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Inferring foliar water uptake using stable isotopes of water

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Author Contributions

GRG and RS conceived of the project. GRG, MML, and LC performed the research with assistance from MA. GRG analyzed the data and wrote the paper with contributions from all the authors.

Abstract

A growing number of studies have described the direct absorption of water into leaves, a phenomenon known as foliar water uptake. The resultant increase in the amount of water in the leaf can be important for plant function. Exposing leaves to isotopically enriched or depleted water sources has become a common method for establishing whether or not a plant is capable of carrying out foliar water uptake. However, a careful inspection of our understanding of the fluxes of water isotopes between leaves and the atmosphere under high humidity conditions shows that there can clearly be isotopic exchange between the two pools even in the absence of a change in the mass of water in the leaf. We provide experimental evidence that while leaf water isotope ratios may change following exposure to a fog event using water with a depleted oxygen isotope ratio, leaf mass only changes when leaves are experiencing a water deficit that creates a driving gradient for the uptake of water by the leaf. Studies that rely on stable isotopes of water as a means of studying plant water use, particularly with respect to foliar water uptake, must consider the effects of these isotopic exchange processes.

Key words: fog, isotope dendrochronology, leaf wetting, plant-water relations, stomata

Body of Text

Foliar water uptake describes the process by which plants absorb water into their leaves, resulting in a net increase in the mass of water in the leaf. This occurs when saturating atmospheric water vapor conditions result in a driving gradient for water to enter into a leaf that is at a more negative water potential (Simonin et al. 2009; Goldsmith 2013; Vesala et al. 2017). The conditions necessary for this phenomenon are often observed in dew- and fog-affected ecosystems such as coastal Mediterranean ecosystems (Burgess and Dawson 2004; Baguskas et al. 2016) and tropical montane cloud forests (Eller et al. 2013; Gotsch et al. 2014; Malhi et al. 2017), where fog (often leading to leaf wetting) serves as an alternative plant water source during the dry season. However, the effects of precipitation events are similar and foliar water uptake has now been described as affecting plant water and carbon relations in > 70 species from a number of different ecosystems (Goldsmith et al. 2013).

The capacity for species to do foliar water uptake has frequently been established by means of water isotope labeling experiments (Burgess and Dawson 2004; Breshears et al. 2008; Limm et al. 2009; Eller et al. 2013; Berry and Smith 2014; Berry et al. 2014; Gotsch et al. 2014; Cassana et al. 2015; Eller et al. 2016; Emery 2016; Schwerbrock and Leuschner 2017). This method relies on exposing leaves to a water source that is highly enriched or depleted in heavy isotopes of oxygen or hydrogen compared to that of the xylem source water and measuring for the presence of this label in the leaf. The labeled water is delivered through a simulated fog or leaf wetting event using an ultrasonic fog machine or a simple spray bottle. A shift in the bulk leaf water isotope ratio towards that of the labeled water source is interpreted as foliar water uptake.

Water molecules can also exchange back and forth between the leaf and the atmosphere without a net increase in the mass of water in the leaf (Kim and Lee 2011). This occurs when atmospheric vapor pressure (e_a) increases relative to leaf vapor pressure (e_i). As a result, the air to leaf vapor pressure deficit decreases and reduces the driving gradient for water loss from the leaf. Stomata generally open in response to decreasing VPD (Lange et al. 1971), leading to an increase in leaf stomatal conductance (g_s) in the light, even though net transpiration (E) is decreasing to zero. Thus, with the stomata open, but transpiration suppressed due to e_a/e_i reaching unity (i.e. 100% relative humidity), water molecules simply move from the leaf to the atmosphere and vice versa with no net flux. This is akin to isotopic exchange between two pools of water in a closed system (Clark and Fritz 1997). Notably, this changes the isotope ratio of the leaf water, but does not lead to a net increase in the amount of water in the leaf. Thus, it is not possible to distinguish between a change in the leaf water isotope ratio due to foliar water uptake (net gain H_2O) versus a change caused solely by water isotopes simply exchanging back and forth (no net change H_2O) between the leaf and the atmosphere (Figure 1).

The effect of the isotope exchange of water across the leaf surface on leaf water isotope ratios at steady state is described by the Craig-Gordon Model (Craig and Gordon 1965; Dongmann et al. 1974):

$$_e = ^+ + _k + (_v \quad _k)\frac{e_a}{e_i} \qquad (1)$$

where the enrichment in leaf water isotopes relative to the source (Δ_e) is a function of equilibrium (ε^+) and kinetic fractionation factors (ε_k), the enrichment in atmospheric water vapor isotopes relative to source water isotopes (Δ_v), and the ratio of ambient air vapor pressure to leaf intracellular vapor pressure (e_a/e_i). Equilibrium fractionation occurs with the phase change of water from liquid to vapor within the stomata, whereas kinetic fractionation occurs with the diffusion of that vapor through the stomata and boundary layer into the atmosphere. Dongmann et al. (1974) notes that when e_a/e_i is at unity, the model simplifies to:

$$\Delta_e = \varepsilon^+ + \Delta_v \qquad (2)$$

In Eq. 2, the stable isotope ratio of water in leaves is not subject to kinetic fractionation and can be explained solely by a temperature-dependent equilibrium fractionation factor and the difference in atmospheric water vapor isotopes relative to source water isotopes. It is important to note that this theory cannot distinguish between the effects of foliar water uptake versus bi-directional exchange.

The exchange of water isotopes between leaves and the atmosphere in the absence of foliar water uptake can be demonstrated experimentally. We exposed leaves from well-watered poplar (*Populus x canescens*) plants growing in a high humidity (~80%) growth chamber to a fog event using water with a depleted oxygen isotope ratio. We excised leaves at full water content, measured the leaf mass, and sealed the petioles from water entry. Leaves were either immediately exposed to fog, or allowed to lose 5 or 10% of their initial mass prior to fog exposure to create a driving gradient for water to enter the leaf through foliar water uptake (n = 1 leaf each from 5 individuals per treatment). Fog was generated using an ultrasonic fog machine (Ultrasonic 3, CIS Products, France) and supplemented by periodic physical spraying for 1 h (i.e. fog leading to leaf wetting). We then quickly and carefully dried the leaf surfaces, re-measured the leaf mass, and sealed the leaf in a glass vial to later measure the oxygen isotope ratios of bulk leaf water via isotope ratio mass spectrometry. We compared the three treatments (0, 5, and 10% mass loss) with the isotope ratio of the source water provided to the plants (for methods, see Online Resource). All data are available in the KNB data repository (Goldsmith et al. 2017).

The leaf water isotope ratios of all three treatments shifted towards the depleted isotopic label following fog exposure (Figure 2). However, the leaf mass of the 0% treatment did not change, indicating the exchange of water isotopes even though there was no foliar water uptake. The 5 and 10% treatments recovered some (but not all) of their initial mass, as would be predicted by the establishment of a driving gradient for foliar water uptake. Due to the short duration of the experiment, the leaf water isotope ratios did not converge with the labeled water vapor, as would be predicted by theory and has been observed in other experimental approaches (Kim and Lee 2011).

The observation that leaf water isotopes exchange even in the absence of a change in leaf mass is of particular importance if the primary pathway for foliar water uptake is stomata (Burkhardt et al. 2012). However, the pathways for foliar water uptake are not yet fully resolved. There is evidence for water entry through hydathodes, trichomes, fungal hyphae and the cuticle, depending on the species under study (Burgess and Dawson 2004; Oliveira et al. 2014). However, even if the primary pathways were to be something other than the stomata, inferences regarding foliar water uptake could still be confounded by bi-directional exchange through open (or at night, partially open) stomata.

There are other analogous applications of water isotope tracers that should also be considered. Branch water uptake directly through bark has been studied by submerging branch segments into labeled water sources; even in the absence of stomata, it is likely that isotopic exchange will occur given sufficient time (Mayr et al. 2014; Earles et al. 2015). Several studies have also used differences between the stable isotope ratios of fog versus soil water to infer the proportional use of these two sources through sampling of xylem water isotope ratios (Berry et al. 2013; Fischer et al. 2016; Fu et al. 2016). Here, it must be assumed that the water in the xylem could come from a combination of the 1) soil water derived from precipitation, 2) drip of intercepted fog water from the plant canopy into the soil, 3) foliar uptake of fog water, or 4) bi-directional exchange of fog water with leaf water. The subsequent incorporation of these water isotopes into plant tissue (e.g. $\delta^{18}O$ of tree rings) has also been proposed as a means of tracing the contribution of fog water to plant water use over time (Hu and Riveros-Iregui 2016); our results may help explain patterns observed in fog-affected environments (Anchukaitis et al. 2008; Zhu et al. 2012).

Foliar water uptake remains a real phenomenon. There are a number of different methods to independently establish its existence, including sapflow (Burgess and Dawson 2004), dye tracers (Eller et al. 2013), gravimetric approaches (Limm et al. 2009), and plant water potentials (Goldsmith et al. 2013). In fact, many of the research studies cited above combined stable isotope labeling experiments with other methods and thus the results are likely to stand. As no single method is perfect, we recommend that investigators try to use multiple means to establish foliar water uptake wherever possible.

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Figure Legend

Figure 1. Leaf water isotope ratios can vary due to (a) the effects of transpiration when the ratio of ambient air vapor pressure to leaf intracellular vapor pressure (e_a/e_i) < 1, (b) the effects of transpiration suppression when e_a/e_i = 1, which results in no change in the mass of leaf water and (c) the effects of foliar water uptake when $e_a/e_i \geq 1$, which results in a net increase in the mass of leaf water. The effects of bi-directional exchange of water isotopes between the leaf and the atmosphere observed in (b) cannot be distinguished from the net uptake of water isotopes from the atmosphere in (c).

Figure 2. Changes in (a) the $\delta^{18}O$ of water and (b) the percent change in fresh leaf mass of poplar (*Populus x canescens*) leaves exposed to a fog source water depleted in $\delta^{18}O$ after 0, 5, and 10% loss of fresh leaf mass achieved through bench drying (n = 1 leaf each from 5 individuals per treatment). Data are means ± 1 SD.

Figure 1

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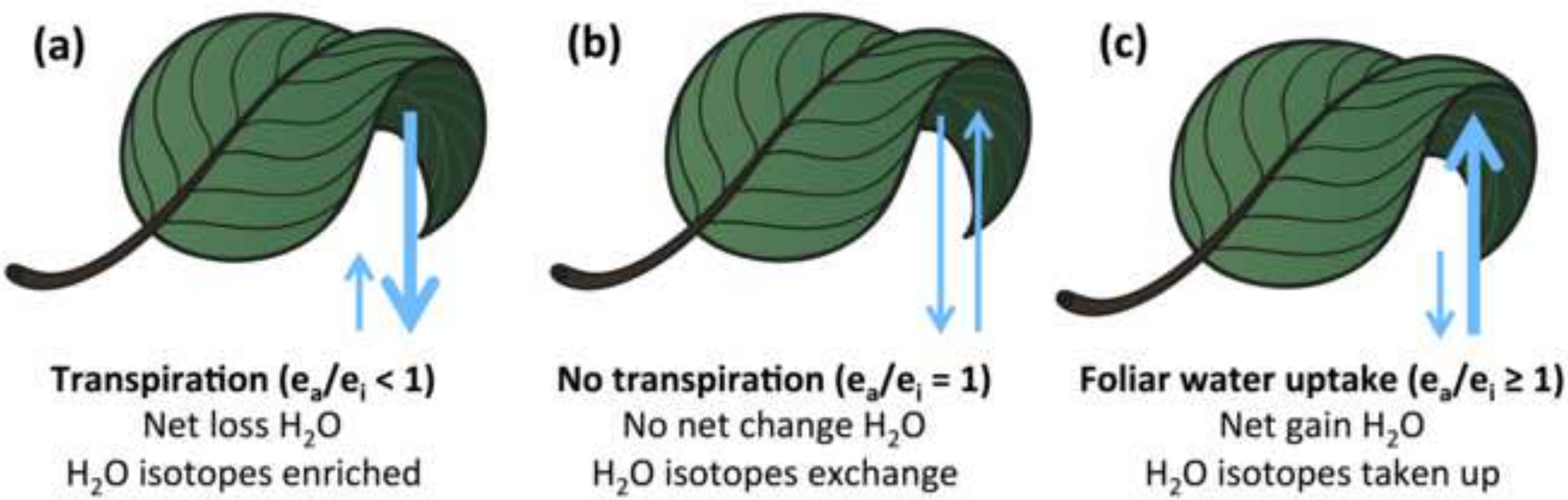
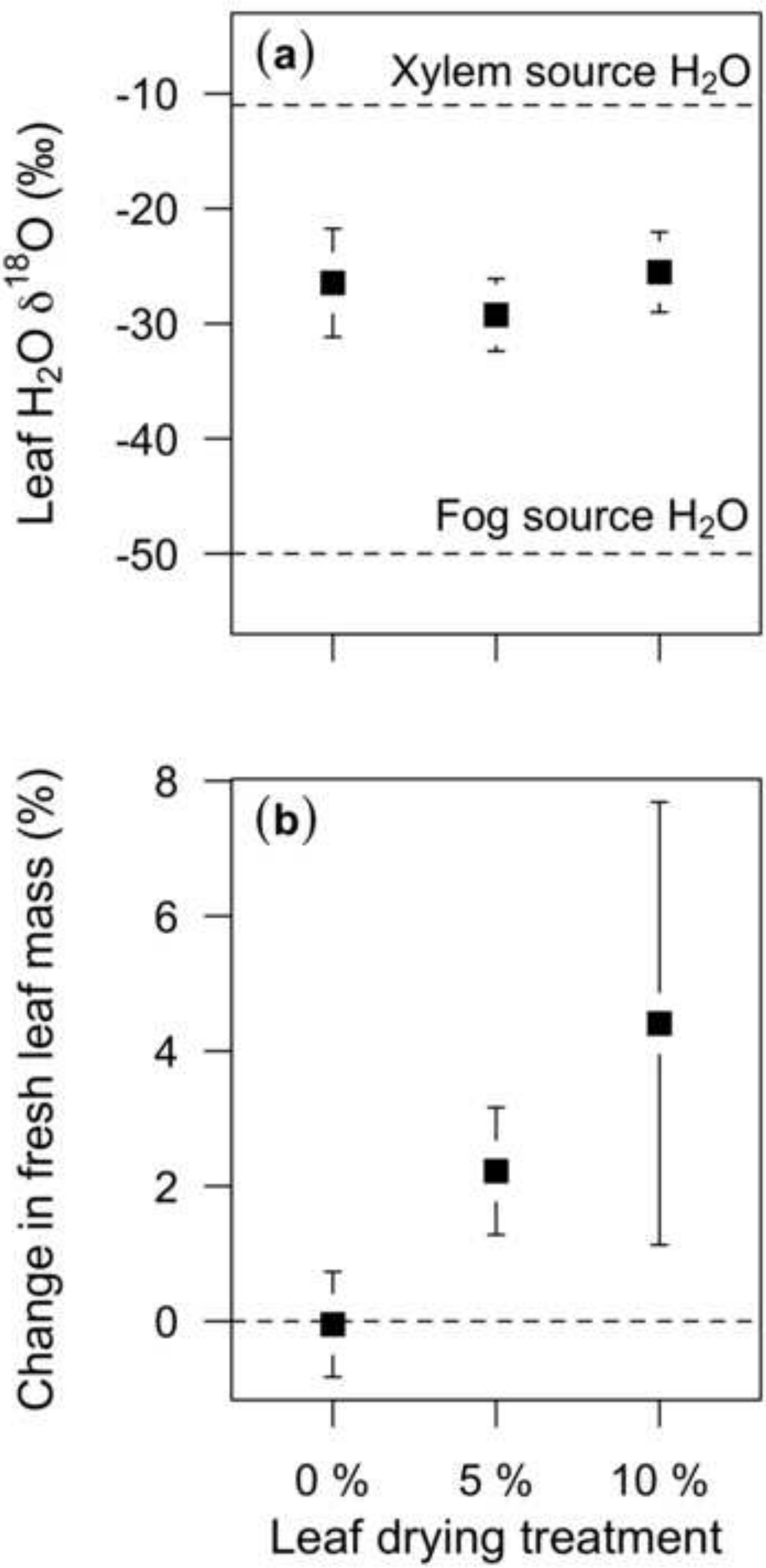


Figure 2


(a)
Xylem source H2O
Fog source H2O
Leaf H2O δ18O (‰)
-10
-20
-30
-40
-50
(b)
Change in fresh leaf mass (%)
8
6
4
2
0
0 %
5 %
10 %
Leaf drying treatment