicity and polarity of the polymer relative to the solute, the potential for hydrogen or van de Waals bonding between the solute and the polymer, solute concentration gradient as the driving force for diffusion, distribution of the repeat units in a polymer, geometry of the polymeric implant, crystallinity, end group character and identity, chain defects and rigidity, et cetera; however, not all of them are readily tunable in a sense in which this term is being used in this study.

The characteristic U-shaped degradation profile of PLGA as a function of LA/GA ratio\[49\] has been frequently used to tune the release rate of drugs entrapped in PLGA as a function of the weight ratio of these two monomers \[50\]. The release of thyrotropin releasing hormone from PLGA microspheres increased with the increase in the GA content of the polymer in the LA/GA ratio range from 50/50 to 100/0 \[27\]. Varying the LA/GA ratio from 70/30 to 90/10 while keeping other parameters constant decreased mitomycin-C release from 70 to 30 wt.% after three months of release \[52\]. Likewise, lowering the feed ratio between acrylamide (AAm) and (r)-α-acryloyloxy-β,β-dimethyl-γ-butyrolactone (DBA) in a thermoresponsive copolymer decreased the rate of release of dexamethasone from 100 % after a week for AAm/DBA = 5.25 to 15 % for AAm/DBA = 0.9\[53\]. The drug release from electrospun polymeric fibers composed of polycaprolactone (PCL) and PLGA was also made tunable by controlling the weight ratio between the two polymeric components \[54\]. Specifically, anti-HIV drug Tenofovir disoproxil was released faster at larger PCL contents and more sustainably at larger PLGA contents. Also, micelles made of the block copolymer composed of poly(ethylene glycol)-b-poly(lactic acid) (PEG-b-PLA) and PLA-b-PNIPAm displayed a reduced release of ibuprofen as the content of PLA-b-PNIPAm increased \[55\].

Tuning the release as a function of the weight ratio between the two components of a carrier is often the consequence of their different affinity for the drug, and one example comes from the tunability of the release of recombinant human insulin-like growth factor 1 (rHIGF-1): with rHIGF-1 binding more intensely to more biodegradable keratine (KTN) than to less biodegradable keratose (KOS), increasing the KTN/KOS weight ratio in a polymer increased the rate of release of rHIGF-1\[56\]. The release was tunable within the 10 – 70 % range after 7 days, as defined by the release from pure KTN (10 %) and pure KOS (70 %), respectively.

Still, although polymeric microspheres have been used as drug delivery carriers since the 1980s \[58\], the effects other than variations in the bulk composition and crosslinking ratio have been rarely used for tunable release purposes. Controlling the thickness of the polymeric film deposited over a drug reservoir can produce finely tuned release profiles\[59\]; however, this concept has been rarely ever translated to polymeric particle systems. Yet, there are notable examples. For example, hollow composite gold/polymeric microcapsules were 3D printed and the variation in the weight ratio of PLGA in the precursor solution from 0.5 to 5 wt.% controlled the capsule thickness and allowed the release of horseradish peroxidase to be tuned to profiles shown in Fig. 5d. Also, hybrid PLGA/TiO\(_2\) microspheres were fabricated using droplet-based microfluidics \[60\] and their surface texture was varied by changing the mass ratio between titanium tetraisopropoxide (TTIP) and PLGA in the dispersed phase. The higher the content of TTIP, the deeper the wrinkles and the faster the
drug release, a phenomenon which was attributed to a larger specific surface area leading to a more rapid release of Tanshinone IIA as the drug. The degree of oxidation of carboxymethyl cellulose microspheres with periodate was used to increase the release of doxorubicin from 15 to 20 to 25 % as the oxidation degree increased from 9 to 17 to 35 %, respectively[61]. The use of multiple other, finer polymer microparticle properties, - e.g., surface texture and chemistry, core/shell size/thickness, self-assembly features, etc. - is expected to open up a whole new avenue in controlled drug release research.

Other effects are also noted in the literature. Changing the degree of substitution of deoxycholic acid in nanoparticles based on hyaluronic and deoxycholic acid conjugates (HA-DOCA) from 5.9 to 7.6 to 9.4 changed the released amount of doxorubicin from 49 to 58 to 70 %[62]. Changing the particle size of magnesium carbonate from 25 – 50 to 75 – 100 to > 200 μm on average decreased the amount of released ibuprofen after 24 h, albeit within a very narrow range, from 98 to 93 to 87 % [63]. The effect of the drug on the structure of the carrier is never to be underestimated either, as the result of which the concentration of the drug loaded into the carrier can often be used as a parameter to produce tunable release profiles. One example comes from the increase in the released percentage of the loaded dexamethasone paralleling an increase in the amount of the drug present in PCL/silk fibroin nanoparticles as the carrier[64]. The release rate of doxorubicin from liposomes similarly increased in parallel with the increase in the concentration of the encapsulated drug and was made tunable using drug-to-lipid ratio as the control parameter[65]. The opposite effect was observed in the case of loading HA-DOCA nanoparticles with different amounts of doxorubicin: as the content of the drug in the carrier increased, the cumulative rate of the release of the drug decreased (71 vs. 42 vs. 32 % after 24 h for 5, 10 and 20 wt.% loadings, respectively)[62]. Such a disparity between the effects of technically identical parameters illustrates the complexity of understanding and controlling the mechanisms of drug release in general. Tunability achievable in a system with a specific drug/carrier combination may be obliterated when the identity of the drug changes. One example comes from the aforementioned tunable release of rhIGF-1 from KOS/KTN polymer, impossible to repeat with another growth factor, recombinant human bone morphogenetic protein 2, or with an antibiotic, ciprofloxacin[56]. Finally, Table 2 lists some of the patented carriers with intrinsically tunable release properties. Table 3 provides a summary of the carriers referred to in the text and of their categorization based on two major groups - intrinsic vs. extrinsic - and two subgroups: tunable vs. tailorable and smart vs. manual, respectively (Fig. 2).

3. Expert Opinion

A central problem in the elucidation of parameters that could be used as “magic knobs” and allow the drug release to be made intrinsically tunable is the inevitable entanglement of variables defining the microstructure of the carrier. Changing one of these parameters is bound to have an effect on other parameters, which may affect the release in an uncontrolled manner and deter the tunable effect. For example, increasing the size of sucrose acetate isobutyrate microsphere depots at first decreased the release rate of risperidone because of a lesser particle surface area conducive to drug diffusion, but then increased it at even larger particle sizes because of the larger porosity of the large particles[77]. Increasing the particle size also does not only lower the specific surface area of the material, but also lowers the
particle curvature and, thus, the surface energy, affecting particle-particle interaction, surface binding and release of the drug, and the diffusion of the drug across the particle/solution interface. Thus, although increased surface area of a flat surface directly translates to increased released rate[78], increased surface area in a particulate system comes at the cost of increased surface tension and compaction of the crystalline order at the particle surface, which can act as a barrier to the release. Still, the medical need for such carriers is obvious and its fueling the drive for the discovery is expected to overcome these fundamental challenges and ambiguities. The ease, the elegance and the practical appeal of using a single parameter to control the release rate as opposed to compromising between a multitude thereof will also be a crucial contributor to the development of a new generation of intrinsically tunable release carriers.

Much has been said about the necessity for the input of materials scientists on devising more advanced drug delivery platforms. However, the ability to control and theoretically predict physicochemical properties of materials in a tunable manner still greatly surpasses the ability to control and model their drug release properties. Insufficient versatility and inferior stimuli-response sophistication of drug delivery devices compared to those of electronic devices, the majority of which owe their function to the tunability of one or more of their components, speaks in favor of this disparity. For example, today we know that nanomaterials exhibit a markedly broader variety of potential properties than their bulk counterparts, and tunability is one of their hallmarks. Owing to the quantum confinement effect, the size of semiconducting particles at the nanoscale can be controlled to tune their bandgap and, thus, the photon emission frequency to the desired value[79]. Simple variations of graphene nanoribbon width, density or the doping level also allow for the tunability of the bandgap and the corresponding metal–oxide–semiconductor field-effect transistor properties[80][81][82]. Albeit having an insignificant effect in many other 2D materials, including MoS$_2$, a precise control over the number of deposited monoatomic layers of phosphorene gives a precise control over the magnitude of the transport gap of the material[83]. Plasmon resonance in gold nanostructures could be tuned by varying the particle diameter[84], aspect ratio[85], thickness in a layer[86], et cetera. Sophisticated superstructures, such as photonic crystals and metamaterials, also allow for the tunability of optical properties through a control over their geometric features and translational symmetry[87][88]. An unexplored, albeit bold question is if the tunability that typifies the quantum properties of nanoparticles could be translated to the tunability of drug release profiles.

Magnetic ceramics are instructive in terms of the intrinsic tunability of properties. By controlling the distribution of cations at octahedral and tetrahedral sites of the spinel lattice of ferrites, more than one of their magnetic properties, ranging from susceptibility to permeability to Neel temperature to coercivity to remanence, can be tuned[89]. Variation of the stoichiometric ratio between a couple of metallic dopants at the A site of the ABO$_3$ perovskite structure of nanostructured manganites, e.g., $x$ in La$_{1-x}$Sr$_x$MnO$_{3+y}$, can be employed to vary their Curie temperature and adjust it to the hyperthermically or thermoblastically targeted tissue in magnetic cancer therapies[90]. Controlling a similar stoichiometric parameter $x$, though defining Co/Ni molar ratio in magneto-dielectric ferrite, BaSrCo$_{2-x}$Ni$_x$Fe$_{12}$O$_{22}$, allows for the simultaneous tuning of the narrowband reflection
reduction in a wide microwave frequency range at low $x$ and high Co contents and of the wideband transmittance attenuation in a similarly wide frequency range at high $x$ and high Ni contents[91]. Atomic layer engineering has also enabled the tuning of an array of properties and transitions in alloys and ceramics depending on the atomically precise doping levels and deposited geometries, ranging from 1D ribbons and strings to nanoladders to sheets and films with an atomically defined thickness[92][93]. The question is whether similar materials, methods and structural effects could be employed to yield tunable release profiles too.

This question may be the starting point for the expansion of the repertoire of materials with tunable release properties. Traditionally, polymers have been the main choice for tunable release agents because of the ease with which this control be intrinsically achieved using parameters such as monomeric unit ratio, crosslinking ratio, etc. Until recently, it was thought that calcium phosphates, like most ceramics, did not have the potential for controllable release, let alone tunable. However, this was refuted with the development of self-setting pastes in which the release of antibiotics could be modeled with simplistic equations allowing for the prediction of release properties depending on a single, easily controllable compositional parameter[22]. As far as the externally triggered release platforms are concerned, piezoelectric and magnetostrictive materials, deforming in direct proportionality with the intensity of the applied electric[94] or magnetic[95] fields, respectively, and potentially releasing the captured drug in a kinetically controllable fashion, present another class of ceramics that could be investigated for tunable delivery. Nanotubes, e.g. boron nitride[96], or composites with polymers[97] present particularly attractive subclasses of such materials owing to their proven drug loading and release capabilities. Electrically responsive polymers[98] will also offer a whole new avenue in the utilization of the proposed effects. Therefore, it is important that research aiming to discover new materials for controlled release continues. Along the way, therapeutic synergies emerging from the carrier properties other than those enabling the tunable release are expected to be derivable by serendipity, if not by design.

One such diversification of the types of materials capable of exhibiting tunable release may go hand-in-hand with the distancing from the now classical tissue engineering paradigm of devising platforms that would regenerate, but not augment the replaced tissues. Thinking in this bold direction and engaging in the use of materials whose therapeutic effects may be due to their moderate toxicity and invasiveness of biophysical structures (every therapy is conditioned upon the creation of a wound) may bring us closer to other advanced concepts in the intrinsically tunable drug delivery. One of them would be the use of particle aggregation degrees through the control of surface chemistry and charge. Another direction worth exploring belongs to the exploitation of transitory, metastable, non-equilibrium states for the kinetic control of release patterns. Polymorphic transitions have been used to mediate drug release [99], but no tunable effects have been reported so far. Particle shell-to-core volume ratio is another parameter that has been used to produce different release profiles; however, the correlation between this parameter and release rate was rather volatile, not allowing for the tunable effect [100].
Finally, although the division to extrinsic/intrinsic and tunable/tailorable release carriers was introduced to clarify the perspective on them, it is expected that, as ever, conditions for the creation of the most prolific systems would be set on the grounds where the boundaries crush and multiple concepts merge into one.

Acknowledgments

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## Five bulleted summary points

- The fact that many biological processes take place at precise timescales implies that ideal therapeutic platforms would be capable of achieving a sequential, precisely temporally tuned delivery of the right therapeutic agents.

- Etymological clarification of the term “tunable” and its distinction from the term “tailorable” is provided: in the case of intrinsically tunable release, a particular compositional or structural parameter is tuned to give a desired release profile, whereas in the case of intrinsically tailorable release, adding or subtracting different carrier components is used to modify the release profile.

- Tunability can also be achieved extrinsically, being caused by the indiscrete variation of an environmental parameter; included in this category are carriers whose release is controlled by a specific disease marker and carriers whose release is controlled by external effects, e.g., heat, light, magnetic field, ultrasound, mechanical force, and electric current.

- A central problem in the elucidation of parameters that could be used as “magic knobs” that allow the drug release to be made intrinsically tunable is the inevitable entanglement of variables defining the microstructure of the carrier.

- Identical parameters often exert diametrically opposite effects on drug release in different carriers, illustrating the complexity of understanding and controlling the mechanisms of drug release in general.
Fig. 1.
Four types of kinetically controlled drug release. Two intrinsic types of control (a, b) can be either tailorable (a) if the rate of release (k) is directly proportional to a particular microstructural or compositional parameter of the carrier (A), or tunable (b) if the release is controlled by adding or subtracting different components of the carrier. Two extrinsic types of tunable release can be either smart, environmental (c) if the rate of release directly corresponds to the concentration of a chemical or biochemical marker in the environment surrounding the carrier or (d) manual if the rate of release is proportional to the intensity of an electromagnetic or acoustic signal produced by an externally operated device. A in the image refers to the microstructure or composition of the carrier, not the amount or concentration of the loaded drug. The three curves at the bottom refer to three kinetically distinct drug release profiles corresponding to the three different states of A.
Fig. 3.
Different release profiles of: (a) bovine serum albumin depending on the qualitative nature of the mesoporous calcium phosphate carrier; (b) vancomycin depending on the qualitative, not quantitative change in the cement composition; (c) SH3-IGF-1 from HAMC depending on whether the hydrogel is bound to a weak binding peptide (WBP) or to a strong binding peptide (SBP); (d) doxorubicin depending on the qualitative modification of silk films as carriers. Reprinted with permissions from Refs. [21] (a), [25] (b), [26] (c), and [27] (d).
Fig. 4.
pH-triggered release of PX, FL (a) and R6G (b) from PPy nanoparticles. (c) pH-responsive release of gentamicin from PEO-based polymeric nanoparticles covalently grafted onto titanium surface and functionalized with the antibiotic. (d) redox-responsive release of the antineoplastic drug, SN-38 from aniline-based polymeric nanoparticles. Reprinted with permission from Refs. [29] [30] and [31].
Fig. 5.
(a) Release of vancomycin from calcium phosphate pastes being tunable depending on the weight ratio between two of its components, HAP1 and HAP2. The linear function of the release rate versus the given weight ratio is shown in (b). (c) Release of daunorubicin from a silica-based drug delivery carrier with different nanopore sizes. (d) Release of horseradish peroxidase from gold-nanorod/PLGA microcapsules depending on the amount of PLGA in the precursor solution and the resulting microcapsule thickness. Reprinted with permission from Refs. [22] (a–b), [40] (c), and [57] (d).
Table 1

Types of drugs delivered with the controlled release and the associated pharmaceutical product names.

<table>
<thead>
<tr>
<th>Delivered type of molecule</th>
<th>Pharmaceutical product</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Anti-inflammatory drugs</td>
<td>Surodex</td>
<td>[6]</td>
</tr>
<tr>
<td>Corticosteroids</td>
<td>Ozurdex®, Retisert®, Iluvien®</td>
<td>[7]</td>
</tr>
<tr>
<td>Anticancer chemotherapeutics</td>
<td>Gliadel, ThermoDox, Decapepty®, Trelstar™</td>
<td>[8], [9].</td>
</tr>
<tr>
<td>Contraceptives</td>
<td>Norplant, Jadelle</td>
<td>[10]</td>
</tr>
<tr>
<td>Antipsychotics</td>
<td>Risperdal Consta™</td>
<td>[12]</td>
</tr>
<tr>
<td>Opioids</td>
<td>MS-Contin, Duragesic® (Fentanyl), Butrans®</td>
<td>[13]</td>
</tr>
<tr>
<td>Opioid antagonists</td>
<td>Vivitrol™</td>
<td>[14]</td>
</tr>
<tr>
<td>Antibiotics</td>
<td>To bone: Palacos® G (gentamicin), Cemex® Genta (gentamicin), Simplex® P (tobramycin), Combalt® G-HV (gentamicin), Samartset® GHV (gentamicin), and Versabond®; as oral tablets: Moxatag® (amoxicillin), Augmentin® XR (amoxicillin/ clavulanate potassium) and Cipro XR (ciprofloxacin)</td>
<td>[16], [17], [18]</td>
</tr>
</tbody>
</table>
Table 2
Examples of intrinsically tunable release carriers with intellectual property rights.

<table>
<thead>
<tr>
<th>Material</th>
<th>Filing date</th>
<th>Control parameter</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Organosiloxane-Poly(ethylene glycol) (PEG)</td>
<td>1988</td>
<td>Amount of the water-in-oil emulsion mixed in with the elastomer-forming composition</td>
<td>[66]</td>
</tr>
<tr>
<td>Alpha-cyclodextrin and PEG-poly(propylene glycol)-PEG</td>
<td>2001</td>
<td>Copolymer weight ratio</td>
<td>[67]</td>
</tr>
<tr>
<td>Polyurethane coating</td>
<td>2002</td>
<td>Duration of the low-energy plasma polymerization process</td>
<td>[68]</td>
</tr>
<tr>
<td>Poly(D, L-lactide-co-glycolide) (PLGA)-Hyaluronic acid (HA)</td>
<td>2007</td>
<td>Ratio of the hydrophilic (HA) to the hydrophobic matrix (PLGA)</td>
<td>[69]</td>
</tr>
<tr>
<td>PLGA coating</td>
<td>2008</td>
<td>Weight percent of acetone in acetone/methyl ethyl ketone spraying solution containing 2 wt. % polymer</td>
<td>[70]</td>
</tr>
<tr>
<td>Polycaprolactone (PCL)-PLGA core-sheath fibers</td>
<td>2009</td>
<td>Ratio between the feed rate of the inner (PCL) and outer (PLGA) portion solutions during electrospinning</td>
<td>[71]</td>
</tr>
<tr>
<td>PCL coating</td>
<td>2009</td>
<td>PCL molecular weight in the 10 – 80 kDa range</td>
<td>[72]</td>
</tr>
<tr>
<td>Hyaluronan and Methylcellulose (HAMC)</td>
<td>2010</td>
<td>MC content in HAMC, drug particle size</td>
<td>[73]</td>
</tr>
<tr>
<td>Hydroxyethylmethacrylate (HEMA)-chloroformate</td>
<td>2012</td>
<td>Conjugation degree of the prodrug to polymeric carrier moieties</td>
<td>[74]</td>
</tr>
<tr>
<td>Polystyrene-polyisobutylene</td>
<td>2015</td>
<td>Drug particle size, the weight ratio of volatile solvents in the solvent mixture</td>
<td>[75]</td>
</tr>
</tbody>
</table>
Table 3
Summary of the drug delivery carriers mentioned in the text and their categorization based on two major groups (intrinsic/extrinsic) and two subgroups (tunable/tailorable and smart/manual) described in Fig. 2, along with the key parameters for controlling the release kinetics.

<table>
<thead>
<tr>
<th>Drug release type</th>
<th>Material</th>
<th>Control parameter</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Intrinsic</td>
<td>Silicon</td>
<td>Porosity</td>
<td>[40], [41], [42]</td>
</tr>
<tr>
<td></td>
<td>Calcium alginate, Starch-gelatin, Gelatin, Polyethylene glycol (PEG),</td>
<td>Crosslinking degree</td>
<td>[33], [43], [45],</td>
</tr>
<tr>
<td></td>
<td>Trime thylene carbonate (TMC)-D,L-lactide (DLLA)</td>
<td></td>
<td>[46], [48]</td>
</tr>
<tr>
<td></td>
<td>Polyethylene glycol (PEG)</td>
<td>Molecular weight,</td>
<td>[46]</td>
</tr>
<tr>
<td></td>
<td>Poly(D, L-lactide-co-glycolide) (PLGA)</td>
<td>LA/GA ratio</td>
<td>[76], [49], [50],</td>
</tr>
<tr>
<td></td>
<td>Acrylamide (AAm)-(r)-α-acyrloyloxy-β,β-dimethyl-γ-butyrolactone (DBA)</td>
<td>Feed ratio</td>
<td>[53]</td>
</tr>
<tr>
<td></td>
<td>Polycaprolactone (PCL), PLGA, PEG-b-PLA, PLA-b-PNIPAm, Keratine (KTN)</td>
<td>Polymer content</td>
<td>[54], [55], [56],</td>
</tr>
<tr>
<td></td>
<td>/keratose (KOS), Gold/PLGA</td>
<td></td>
<td>[57]</td>
</tr>
<tr>
<td></td>
<td>Keratine (KTN)/keratose (KOS)</td>
<td>Drug identity</td>
<td>[56]</td>
</tr>
<tr>
<td></td>
<td>PLGA/titania</td>
<td>Polymer-metal mass</td>
<td>[60]</td>
</tr>
<tr>
<td></td>
<td>Carboxymethyl cellulose</td>
<td>Degree of oxidation</td>
<td>[61]</td>
</tr>
<tr>
<td></td>
<td>Hyaluronic and deoxycholic acid conjugates (HA-DOCA)</td>
<td>Degree of substitution, Drug content</td>
<td>[62]</td>
</tr>
<tr>
<td></td>
<td>Magnesium carbonate</td>
<td>Particle size</td>
<td>[63]</td>
</tr>
<tr>
<td></td>
<td>PCL/silk fibroin</td>
<td>Drug content</td>
<td>[64]</td>
</tr>
<tr>
<td>Tailorable</td>
<td>Alginate, Sodium carboxymethyl cellulose (Na-CMC), Viscose Rayon and</td>
<td>Different material combinations</td>
<td>[23]</td>
</tr>
<tr>
<td></td>
<td>Poly(ethylene tere phthalate) (PET)</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>Calcium phosphate</td>
<td>Phase composition, addition of the drug</td>
<td>[21], [25]</td>
</tr>
<tr>
<td></td>
<td></td>
<td>or of poly(D, L-lactide-co-glycolide)</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>(PLGA)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Hyaluronan and methylcellulose (HAMC)</td>
<td>Binding of peptides</td>
<td>[26]</td>
</tr>
<tr>
<td></td>
<td>Chitosan</td>
<td>Conjugation of L-leucine</td>
<td>[24]</td>
</tr>
<tr>
<td></td>
<td>Silk</td>
<td>Modification by carboxyl or sulfonate</td>
<td>[27]</td>
</tr>
<tr>
<td>Smart</td>
<td>Polypyrrole (PPy), Polyethylene oxide/titanium</td>
<td>pH</td>
<td>[29], [30]</td>
</tr>
<tr>
<td></td>
<td>2,6-bis(hydroxymethyl)aniline-based polymers</td>
<td>Cytosolic cleavage of disulfide or</td>
<td>[31]</td>
</tr>
<tr>
<td></td>
<td></td>
<td>thioester bonds</td>
<td></td>
</tr>
<tr>
<td></td>
<td>O2-(β-D-galactopyranosyl)1-(N-isopropylamino)di azene-1-im-1,2-diolate</td>
<td>Enzyme concentration</td>
<td>[28]</td>
</tr>
<tr>
<td></td>
<td>(Gal-IPA/NO)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Manual</td>
<td>PPy, Calcium alginate</td>
<td>Electric field strength</td>
<td>[32], [33]</td>
</tr>
<tr>
<td></td>
<td>Gold/Nickel/Gold/Nickel/Chromium nanomotor</td>
<td>Rotation speed</td>
<td>[36]</td>
</tr>
<tr>
<td></td>
<td>Gelatin-PNIPAm-co-AAm</td>
<td>Temperature</td>
<td>[34]</td>
</tr>
<tr>
<td></td>
<td>Polystyrene-PDMS block co-polymers</td>
<td>Magnetic field intensity</td>
<td>[37]</td>
</tr>
<tr>
<td></td>
<td>Hyaluronic acid-copper sulfide</td>
<td>Ultrasound irradiation on/off modulation</td>
<td>[38]</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Photo thermal laser irradiation on/off</td>
<td>[39]</td>
</tr>
</tbody>
</table>

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