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### Species Substitution and Mislabeling of Ceviche, Poke, and Sushi Dishes Sold in Orange County, California

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## Species Substitution and Mislabeling of Ceviche, Poke, and Sushi Dishes Sold in Orange County, California

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3

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## Abstract

Raw, ready-to-eat (RTE) seafood products have become increasingly popular globally, but they are vulnerable to species substitution and mislabeling. DNA barcoding allows for fish species identification by extracting, amplifying, and sequencing a standardized gene target. A wide variety of fish products have been studied with DNA barcoding, but little investigation of ceviche and poke has occurred in the United States. Sushi is known to be a target of mislabeling but has not been extensively studied in Orange County, CA. The objective of this study was to investigate species substitution and mislabeling of sushi, poke, and ceviche dishes sold at restaurants in Orange County, CA. A total of 105 raw, RTE seafood products were collected, including sushi (n = 35), poke (n = 35), and ceviche (n = 35). All samples were sequenced with DNA barcoding or mini-barcoding. The identified species were compared against the menu names, verbal declarations by restaurant staff, and the Food and Drug Administration (FDA) Seafood List to verify whether acceptable market names were used for each product. Of the 103 samples identified with DNA barcoding, species substitution was detected at a rate of 23.3% and unacceptable market names were found in 45.6% of samples. Overall, 63.1% of samples had some form of mislabeling. When the mislabeling rates were divided based on product category, ceviche had the highest overall mislabeling rate (85.3%), followed by poke (61.8%), and sushi (42.9%). Mislabeling of ceviche and poke was primarily driven by the use of unacceptable market names, while species substitution was more common in sushi dishes. These rates reveal widespread mislabeling among raw, RTE seafood products and suggest the need for outreach efforts to ensure proper labeling of fish using acceptable market names, as well as further research into mislabeling within the raw, RTE seafood supply chain.

**Keywords:** mislabeling; substitution; DNA barcoding; ready-to-eat; seafood

47 **1. Introduction**

48 Global annual per capita consumption of aquatic foods reached a record high of 20.5 kg in  
49 2019 (FAO, 2022). In the United States in 2020, a total of 3.8 billion kg of seafood landings  
50 were recorded, valued at US\$4.8 billion (NOAA, 2022). Due to the health benefits of seafood,  
51 the Dietary Guidelines for Americans recommends consuming at least 227 g of seafood per week  
52 (USDA, 2020). Live, fresh, and chilled products make up the largest share of seafood sold for  
53 direct human consumption (FAO, 2022). Some of the most widely consumed raw, ready-to-eat  
54 (RTE) seafood dishes in the U.S. include ceviche, poke, and sushi. Ceviche is a raw, marinated  
55 fish dish originating from Peru that is typically mixed with other ingredients, such as onions,  
56 peppers, and seasonings. The fish is cut into small pieces and marinated in acidic citrus juices,  
57 with the most common fish used in U.S. recipes being tilapia (Mathur & Schaffner, 2013). Poke  
58 is a cubed raw fish product originating from Hawaii that is mixed with seasonings such as soy  
59 sauce, onions, and peppers (Hospital & Beavers, 2014). Some of the most common fish used to  
60 make poke are tuna, salmon, and yellowtail. Sushi is a broad category of Japanese cuisine  
61 involving small cuts of seafood and/or vegetables served with rice. Numerous fish species are  
62 used in sushi, including tuna, salmon, halibut, and snapper (Warner et al., 2013).

63 Raw, RTE seafood products are particularly vulnerable to species substitution and  
64 mislabeling due to their high price points in the market and lack of morphological identifiers.  
65 Mislabeling of seafood can be intentional or accidental, with the former typically driven by  
66 economic motivation (Silva et al., 2021). Mislabeling can happen at any point between harvest  
67 and consumption; incorrect use of common names, lost product, or outright substitution of  
68 species can all contribute to species mislabeling. Complexity within the global seafood supply

69 chain and varying labeling requirements across countries further increase the risk of seafood  
70 substitution and mislabeling (Silva et al., 2021).

71 Identifying seafood substitution and mislabeling within the seafood market is critical for  
72 safety, cultural significance, conservation measures, and protecting consumer interests. With  
73 regards to safety, the primary concerns surround allergies, contaminants and toxins that may be  
74 present (Silva et al., 2021). Seafood species substitution can be dangerous for consumers with  
75 allergies to fish or shellfish if the substituted species contains allergens that are not present in the  
76 declared species. Food safety is also a concern with species that contain toxins, such as  
77 gempylotoxin (oilfish), tetrodotoxin (pufferfish), or elevated levels of mercury (e.g., bigeye tuna)  
78 (Silva et al., 2021). The mislabeling of threatened species, such as substituting the endangered  
79 southern bluefin tuna (*Thunnus maccoyii*) for yellowfin tuna (*Thunnus alalunga*), can also  
80 impact conservation programs (Liou et al., 2020). Finally, the detection of seafood substitution  
81 and mislabeling is essential to protecting customer interests by ensuring that consumers receive  
82 what they paid for.

83 Due to the lack of morphological features in raw, RTE seafood, analytical methods are often  
84 required for species identification. DNA barcoding is a sequencing-based method that relies on  
85 the differentiation of species using a short, standardized genetic target (Hebert et al., 2003). The  
86 primary marker used with animal species, including fish, is a ~650 bp region of the gene coding  
87 for cytochrome c oxidase subunit I (COI). This genetic target is ideal for most instances of  
88 seafood species identification because it demonstrates relatively low intra-species variation and  
89 high inter-species variation. DNA barcoding of COI is widely used for the DNA-based  
90 identification of seafood and is utilized by the U.S. Food and Drug Administration (FDA) for  
91 regulatory testing of fish species (Handy et al., 2011). However, COI barcoding is frequently

92 insufficient for species identification within the *Thunnus* genus due low genetic divergences  
93 (Pollack et al., 2018; Viñas & Tudela, 2009). In these cases, the mitochondrial control region  
94 (CR) is often used (Gordoa et al., 2017; Mitchell & Hellberg, 2016; Rongchun et al., 2022;  
95 Viñas & Tudela, 2009).

96 DNA barcoding has been used in several studies to reveal mislabeling of raw, RTE seafood,  
97 specifically ceviche (Velez-Zuazo et al., 2021) and sushi (Chang et al., 2021; Lowenstein et al.,  
98 2009; Rongchun et al., 2022; Velez-Zuazo et al., 2021; Warner et al., 2013; Willette et al.,  
99 2017). A study evaluating fish mislabeling in Southern California reported a mislabeling rate of  
100 84% for sushi restaurants, including raw "white tuna" samples that were actually escolar  
101 (*Lepidocybium flavobrunneum*), which contains gempylotoxin (Warner et al., 2012, 2013).  
102 Another study investigating mislabeling in Los Angeles, CA, from 2012 to 2015 reported a  
103 mislabeling rate of 47%, with elevated mislabeling rates in sushi products labeled as halibut, red  
104 snapper, yellowfin tuna and yellowtail (Willette et al., 2017). Seafood mislabeling is a major  
105 issue globally as well (Pardo et al., 2016); for example, 17% of sushi samples were found to be  
106 mislabeled in a study in Taiwan (Chang et al., 2021), and a mislabeling rate of 26% was found in  
107 Japanese restaurants in Brazil (Staffen et al., 2017). Ceviche was previously reported to be  
108 mislabeled in 78% of samples in Peru; however, no studies have been conducted on ceviche  
109 mislabeling in the United States (Velez-Zuazo et al., 2021).

110 While relatively high levels of seafood mislabeling have been reported for sushi dishes sold  
111 in Southern California, there is a lack of research on mislabeling of ceviche dishes sold in the  
112 U.S. Furthermore, to the authors' knowledge, there are no studies focused specifically on  
113 mislabeling of poke dishes. Therefore, the objective of this study was to evaluate species  
114 substitution and mislabeling of ceviche, poke, and sushi products sold at restaurants in Orange

115 County, CA. Sushi was included in this study to allow for a direct comparison of mislabeling  
116 across the three categories of raw, RTE seafood.

## 117 **2. Materials and Methods**

### 118 **2.1. Sample Collection**

119 A total of 105 raw, RTE fish products marketed as sushi, poke, or ceviche were collected  
120 from restaurants (n = 67) across Orange County, CA. Equal numbers of samples were collected  
121 from each RTE category (i.e., 35 samples each of ceviche, poke, and sushi dishes). Additional  
122 details on the individual samples collected are provided in Table S1. The selection of seafood at  
123 each location was based on availability and each sample was unique (i.e., no duplicate samples  
124 from the same location). In cases of chain restaurants, only one location of the same chain  
125 restaurant was visited. The market name and menu description for each fish product was  
126 documented. In cases where dishes were labeled generically as “fish”, “white fish”, or “pescado”  
127 (a Spanish word for edible fish), verbal confirmation of the species was requested from staff and  
128 documented. The fish samples were then transported to the laboratory in a cooler with ice packs  
129 and stored in a freezer at  $-80^{\circ}\text{C}$  or immediately processed for DNA extraction. To prepare for  
130 DNA extraction, a  $\sim 10$  mg sample from the interior of the product was removed using sterile  
131 forceps and placed in a 1.5 mL sterile microcentrifuge tube (Handy et al., 2011). The  
132 microcentrifuge tubes containing sample tissue were stored at  $4^{\circ}\text{C}$  for up to 3 days until DNA  
133 extraction. This allowed for collection of a sufficient number of samples for a batch DNA  
134 extraction rather than extracting individual samples on each collection day. The remaining  
135 product was stored at  $-80^{\circ}\text{C}$ .

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137



## 138 **2.2. DNA Extraction**

139 The spin column DNA extraction protocol outlined in Handy et al. (2011) was performed on  
140 each tissue sample using a DNeasy Blood and Tissue Kit (Qiagen, Germany). Lysis was carried  
141 out using an Eppendorf ThermoMixer C (Germany) set at 56°C with a mixing speed of 300 rpm  
142 for 2 h. DNA was eluted in 100 µL of pre-heated (37°C) Buffer AE (DNeasy Blood and Tissue  
143 Kit). The extracted DNA underwent PCR immediately or was stored at –80°C for future use.  
144 Each set of DNA extractions included a negative control with no fish tissue (i.e., reagent blank).

## 145 **2.3. PCR and DNA Sequencing**

146 COI DNA barcoding was conducted on all samples using the primers and cycling conditions  
147 described in Handy et al. (2011). To prepare the samples for PCR, 9 µL of sterile water, 11.5 µL  
148 of HotStarTaq Plus Master Mix (Qiagen), 0.25 µL of each 10 µM primer and 2 µL of DNA  
149 template were added to each reaction tube. Each PCR run included a negative control with sterile  
150 water instead of template DNA (i.e., non-template control). All samples labeled as tuna were also  
151 tested with the CR mini-barcode primers and cycling conditions described in Mitchell and  
152 Hellberg (2016) to facilitate species discrimination. For CR mini-barcoding, each reaction tube  
153 contained 7 µL sterile water, 11.5 µL HotStarTaq Plus Master Mix, 0.5 µL of each 10 µM  
154 primer, and 3 µL DNA template. Samples that could not be identified with COI DNA barcoding  
155 or CR mini-barcoding (in the case of tuna) were tested with COI mini-barcoding using the SH-E  
156 primers and cycling conditions described in Shokralla et al. (2015). For COI mini-barcoding,  
157 each reaction tube contained 8.5 µL sterile water, 11.5 µL HotStarTaq Plus Master Mix, 0.5 µL  
158 of each 10 µM primer, and 2 µL DNA template.

159 PCR products were confirmed with agarose gel electrophoresis as described previously  
160 (Roungchun et al. 2022). Gels were visualized with an E-Gel Imager (ThermoFisher Scientific,

161 USA) containing a UV base and universal filter. All PCR products were purified with ExoSAP-  
162 IT (ThermoFisher Scientific) according to the manufacturer's instructions and sent to Eurofins  
163 Genomics (USA) for sequencing. Bidirectional sequencing was carried out with a BigDye  
164 Terminator v3.1 cycle Sequencing Kit (Applied Biosystems, USA) and a 3730xl DNA Analyzer  
165 (Applied Biosystems).

166 Geneious R7 (Biomatters, Ltd., New Zealand) was utilized to assemble and edit the raw  
167 sequence data. The sequences were trimmed to the target regions for the respective barcode  
168 targeted. The QC parameters described in Handy et al. (2011) were applied to the COI DNA  
169 barcodes and the QC parameters described in Pollack et al. (2018) were applied to the COI and  
170 CR mini-barcodes. The resulting DNA barcode consensus sequences associated with each  
171 sample can be found in Table S1.

172 COI DNA barcode and mini-barcode sequences were queried against the Barcode of Life  
173 Database (BOLD), Public Records. CR mini-barcode sequences were queried against GenBank  
174 utilizing the Basic Local Alignment Search Tool (BLAST). Each species that was identified was  
175 compared against the species that was advertised and checked for the use of an acceptable  
176 market name based on guidance provided in *The Seafood List* (FDA, 2022).

#### 177 **2.4. Statistical Analysis**

178 The mislabeling rates of raw, RTE seafood products were compared across the three product  
179 categories (i.e., poke, ceviche, and sushi) using a Fisher's exact test with a Bonferroni correction  
180 for multiple tests. This test was used to compare the rates of species substitution, use of  
181 unacceptable market names, and overall mislabeling across the three product categories. Price  
182 was examined as a potential factor affecting mislabeling (i.e., species substitution, unacceptable  
183 market names, and overall mislabeling) within each product category using a Kruskal-Wallis H

184 test. Three ceviche samples were missing price data and therefore were excluded from the  
185 analysis (Table S1). Mislabeling rates (i.e., species substitution, unacceptable market names, and  
186 overall mislabeling) of chain and independent stores were also compared across the combined  
187 data set using Fisher's exact test. Statistical analysis was performed using R Studio version 4.0.2  
188 (RStudio, PBC, USA) (R Core Team, 2020).

### 189 **3. Results**

#### 190 **3.1. DNA Barcoding**

191 Of the 105 raw, RTE seafood samples tested in this study, 103 passed DNA sequencing  
192 quality control and were identified with at least one of the DNA barcoding methods utilized (Fig.  
193 1). Among the COI DNA barcoding methods, the standard COI barcode allowed for  
194 identification of 91 samples and the COI mini-barcode identified 10 samples (Table S1). The 10  
195 samples that failed COI DNA barcoding and were instead identified with COI mini-barcodes  
196 were found to be Atlantic salmon (*Salmo salar*; n = 2), Atlantic mackerel (*Scomber scombrus*; n  
197 = 2), buri (*Seriola quinqueradiata*; n = 1), walleye pollock (*Gadus chalcogrammus*; n = 1),  
198 walleye pollock/Arctic cod (*Gadus chalcogrammus/Boreogadus saida*; n = 1), whiteleg shrimp  
199 (*Litopenaeus vannamei*; n = 2), and yellowtail (*Seriola lalandi*; n = 1). CR mini-barcoding  
200 allowed for identification of an additional two samples that failed COI DNA barcoding, as well  
201 as species confirmation for 29 samples that were also sequenced with the COI DNA barcode  
202 (results discussed in section 3.2).

203 A poke sample labeled as albacore tuna failed assembly for the COI DNA barcode and  
204 failed quality control for the CR mini-barcode due to having 2.3% ambiguities. The sample was  
205 identified as albacore tuna with 96.8% identity in GenBank. However, due to the relatively high  
206 number of ambiguities (>2%), this identification is not considered reliable and was not used in

207 the determination of species labeling. The other sample that did not pass quality control was a  
208 ceviche dish labeled as “fish”. This sample failed assembly for the COI DNA barcode and failed  
209 quality control for the COI mini-barcode due to its short sequence length (125 bp). This sequence  
210 was poor quality with 0% high quality bases and multiple overlapping peaks, indicating the  
211 possibility of a species mixture.

212         When comparing DNA barcoding success across the three raw, RTE product categories,  
213 the ceviche samples had the lowest success rate (82.9%) for COI DNA barcoding, with five  
214 samples identified with COI mini-barcoding instead and one sample that was unable to be  
215 identified with COI DNA barcoding or mini-barcoding (Table S1). The poke and sushi  
216 categories showed a slightly higher success rate (88.6%) for COI DNA barcoding, with four  
217 samples each that could not be identified with COI DNA barcoding. Of the four poke samples  
218 that could not be identified with COI DNA barcoding, two were identified with COI mini-  
219 barcoding, one was identified with CR mini-barcoding, and one sample could not be identified.  
220 Of the four sushi samples that could not be identified with COI DNA barcoding, three were  
221 identified with COI mini-barcoding and one was identified with CR mini-barcoding. The  
222 preparation methods used for these dishes, which include soaking the fish in citrus juices/citric  
223 acid (ceviche), as well as the use of acidic sauces (poke and sushi), may have contributed to  
224 DNA degradation. However, the results of this study show that the use of smaller target  
225 sequences, such as the COI mini-barcode (Shokralla et al., 2015), can alleviate this issue.

### 226 **3.2. Species Identification**

227         All 101 of the samples identified with COI DNA barcoding or mini-barcoding had one or  
228 more top species matches in BOLD with > 98% similarity and the 31 samples identified with CR  
229 mini-barcoding had one or more top species matches in GenBank with > 95% identity. These

230 results are consistent with previous studies, which have reported > 98% similarity for COI  
231 species matches and > 95% identity for CR mini-barcode species matches (Liou et al., 2020;  
232 Mitchell & Hellberg, 2016). The CR genetic marker typically shows lower levels of intraspecies  
233 similarity compared to COI due to its greater rate of divergence (Roungchun et al., 2022).  
234 Among the 31 tuna samples collected in this study, CR mini-barcoding confirmed the species  
235 level identifications obtained with COI DNA barcoding in 15 samples and produced species-  
236 level identifications for the remaining 16 samples that either failed COI DNA barcoding (n = 2)  
237 or were only identified to the genus level with COI DNA barcoding (n = 14). The tuna species  
238 identified in this study included albacore tuna (*Thunnus alalunga*; n = 11), yellowfin tuna  
239 (*Thunnus albacares*; n = 10), southern bluefin tuna (*Thunnus maccoyii*; n = 5), and bigeye tuna  
240 (*Thunnus obesus*; n = 5).

241         When taking into account the combined results for COI and CR barcoding, 94 samples  
242 were identified to the species level (i.e., showed a top genetic match to a single species), eight  
243 samples were identified to the genus level (i.e., showed equivalent top genetic matches to  
244 multiple species from the same genus), and one had equivalent top genetic matches to species  
245 from multiple genera within the same family (Fig. 1). One sushi sample labeled as seabream had  
246 equivalent genetic matches to both madai (*Pagrus major*) and Nile tilapia (*Oreochromis*  
247 *niloticus*). However, upon further inspection it was found that the Nile tilapia identification was  
248 likely due to an erroneous sequence (Accession ID MT525054) in the database and the sample  
249 was therefore determined to be madai.

250         The eight samples identified to the genus level showed equivalent top matches to  
251 multiple tilapia species (*Oreochromis* sp.), primarily Nile tilapia (*Oreochromis niloticus*), blue  
252 tilapia (*Oreochromis aureus*), and Mozambique tilapia (*Oreochromis mossambicus*). Tilapia

253 species are often cross-bred, and the resulting hybrids cannot be differentiated with  
254 mitochondrial DNA (Liou et al., 2020; Peterson et al., 2021). However, the identification of  
255 tilapia to the genus level is sufficient for confirming samples labeled as “tilapia” and is  
256 consistent with previous studies that have used COI barcoding methodologies (Liou et al., 2020;  
257 Peterson et al., 2021).

258         The one sample with a family level match (Gadidae) was a ceviche dish that showed  
259 equivalent top genetic matches to walleye pollock (*Gadus chalcogrammus*) and Arctic cod  
260 (*Boreogadus saida*). Among the top 100 genetic matches to the query sequence for this sample,  
261 97 entries were associated with walleye pollock and only three entries were associated with  
262 Arctic cod. All three entries for Arctic cod were associated with the same published study that  
263 performed COI DNA barcoding on fish larvae and juveniles (Bouchard et al., 2021), which can  
264 be especially difficult to differentiate (Wildes et al., 2022). Furthermore, Wildes et al. (2022)  
265 reported that walleye pollock is frequently misidentified as Arctic cod based on morphological  
266 features. Taken together, it is possible that the genetic matches to Arctic cod were associated  
267 with walleye pollock specimens that were morphologically misclassified as Arctic cod. In this  
268 case the use of additional genetic markers, such as microsatellites or other mitochondrial targets,  
269 would be helpful to determine whether the sample is indeed walleye pollock (Wildes et al.,  
270 2022). However, the identification of this sample to the family level of Gadidae was considered  
271 sufficient in the current study to confirm that the species on the label was different from the  
272 identified species (discussed in section 4.1.1).

## 273 **4. Discussion**

### 274 **4.1. Species Substitution**

275 Species substitution was detected in 24 of the 103 raw, RTE seafood samples identified  
276 in this study (Table 1). These results include six samples that were listed as “fish” or “white fish”  
277 on the menu, but were verbally declared by staff to be halibut, sea bass, tilapia, or snapper. While  
278 several cases of species substitution were associated with economic motivation (discussed in the  
279 following subsections), instances of substitution resulting from verbal declarations of restaurant  
280 staff may have been due to a lack of communication among restaurant staff and managers  
281 regarding fish species. Among the 67 restaurants sampled in this study, 20 had at least one case  
282 of species substitution. Interestingly, the species substitution rate found in this study for raw,  
283 RTE seafood (23.3%) is higher than rates of species substitution reported previously in the  
284 Southern California region for fresh or thawed fish fillets (13.3%) sold at grocery stores (Liou et  
285 al., 2020) and pre-packaged frozen fish fillets (1.8%) sold at grocery stores (Peterson et al.,  
286 2021). When the results were separated based on product category (Fig. 2), the highest rate of  
287 species substitution was observed with sushi samples at 28.6% (10/35), followed by ceviche  
288 samples at 23.5% (8/34), and then poke samples at 17.6% (6/34). These differences were not  
289 statistically significant ( $p > 0.017$ ), according to Fisher’s exact test of independence with the  
290 Bonferroni correction.

291 Products declared to be yellowtail were the most frequently detected targets of  
292 substitution (11/12), followed by those declared (verbally or on the menu) to be halibut (4/5),  
293 various types of snappers (3/3), tilapia (2/6), sea bass (2/2), and amberjack (1/1). Previous  
294 studies conducted in the United States and globally also reported relatively high rates of  
295 substitution for sea bass, yellowtail, snapper and halibut (Filonzi et al., 2010; Hanner et al., 2011;  
296 Hu et al., 2018; Liou et al., 2020; Pardo et al., 2016; Warner et al., 2013; Willette et al., 2017).

#### 297 **4.1.1. Yellowtail and Halibut Substitution**

298        Among the 12 samples labeled as yellowtail (*Seriola lalandi*), 11 were identified as buri  
299 (*Seriola quinqueradiata*). While these species share amberjack as an acceptable market name,  
300 yellowtail is not an acceptable market name for buri (FDA, 2022). This form of species  
301 substitution was observed across all three product categories (ceviche, sushi, and poke). The high  
302 rate of substitution found in yellowtail (91.7%) is consistent with Warner et al. (2013), who  
303 reported that 100% of sushi samples collected nationwide labeled as yellowtail or hamachi were  
304 actually buri (n = 23). While this particular species substitution is common, it is believed to be  
305 unintentional and possibly due to issues in foreign name translation (Warner et al., 2013).  
306 However, the detection of this substitution event in ceviche and poke is a novel finding that  
307 indicates this is a pervasive labeling error that is not limited to sushi dishes. An additional  
308 substitution event associated with the *Seriola* genus was the substitution of amberjack (*Seriola*  
309 sp.) for white trevally (*Pseudocaranx dentex*) in a sample of sushi. This substitution may have  
310 been accidental or due to a lack of product supply, as white trevally has a high commercial value  
311 and is utilized in sushi dishes (Agawa et al., 2022). Similarly, Wallstrom et al. (2020) previously  
312 reported the mislabeling of white trevally as another species of jack in a sushi sample.

313        Of the five dishes declared to be halibut in this study, only one was verified as halibut,  
314 with an identification of Pacific halibut (*Hippoglossus stenolepis*) for a ceviche dish. The  
315 remaining samples were found to be substituted with other species, specifically two sushi  
316 samples were identified as olive flounder (*Paralichthys olivaceus*), one sushi sample was  
317 identified as almaco jack (*Seriola rivoliana*), and one ceviche sample showed equivalent top  
318 matches to walleye pollock (*Gadus chalcogrammus*) and Arctic cod (*Boreogadus saida*). Halibut  
319 is a highly valued species and there are economic incentives to substituting it with other species.  
320 According to federal regulations (21 CFR §102.57), the term “halibut” is reserved for Atlantic



321 halibut (*Hippoglossus hippoglossus*) and Pacific halibut (*Hippoglossus stenolepis*). However, the  
322 findings of the current study combined with previous studies (Filonzi et al., 2010; Liou et al.,  
323 2020; Wallstrom et al., 2020; Warner et al., 2013; Willette et al., 2017), indicate that halibut  
324 substitution is a consistent problem, with flounders often mislabeled as halibut in sushi products.  
325 Consistent with the current study, Willette et al. (2017) also reported the mislabeling of olive  
326 flounder sushi as halibut. The use of undeclared raw olive flounder is problematic because this  
327 species has been associated with a myxosporean parasite, *Kudoa septempunctata*, which causes  
328 gastroenteritis in humans (Kawai et al., 2012).

#### 329 **4.1.2. Snapper, Sea Bass, and Tilapia Substitution**

330 Two instances of substitution involving snappers were detected in sushi products: red  
331 snapper (*Lutjanus campechanus*) substituted with tilapia (*Oreochromis* sp.) and black snapper  
332 (*Apsilus dentatus*) substituted with gilthead bream (*Sparus aurata*). These results are similar to  
333 snapper substitution events reported in Warner et al. (2013), in which snapper sushi was  
334 substituted with tilapia and bream, among other species. Similarly, studies conducted in Canada  
335 have reported a high frequency of red snapper/snapper substituted with tilapia (Hanner et al.,  
336 2011; Hu et al., 2018). The substitution of red snapper sushi with tilapia is likely economically  
337 motivated, as red snapper is a highly valued species while tilapia is generally inexpensive (Hu et  
338 al., 2018; Isaacs & Hellberg, 2020; Khaksar et al., 2015; Willette et al., 2017). Bream is often  
339 referred to as genuine snapper in sushi culture, and there does not appear to be an economic  
340 motivation for mislabeling it as black snapper (Hu et al., 2018). One sample of ceviche was  
341 labeled on the menu as fish and declared by staff to be snapper but was identified as catfish.

342 Two ceviche samples declared to be sea bass in this study were instead identified as  
343 Sutchi catfish (*Pangasianodon hypophthalmus*) and tilapia (*Oreochromis* sp.). *The Seafood List*

344 includes 16 species listed that are acceptable to be marketed as sea bass, none of which are from  
345 the *Oreochromis* or *Pangasianodon* genera (FDA, 2022). Sea bass is a highly valued species that  
346 is frequently substituted with other species, as reported in previous studies conducted in the  
347 United States and globally (Hanner et al., 2011; Hu et al., 2018; Khaksar et al., 2015; Velez-  
348 Zuazo et al., 2021; Warner et al., 2013). The ceviche sample identified as Sutchi catfish was  
349 described as “fish” on the menu and verbally declared to be “sea bass” by the staff, while the  
350 ceviche sample identified as tilapia specifically declared “sea bass” on the menu.

351         Of the three substituted ceviche samples declared to be tilapia, two were identified as  
352 whiteleg shrimp (*Litopanaeus vannamei*) and the other was identified as Sutchi catfish  
353 (*Pangasianodon hypophthalmus*). These samples were described as “fish” or “white fish” on the  
354 menu but declared by restaurant staff to be “tilapia”. Species substitution in products labeled as  
355 tilapia is not a common occurrence, as tilapia is a relatively low-value species. Of note, both of  
356 the restaurants that sold undeclared shrimp also had “shrimp ceviche” dishes on their menus.  
357 Although these substitution events appear to be accidental, individuals with shellfish allergies  
358 could be greatly impacted by the undeclared presence of shrimp due to the potential for  
359 anaphylactic shock. These substitutions also have cultural impacts, as the consumption of  
360 shellfish is prohibited in those with kosher diets.

#### 361 **4.2. Unacceptable market names**

362         Use of the common name or acceptable market name(s) for a fish species as provided in  
363 *The Seafood List* is recommended by the FDA in order to provide a clear representation of a  
364 product to the consumer (FDA, 2022). Of the 103 samples identified with DNA barcoding, 47  
365 (45.6%) did not have common names or acceptable market names on the menu (Table 2).  
366 Among the 67 restaurants sampled in this study, 43 had at least one unacceptable market name

367 on the menu. Six of the products with unacceptable market names were also determined to be  
368 substituted based on the verbal declarations of restaurant staff (discussed above). Among the  
369 three product categories (Fig. 2), ceviche had the greatest rate of unacceptable market names at  
370 79.4% (27/34), followed by poke at 44.1% (15/34), and sushi at 14.3% (5/35). These three rates  
371 were found to be significantly different from each other using Fisher's exact test of independence  
372 with the Bonferroni correction ( $p < 0.017$ ). However, there were no significant differences in the  
373 rates of unacceptable market names based on price within each product category (Kruskal Wallis  
374 H test,  $p > 0.05$ ) or when comparing independent vs. chain restaurants (Fisher's exact test,  $p >$   
375 0.05).

#### 376 **4.2.1. Ceviche**

377 Ceviche's high mislabeling rate was driven primarily by restaurants referring to their  
378 product using generic terms such as "fish" ( $n = 22$ ), "pescado" ( $n = 4$ ), or "white fish" ( $n = 2$ ),  
379 none of which are acceptable market names for fish sold in the United States. These generically  
380 labeled samples were identified as a wide variety of species, including Atlantic salmon, tilapia,  
381 Sutchi catfish, whiteleg shrimp, yellowtail, and widow rockfish. When the restaurant staff were  
382 asked for clarification regarding the species in the dish, their answers were uninformative for  
383 67.9% of samples and resulted in species substitution for 21.4% of samples (discussed above).  
384 However, in three instances (10.7%), the restaurant staff were able to provide an accurate market  
385 name (swai or tilapia) that was confirmed with DNA barcoding. These results are similar to a  
386 study conducted in Chile, which indicated that 64% of salespeople were unable to identify the  
387 species in sushi products (Prida et al., 2020). In comparison, a U.S. study on tuna sushi reported  
388 that 32% of species descriptions declared by restaurant staff were incorrect and 9% were  
389 uninformative for generically labeled tuna sushi samples (Lowenstein et al., 2009).

#### 390 4.2.2. Poke and Sushi

391 Unacceptable market names among poke products were driven by incorrectly labeled  
392 tuna and salmon samples. Four poke samples labeled as “ahi” or “ahi tuna” were identified as  
393 yellowfin tuna (*Thunnus albacares*) and one was identified as southern bluefin tuna (*Thunnus*  
394 *maccoyii*). The term “ahi poke” is often used to describe poke containing yellowfin tuna or  
395 bigeye tuna (Hospital & Beavers, 2014). However, “ahi” is considered a vernacular name and is  
396 not an acceptable market name for any species according to *The Seafood List* (FDA, 2022). The  
397 mislabeling of southern bluefin tuna with a vernacular term for yellowfin or bigeye tuna is  
398 potentially a case of “reverse substitution”, in which the highly valuable bluefin tuna is illegally  
399 overfished and sold as a lesser species (Gordoa et al., 2017). Reverse substitution involving  
400 bluefin tuna has been reported previously in studies conducted in the United States (Liou et al.,  
401 2020) and Spain (Gordoa et al., 2017).

402 Ten poke samples and four sushi samples with unacceptable market names were  
403 associated with improper labeling of salmon. These samples were labeled as “salmon” and  
404 identified as Atlantic salmon (*Salmo salar*). While labeling of Atlantic salmon generically as  
405 “salmon” is a commonly observed practice (Khaksar et al., 2015; Liou et al., 2020), it should be  
406 referred to as “Atlantic salmon” according to *The Seafood List* (FDA, 2022). The specification is  
407 important because Atlantic salmon is a farmed species, whereas the majority of Pacific salmon  
408 are wild-caught (Cline, 2012). Therefore, designating Atlantic salmon generically as “salmon”  
409 does not provide consumers with sufficient information to make informed purchasing decisions.  
410 The remaining sushi sample with an unacceptable market name was labeled as “izumidai” and  
411 identified as tilapia. While izumidai is used in sushi culture to refer to tilapia, it is not a  
412 recommended market name according to *The Seafood List* (FDA, 2022).

### 413 4.3. Overall mislabeling rates

414 When the instances of species substitution and use of unacceptable market names were  
415 combined, the overall mislabeling rate was found to be 63.1% (65/103; Fig. 2). Among the 67  
416 restaurants where samples were collected, 55 had at least one instance of species substitution or  
417 use of unacceptable market names. The overall mislabeling rate reported here for raw, RTE  
418 seafood (63.1%) is similar to the rate of 61% reported by Velez-Zuazo et al. (2021) for  
419 mislabeling at ceviche and sushi restaurants in Peru, and within the range of previous studies  
420 conducted globally on sushi which have reported 12-84% species mislabeling (Gordoa et al.,  
421 2017; Hu et al., 2018; Khaksar et al., 2015; Liu et al., 2022; Lowenstein et al., 2009; Prida et al.,  
422 2020; Wallstrom et al., 2020; Warner et al., 2013; Willette et al., 2017). However, it is difficult  
423 to make direct comparisons among mislabeling rates reported in other studies due to differences  
424 in sampling methods and the way in which mislabeling is determined (e.g., species substitution  
425 versus unacceptable market names).

426 When the overall mislabeling rates were separated based on product category, the rate for  
427 ceviche (85.3%, 29/34) was significantly higher ( $p < 0.017$ ) than the rate for sushi (42.9%,  
428 15/35), according to a Fisher's exact test of independence with the Bonferroni correction. On the  
429 other hand, the overall rate of poke mislabeling (61.8%, 21/34) was not significantly different  
430 from the rates for ceviche or sushi mislabeling. Mislabeling in ceviche and poke was primarily  
431 due to the use of unacceptable market names, whereas species substitution was more common in  
432 sushi dishes (Fig. 2). This is likely due to differences in how the products are marketed to  
433 consumers – ceviche dishes are often generically labeled as “fish” and poke restaurants tend to  
434 offer a small range of fish choices (i.e., yellowtail, tuna, and salmon). On the other hand, sushi

435 restaurants tend to offer a larger range of fish choices at various price points depending on the  
436 fish species listed on the menu.

## 437 **5. Conclusion**

438 This study highlights several challenges with species substitution and mislabeling of raw,  
439 RTE seafood. Ceviche mislabeling is not well documented in the United States, and the results of  
440 this study indicate a relatively high mislabeling rate, primarily due to the use of unacceptable  
441 market names. There were six instances where the species that was verbally declared by  
442 restaurant staff resulted in cases of species substitution. These occasions speak to the challenges  
443 of relying on adequate communication among restaurant staff and managers regarding fish  
444 species and suggest potential for improvement in this area. Two instances of shellfish being  
445 present in samples sold as finfish present concerns for public safety. While the substitution was  
446 likely accidental, ingestion of shellfish in those with allergies can result in anaphylactic shock,  
447 leading to hospitalization or death. This study presents the first reports of poke mislabeling,  
448 which was due to a combination of unacceptable market names and species substitution. In  
449 combination with previous studies, the current study demonstrates that sushi species substitution  
450 continues to be problematic, with yellowtail, halibut, sea bass, and snapper samples being  
451 consistently mislabeled. While mislabeled products were observed at the retail level in this study,  
452 it is likely that a portion of these products were originally mislabeled earlier in the supply chain.  
453 Further research into mislabeling within the seafood supply chain and a determination of where  
454 and why mislabeling is occurring is warranted. Additionally, greater outreach efforts at the retail  
455 level to improve menu descriptions of raw, RTE seafood are suggested, as well as requiring the  
456 use of species names or acceptable market names according to *The Seafood List*.

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**Table 1.** Instances of species substitution (n = 24) in raw, RTE seafood samples based on menu description or verbal declaration.

Menu Description	Expected Species <sup>a</sup>	Identified Species	Product Category	No. of Samples
Yellowtail	Yellowtail ( <i>Seriola lalandi</i> )	Buri ( <i>Seriola quinqueradiata</i> )	Sushi	4
			Poke	6
			Ceviche	1
Halibut	Atlantic halibut ( <i>Hippoglossus hippoglossus</i> ) or Pacific halibut ( <i>Hippoglossus stenolepis</i> )	Olive flounder ( <i>Paralichthys olivaceus</i> )	Sushi	2
		Almaco jack ( <i>Seriola rivoliana</i> )	Sushi	1
Fish (halibut <sup>b</sup> )	Atlantic halibut ( <i>Hippoglossus hippoglossus</i> ) or Pacific halibut ( <i>Hippoglossus stenolepis</i> )	Walleye Pollock ( <i>Gadus chalcogrammus</i> )/Arctic cod ( <i>Boreogadus saida</i> )	Ceviche	1
Sea bass	Sea bass (range of species from 6 genera)	Tilapia ( <i>Oreochromis sp.</i> )	Ceviche	1
Fish (sea bass <sup>b</sup> )	Sea bass (range of species from 6 genera)	Sutchi Catfish ( <i>Pangasianodon hypophthalmus</i> )	Ceviche	1
Red snapper	Red snapper ( <i>Lutjanus campechanus</i> )	Tilapia ( <i>Oreochromis sp.</i> )	Sushi	1
Black snapper	Black snapper ( <i>Apsilus dentatus</i> )	Gilthead bream ( <i>Sparus aurata</i> )	Sushi	1
Fish (snapper <sup>b</sup> )	Snapper (range of species from 12 genera)	Sutchi Catfish ( <i>Pangasianodon hypophthalmus</i> )	Ceviche	1
Amberjack	Amberjack ( <i>Seriola sp.</i> )	White trevally ( <i>Pseudocaranx dentex</i> )	Sushi	1
White fish (tilapia <sup>b</sup> )	Tilapia ( <i>Oreochromis sp.</i> , <i>Sarotherodon sp.</i> or <i>Tilapia sp.</i> )	Sutchi Catfish ( <i>Pangasianodon hypophthalmus</i> )	Ceviche	1
Fish (tilapia <sup>b</sup> )	Tilapia ( <i>Oreochromis sp.</i> , <i>Sarotherodon sp.</i> or <i>Tilapia sp.</i> )	Whiteleg Shrimp ( <i>Panaeus vannamei</i> )	Ceviche	2

<sup>a</sup>Expected species were determined based on acceptable market names listed in *The Seafood List* (FDA, 2022)<sup>b</sup>Verbal declaration by restaurant staff

**Table 2.** Samples found to have unacceptable market names (n = 47) according to *The Seafood List* (FDA, 2022). Note: The common name may be used as the market name unless prohibited by regulation or law.

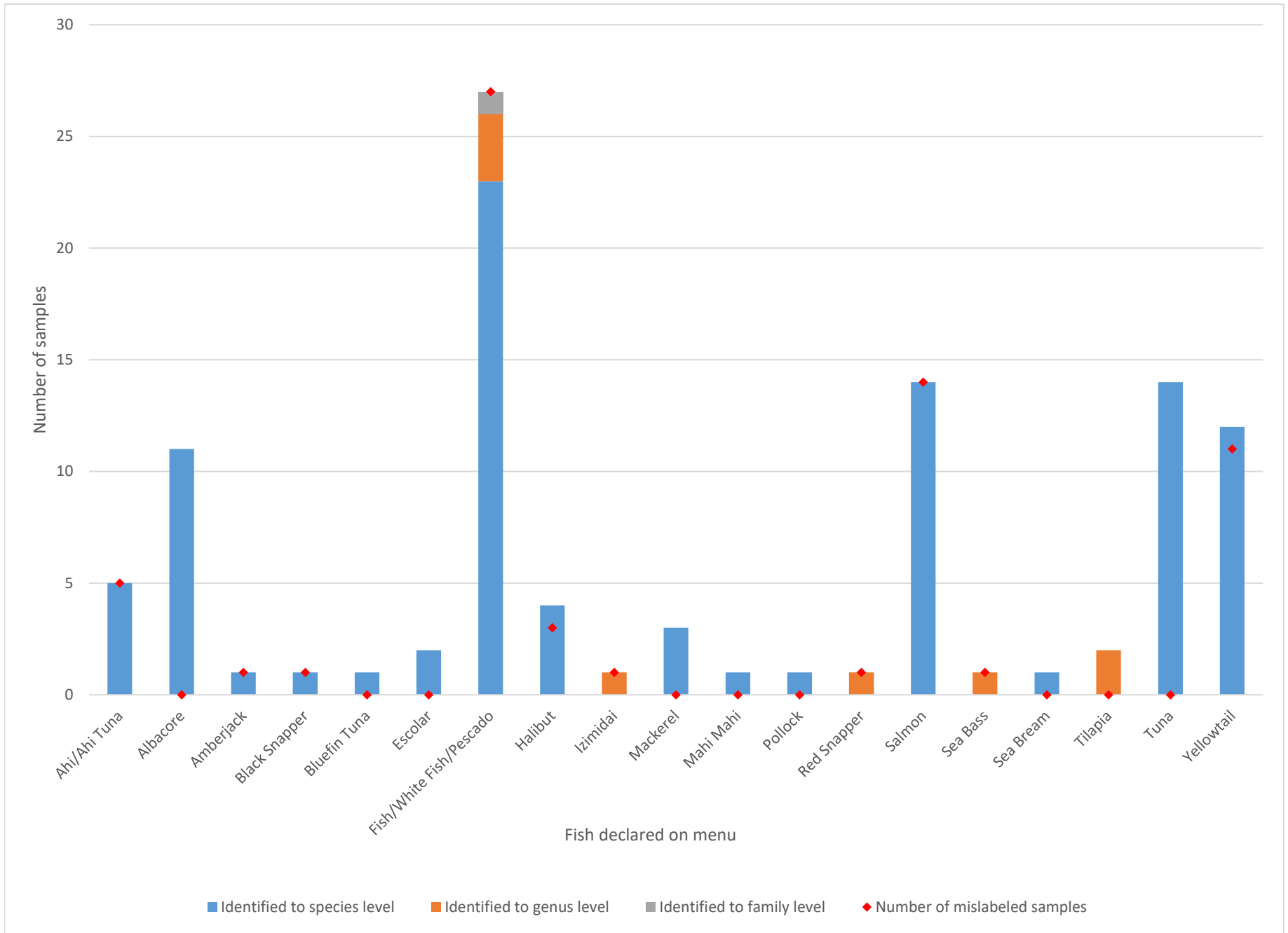
<b>Product Category</b>	<b>Menu Description</b>	<b>Number of Samples</b>	<b>Identified Species</b>	<b>Acceptable market name(s)</b>
<b>Ceviche</b>	Fish/white	15	Sutchi catfish ( <i>Pangasianodon hypophthalmus</i> )	Swai, Sutchi, Striped Pangasius, or Tra
	Fish/pescado	5	Tilapia ( <i>Oreochromis sp.</i> )	Tilapia
	Fish	1	Walleye pollock ( <i>Gadus chalcogrammus</i> )	Pollock
	Fish	1	Walleye pollock ( <i>Gadus chalcogrammus</i> )/Artic cod ( <i>Boreogadus saida</i> )	Pollock/Cod
	Fish	1	Atlantic salmon ( <i>Salmo salar</i> )	Atlantic salmon
	Fish	1	Yellowtail ( <i>Seriola lalandi</i> )	Amberjack or Yellowtail
	Fish	1	Widow rockfish ( <i>Sebastes entomelas</i> )	Rockfish
	Fish	2	Whiteleg shrimp ( <i>Litopenaeus vannamei</i> )	Shrimp
	<b>Poke</b>	Salmon	10	Atlantic salmon ( <i>Salmo salar</i> )
Ahi/ahi tuna		4	Yellowfin tuna ( <i>Thunnus albacares</i> )	Tuna
Ahi tuna		1	Southern Bluefin Tuna ( <i>Thunnus maccoyii</i> )	Tuna
<b>Sushi</b>	Salmon	4	Atlantic salmon ( <i>Salmo salar</i> )	Atlantic salmon
	Izimidai	1	Tilapia ( <i>Oreochromis sp.</i> )	Tilapia

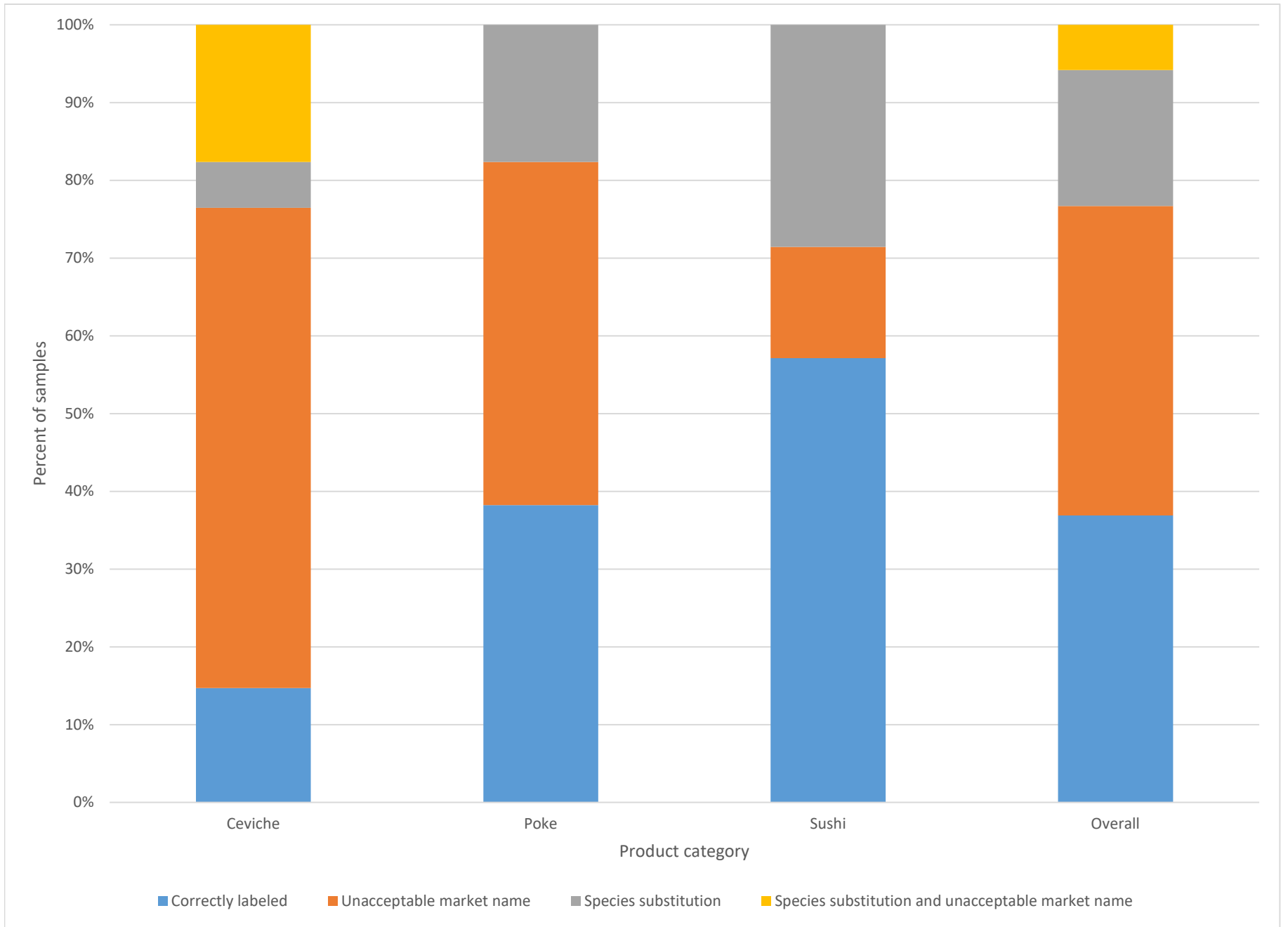


## **Figure captions**

**Figure 1.** Combined results of DNA barcoding and mini-barcoding for raw, RTE seafood samples identified in this study (n = 103). The number of mislabeled samples refers those with unacceptable market names and/or species substitution.

**Figure 2.** Proportion of ceviche (n = 34), poke (n = 34), and sushi (n = 35) samples with unacceptable market names and/or species substitution. The “species substitution and unacceptable market name” category applies to samples that had an unacceptable market name on the menu and were substituted due to a verbal declaration by restaurant staff. The “overall” category represents all samples combined (n = 103).

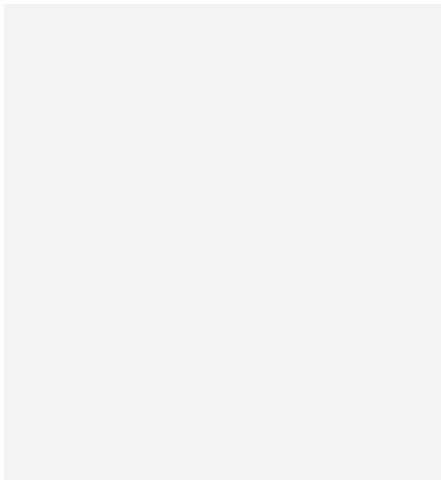




Number for sorting	Sample Number	Product category	Menu Declaration	Verbal Declaration by Staff
1	Q1	Sushi/sashimi	Salmon	N/A
2	Q2	Sushi/sashimi	Tuna	N/A
3	Q3	Sushi/sashimi	Salmon	N/A
4	Q4	Sushi/sashimi	Tuna	N/A
5	Q5	Poke	Tuna	N/A
6	Q6	Poke	Yellowtail	N/A
7	Q7	Poke	Tuna	N/A
8	Q8	Poke	Salmon	N/A
9	Q9	Sushi/sashimi	Tuna	N/A
10	Q10	Sushi/sashimi	Yellowtail	N/A
11	Q11	Sushi/sashimi	Yellowtail	N/A
12	Q12	Sushi/sashimi	Tuna	N/A
13	Q13	Sushi/sashimi	Tuna	N/A
14	Q14	Sushi/sashimi	Salmon	N/A
15	Q15	Sushi/sashimi	Tuna	N/A
16	Q16	Sushi/sashimi	Yellowtail	N/A
17	Q17	Poke	Albacore	N/A
18	Q18	Poke	Salmon	N/A
19	Q19	Sushi/sashimi	Albacore	N/A
20	Q20	Sushi/sashimi	Yellowtail	N/A
21	Q21	Sushi/sashimi	Tuna	N/A
22	Q22	Sushi/sashimi	Salmon	N/A
23	Q23	Sushi/sashimi	Tuna	N/A
24	Q24	Sushi/sashimi	Albacore	N/A
25	Q25	Poke	Salmon	N/A
26	Q26	Poke	Tuna	N/A
27	Q27	Poke	Salmon	N/A
28	Q28	Poke	Ahi tuna	N/A
29	Q29	Poke	Salmon	N/A
30	Q30	Poke	Ahi	N/A
31	Q31	Ceviche	Fish	No additional informa
32	Q32	Poke	Tuna	N/A
33	Q33	Poke	Salmon	N/A
34	Q34	Poke	Tuna	N/A
35	Q35	Poke	Salmon	N/A
36	Q36	Sushi/sashimi	Albacore	N/A
37	Q37	Sushi/sashimi	Mackerel	N/A
38	Q38	Sushi/sashimi	Mackerel	N/A
39	Q39	Sushi/sashimi	Escolar	N/A
40	Q40	Sushi/sashimi	Halibut	N/A
41	Q41	Sushi/sashimi	Red snapper	N/A
42	Q42	Poke	Salmon	N/A
43	Q43	Poke	Yellowtail	N/A
45	Q45	Poke	Salmon	N/A
46	Q46	Poke	Yellowtail	N/A
47	Q47	Poke	Salmon	N/A

48 Q48	Poke	Tuna	N/A
49 Q49	Sushi/sashimi	Albacore	N/A
50 Q50	Sushi/sashimi	Izimidai	N/A
51 Q51	Sushi/sashimi	Mackerel	N/A
52 Q52	Sushi/sashimi	Albacore	N/A
53 Q53	Sushi/sashimi	Halibut	N/A
54 Q54	Sushi/sashimi	Black snapper	N/A
55 Q55	Sushi/sashimi	Amberjack	N/A
56 Q56	Sushi/sashimi	Escolar	N/A
57 Q57	Sushi/sashimi	Albacore	N/A
58 Q58	Sushi/Sashimi	Halibut	N/A
59 Q59	Sushi/Sashimi	Sea bream	N/A
60 Q60	Ceviche	Pescado	No additional informa
61 Q61	Ceviche	Mahi mahi	N/A
63 Q63	Ceviche	Fish	No additional informa
64 Q64	Ceviche	Fish	No additional informa
65 Q65	Ceviche	White fish	No additional informa
66 Q66	Ceviche	Fish	No additional informa
68 Q68	Ceviche	Sea bass	N/A
69 Q69	Ceviche	Pescado	No additional informa
71 Q71	Ceviche	Fish	Halibut
72 Q72	Ceviche	Fish	Tilapia
73 Q73	Ceviche	Halibut	N/A
75 Q75	Ceviche	Pescado	No additional informa
77 Q77	Ceviche	Fish	Swai
79 Q79	Ceviche	Pollock	N/A
80 Q80	Ceviche	Tilapia	N/A
83 Q83	Ceviche	Fish	White fish
84 Q84	Ceviche	Fish	White fish
86 Q86	Ceviche	Fish	Sea bass
88 Q88	Ceviche	Fish	White fish
89 Q89	Ceviche	Fish	White fish
90 Q90	Ceviche	Yellowtail	N/A
95 Q95	Ceviche	Fish	Tilapia
96 Q96	Ceviche	Fish	Tilapia
98 Q98	Ceviche	Fish	Tilapia
99 Q99	Ceviche	Fish	Snapper
101 Q101	Poke	Ahi Tuna	N/A
103 Q103	Poke	Ahi Tuna	N/A
106 Q106	Poke	Albacore	N/A
107 Q107	poke	Yellowtail	N/A
111 Q111	Poke	Albacore	N/A
114 Q114	Poke	Albacore	N/A
123 Q123	Poke	Albacore	N/A
127 Q127	Poke	Ahi tuna	N/A
133 Q133	Poke	Albacore	N/A
134 Q134	Poke	Bluefin tuna	N/A
145 Q145	Ceviche	Fish	No additional informa
146 Q146	Ceviche	Pescado	No additional informa

184 Q184	Ceviche	White Fish	Tilapia
186 Q186	Ceviche	Fish	No additional informa
187 Q187	Ceviche	Fish	No additional informa
188 Q188	Ceviche	Fish	No additional informa
189 Q189	Ceviche	Tilapia	N/A
190 Q190	Ceviche	Fish	No additional informa
191 Q191	Ceviche	Fish	No additional informa
193 Q193	Poke	Yellowtail	N/A
194 Q194	Poke	Yellowtail	N/A
195 Q195	Poke	Yellowtail	N/A













































Price (USD)	Type of Restaurant (Independent or Chain)	Barcode used for Species ID
\$10.45	Independent	Full Barcode
\$11.45	Independent	Full Barcode and CR
\$10.99	Independent	Full Barcode
\$11.99	Independent	Full Barcode and CR
\$9.99	Independent	Full Barcode and CR
\$9.99	Independent	Full Barcode
\$12.99	Independent	Full Barcode and CR
\$12.99	Independent	Full Barcode
\$15.95	Independent	Full Barcode and CR
\$15.95	Independent	Full Barcode
\$14.95	Independent	Full Barcode
\$14.95	Independent	Full Barcode and CR
\$3.99	Independent	Full Barcode and CR
\$3.99	Independent	COI Mini-Barcode
\$3.79	Independent	CR (full barcode identified genus)
\$4.29	Independent	Full Barcode
\$12.99	Chain	CR (full barcode identified genus)
\$12.99	Chain	Full Barcode
\$7.99	Chain	CR (full barcode identified genus)
\$7.99	Chain	Full Barcode
\$12.95	Independent	CR (full barcode identified genus)
\$12.95	Independent	Full Barcode
\$6.99	Independent	Full Barcode and CR
\$6.99	Independent	CR (full barcode identified genus)
\$13.95	Chain	Full Barcode
\$13.95	Chain	CR (full barcode identified genus)
\$13.95	Chain	Full Barcode
\$13.95	Chain	CR (full barcode identified genus)
\$14.00	Independent	Full Barcode
\$14.00	Independent	Full Barcode and CR
\$15.34	Independent	Full Barcode
\$9.99	Independent	CR (full barcode identified genus)
\$9.99	Independent	Full Barcode
\$13.95	Independent	CR (full barcode identified genus)
\$13.95	Independent	Full Barcode
\$15.95	Independent	CR (full barcode identified genus)
\$13.99	Independent	Full Barcode
\$15.95	Independent	COI Mini-Barcode
\$15.95	Independent	Full Barcode
\$6.95	Independent	Full Barcode
\$4.95	Independent	Full Barcode
\$8.95	Independent	Full Barcode
\$8.95	Independent	Full Barcode
\$8.99	Independent	COI Mini-Barcode
\$8.99	Independent	Full Barcode
\$11.75	Chain	Full Barcode

\$11.75 Chain	Full Barcode and CR
\$3.99 Independent	CR (full barcode identified genus)
\$3.99 Independent	Full Barcode
\$3.69 Independent	COI Mini-Barcode
\$3.69 Independent	Full Barcode and CR
\$8.00 Independent	Full Barcode
\$8.00 Independent	Full Barcode
\$8.00 Independent	Full Barcode
\$7.00 Independent	Full Barcode
\$7.00 Independent	CR
\$16.00 Independent	Full Barcode
\$18.00 Independent	Full Barcode
\$4.50 Independent	Full Barcode
\$18.00 Independent	Full Barcode
\$16.50 Independent	Full Barcode
\$4.35 Chain	Full Barcode
\$17.95 Independent	Full Barcode
\$5.45 Independent	COI Mini-Barcode
\$14.50 Independent	Full Barcode
\$8.49 Chain	Full Barcode
\$10.39 Independent	COI Mini-Barcode
\$17.50 Chain	Full Barcode
\$19.00 Chain	Full Barcode
N/A - missing data Independent	Full Barcode
\$13.99 Independent	Full Barcode
\$15.00 Independent	Full Barcode
\$8.00 Chain	Full Barcode
N/A - missing data Independent	Full Barcode
\$9.14 Independent	Full Barcode
\$14.95 Independent	Full Barcode
N/A - missing data Independent	Full Barcode
\$14.00 Independent	Full Barcode
\$17.00 Chain	Full Barcode
\$7.00 Independent	COI Mini-Barcode
\$7.99 Independent	COI Mini-Barcode
\$14.46 Independent	Full Barcode
\$15.00 Independent	Full Barcode
\$11.99 Chain	CR (Full barcode identified genus)
\$11.85 Independent	CR (Full barcode identified genus)
\$8.95 Independent	N/A (did not get an ID with DNA barcoding)
\$8.95 Independent	Full Barcode
\$8.95 Independent	Full Barcode and CR
\$9.99 Independent	CR
\$13.99 Chain	Full Barcode and CR
\$9.95 Independent	Full Barcode and CR
\$13.00 Independent	CR (Full barcode identified genus)
\$12.35 Chain	Full Barcode and CR
\$15.00 Chain	COI Mini-Barcode
\$11.95 Independent	Full Barcode

\$19.00 Chain	Full Barcode
\$16.50 Independent	Full Barcode
\$13.00 Independent	Full Barcode
\$11.25 Independent	Full Barcode
\$9.99 Independent	Full Barcode
\$10.99 Chain	N/A (did not get an ID with DNA barcoding)
\$9.99 Independent	Full Barcode
\$10.19 Independent	Full Barcode
\$10.50 Chain	Full Barcode
\$10.50 Chain	COI Mini-Barcode









































Species ID	Level of Genetic Match	Labeling Status
Atlantic salmon ( <i>Salmo salar</i> )	Species	Unacceptable Market Name
Bigeye tuna ( <i>Thunnus obesus</i> )	Species	Correctly Labeled
Atlantic salmon ( <i>Salmo salar</i> )	Species	Unacceptable Market Name
Southern bluefin tuna ( <i>Thunnus maccoyii</i> )	Species	Correctly Labeled
Southern bluefin tuna ( <i>Thunnus maccoyii</i> )	Species	Correctly Labeled
Yellowtail ( <i>Seriola lalandi</i> )	Species	Correctly Labeled
Southern bluefin tuna ( <i>Thunnus maccoyii</i> )	Species	Correctly Labeled
Atlantic salmon ( <i>Salmo salar</i> )	Species	Unacceptable Market Name
Bigeye tuna ( <i>Thunnus obesus</i> )	Species	Correctly Labeled
Buri ( <i>Seriola quinqueradiata</i> )	Species	Species substitution
Buri ( <i>Seriola quinqueradiata</i> )	Species	Species substitution
Bigeye tuna ( <i>Thunnus obesus</i> )	Species	Correctly Labeled
Bigeye tuna ( <i>Thunnus obesus</i> )	Species	Correctly Labeled
Atlantic salmon ( <i>Salmo salar</i> )	Species	Unacceptable Market Name
Yellowfin tuna ( <i>Thunnus albacares</i> )	Species	Correctly Labeled
Buri ( <i>Seriola quinqueradiata</i> )	Species	Species substitution
Albacore ( <i>Thunnus alalunga</i> )	Species	Correctly Labeled
Atlantic salmon ( <i>Salmo salar</i> )	Species	Unacceptable Market Name
Albacore tuna ( <i>Thunnus alalunga</i> )	Species	Correctly Labeled
Buri ( <i>Seriola quinqueradiata</i> )	Species	Species substitution
Yellowfin tuna ( <i>Thunnus albacares</i> )	Species	Correctly Labeled
Atlantic salmon ( <i>Salmo salar</i> )	Species	Unacceptable Market Name
Bigeye tuna ( <i>Thunnus obesus</i> )	Species	Correctly Labeled
Albacore ( <i>Thunnus alalunga</i> )	Species	Correctly Labeled
Atlantic salmon ( <i>Salmo salar</i> )	Species	Unacceptable Market Name
Yellowfin tuna ( <i>Thunnus albacares</i> )	Species	Correctly Labeled
Atlantic salmon ( <i>Salmo salar</i> )	Species	Unacceptable Market Name
Yellowfin tuna ( <i>Thunnus albacares</i> )	Species	Unacceptable Market Name
Atlantic salmon ( <i>Salmo salar</i> )	Species	Unacceptable Market Name
Yellowfin tuna ( <i>Thunnus albacares</i> )	Species	Unacceptable Market Name
Atlantic salmon ( <i>Salmo salar</i> )	Species	Unacceptable Market Name
Yellowfin tuna ( <i>Thunnus albacares</i> )	Species	Correctly Labeled
Atlantic salmon ( <i>Salmo salar</i> )	Species	Unacceptable Market Name
Yellowfin tuna ( <i>Thunnus albacares</i> )	Species	Correctly Labeled
Atlantic salmon ( <i>Salmo salar</i> )	Species	Unacceptable Market Name
Albacore ( <i>Thunnus alalunga</i> )	Species	Correctly Labeled
Atlantic mackerel ( <i>Scomber scombrus</i> )	Species	Correctly Labeled
Atlantic mackerel ( <i>Scomber scombrus</i> )	Species	Correctly Labeled
Escolar ( <i>Lepidocybium flavobrunneum</i> )	Species	Correctly Labeled
Almaco jack ( <i>Seriola rivoliana</i> )	Species	Species substitution
Tilapia ( <i>Oreochromis</i> sp.)	Genus	Species substitution
Atlantic salmon ( <i>Salmo salar</i> )	Species	Unacceptable Market Name
Buri ( <i>Seriola quinqueradiata</i> )	Species	Species substitution
Atlantic salmon ( <i>Salmo salar</i> )	Species	Unacceptable Market Name
Buri ( <i>Seriola quinqueradiata</i> )	Species	Species substitution
Atlantic salmon ( <i>Salmo salar</i> )	Species	Unacceptable Market Name

Yellowfin tuna ( <i>Thunnus albacares</i> )	Species	Correctly Labeled
Albacore ( <i>Thunnus alalunga</i> )	Species	Correctly Labeled
Tilapia ( <i>Oreochromis</i> sp.)	Genus	Unacceptable Market Name
Atlantic mackerel ( <i>Scomber scombrus</i> )	Species	Correctly Labeled
Albacore ( <i>Thunnus alalunga</i> )	Species	Correctly Labeled
Olive flounder ( <i>Paralichthys olivaceus</i> )	Species	Species Substitution
Gilthead bream ( <i>Sparus aurata</i> )	Species	Species Substitution
White trevally ( <i>Pseudocaranx dentex</i> )	Species	Species Substitution
Escolar ( <i>Lepidocybium flavobrunneum</i> )	Species	Correctly Labeled
Albacore ( <i>Thunnus alalunga</i> )	Species	Correctly Labeled
Olive flounder ( <i>Paralichthys olivaceus</i> )	Species	Species Substitution
Madai ( <i>Pagrus major</i> ) (also matched Nile tilap	Species	Correctly Labeled (sea bream is
Tilapia ( <i>Oreochromis</i> sp.)	Genus	Unacceptable Market Name
Dolphinfish ( <i>Coryphaena hippurus</i> )	Species	Correctly Labeled
Sutchi catfish ( <i>Pangasianodon hypophthalmus</i>	Species	Unacceptable Market Name
Sutchi catfish ( <i>Pangasianodon hypophthalmus</i>	Species	Unacceptable Market Name
Sutchi catfish ( <i>Pangasianodon hypophthalmus</i>	Species	Unacceptable Market Name
Walleye pollock ( <i>Gadus chalcogrammus</i> )	Species	Unacceptable Market Name
Tilapia ( <i>Oreochromis</i> sp.)	Genus	Species Substitution
Sutchi catfish ( <i>Pangasianodon hypophthalmus</i>	Species	Unacceptable Market Name
Walleye pollock ( <i>Gadus chalcogrammus</i> )/Artic	Family/Multi-Genus	Species Substitution (verbal) an
Mozambique tilapia ( <i>Oreochromis mossambic</i>	Species	Unacceptable Market Name (bu
Pacific halibut ( <i>Hippoglossus stenolepis</i> )	Species	Correctly Labeled
Nile tilapia ( <i>Oreochromis niloticus</i> )	Species	Unacceptable Market Name
Sutchi catfish ( <i>Pangasianodon hypophthalmus</i>	Species	Unacceptable Market Name (bu
Walleye pollock ( <i>Gadus chalcogrammus</i> )	Species	Correctly labeled
Tilapia ( <i>Oreochromis</i> sp.)	Genus	Correctly labeled
Sutchi catfish ( <i>Pangasianodon hypophthalmus</i>	Species	Unacceptable Market Name
Sutchi catfish ( <i>Pangasianodon hypophthalmus</i>	Species	Unacceptable Market Name
Sutchi catfish ( <i>Pangasianodon hypophthalmus</i>	Species	Species Substitution (verbal) an
Sutchi catfish ( <i>Pangasianodon hypophthalmus</i>	Species	Unacceptable Market Name
Sutchi catfish ( <i>Pangasianodon hypophthalmus</i>	Species	Unacceptable Market Name
Buri ( <i>Seriola quinqueradiata</i> )	Species	Species Substitution
Whiteleg shrimp ( <i>Penaeus vannamei</i> ) (synonyr	Species	Species Substitution and Unacco
Whiteleg shrimp ( <i>Penaeus vannamei</i> ) (synonyr	Species	Species Substitution and Unacco
Tilapia ( <i>Oreochromis</i> sp.)	Genus	Unacceptable Market Name
Sutchi catfish ( <i>Pangasianodon hypophthalmus</i>	Species	Species Substitution (verbal) an
Yellowfin tuna ( <i>Thunnus albacares</i> )	Species	Unacceptable Market Name
Yellowfin tuna ( <i>Thunnus albacares</i> )	Species	Unacceptable Market Name
N/A	N/A	N/A (not identified with barcod
Buri ( <i>Seriola quinqueradiata</i> )	Species	Species Substitution
Albacore ( <i>Thunnus alalunga</i> )	Species	Correctly Labeled
Albacore ( <i>Thunnus alalunga</i> )	Species	Correctly Labeled
Albacore ( <i>Thunnus alalunga</i> )	Species	Correctly Labeled
Southern Bluefin Tuna ( <i>Thunnus maccoyii</i> )	Species	Unacceptable Market Name
Albacore ( <i>Thunnus alalunga</i> )	Species	Correctly Labeled
Southern bluefin tuna ( <i>Thunnus maccoyii</i> )	Species	Correctly Labeled
Yellowtail ( <i>Seriola lalandi</i> )	Species	Unacceptable Market Name
Sutchi catfish ( <i>Pangasianodon hypophthalmus</i>	Species	Unacceptable Market Name

Sutchi catfish ( <i>Pangasianodon hypophthalmus</i> )	Species	Species Substitution (verbal) an
Sutchi catfish ( <i>Pangasianodon hypophthalmus</i> )	Species	Unacceptable Market Name
Tilapia ( <i>Oreochromis</i> sp.)	Genus	Unacceptable Market Name
Widow rockfish ( <i>Sebastes entomelas</i> )	Species	Unacceptable Market Name
Tilapia ( <i>Oreochromis</i> sp.)	Genus	Correctly Labeled
N/A	N/A	N/A (not identified with barcod
Sutchi catfish ( <i>Pangasianodon hypophthalmus</i> )	Species	Unacceptable Market Name
Buri ( <i>Seriola quinqueradiata</i> )	Species	Species substitution
Buri ( <i>Seriola quinqueradiata</i> )	Species	Species substitution
Buri ( <i>Seriola quinqueradiata</i> )	Species	Species substitution











































Full COI Barcode Consensus Sequence	COI (SH-E) Mini-Barcode Consensus Sequence	CR Mini-Barcode Consensus Sequence
CCTCTATTTAGTATTTGGTGCC N/A	N/A	N/A
CCTCTATCTAGTATTCGGTGCA N/A	N/A	ATATCTATATATAGACCATATACAGTAATGTTCTAGGACATATATC
CCTCTATTTAGTATTTGGTGCC N/A	N/A	N/A
CCTCTATCTAGTATTCGGTGCA N/A	N/A	ATATCTATATATCGACCATACATAATAATGCTTTAGGACATATATC
CCTCTATCTAGTATTCGGTGCA N/A	N/A	ATATTTATATATTGACCATATATAATAATGCTTTAGGACATATATC
CCTCTATCTAGTATTTGGTGCC N/A	N/A	N/A
CCTCTATCTAGTATTCGGTGCA N/A	N/A	ATATTTATATATTGACCATATATAATAATGCTTTAGGACATATATC
CCTCTATTTAGTATTTGGTGCC N/A	N/A	N/A
CCTCTATCTAGTATTCGGTGCA N/A	N/A	ATATTTATATATGAACCATATACAGTAATGTTCTAGGACATATATC
CCTCTATCTGGTATTCGGTGCC N/A	N/A	N/A
AACTTAGTCAACCCGGTGCTC N/A	N/A	N/A
CCTCTATCTAGTATTCGGTGCA N/A	N/A	ATATCTATATATGAACCATATACAATAATGTTCTAGGACATATATC
CCTCTATCTAGTATTCGGTGCA N/A	N/A	ATATTTATATATGGACCATATACAATAATGTTCTAGGACATATATC
Full barcode did not pass QC CCTCTATTTAGTATTTG	N/A	N/A
CCTTTATCTAGTATTCGGTGCA N/A	N/A	ATACTCATATATCGACCATATATAATAATGCTTTAGGACATATATC
CCTCTATCTGGTATTCGGTGCC N/A	N/A	N/A
CCTCTATCTAGTATTCGGTGCA N/A	N/A	ATAATCATATTTGAAACATATATAATAATGCTTTAGGACATATATC
CCTCTATTTAGTATTTGGTGCC N/A	N/A	N/A
TCTATCTAGTATTCGGTGCA N/A	N/A	ATAACTATATTTGAAACATATATAATAATGCTTTAGGACATACATC
CCTCTATCTGGTATTCGGTGCC N/A	N/A	N/A
CCTTTATCTAGTATTCGGTGCA N/A	N/A	ATACTCATATATCGACCATATATAATAATGCTTTAGGACATATATC
CCTCTATTTAGTATTTGGTGCC N/A	N/A	N/A
CCTCTATCTAGTATTCGGTGCA N/A	N/A	ATATCTATATATAGACCATATACAGTAATGTTCTAGGACATATATC
CCTCTATCTAGTATTCGGTGCA N/A	N/A	ATAACTATATTTGAAACATATATAATAATGCTTTAGGACATATATC
CCTCTATTTAGTATTTGGTGCC N/A	N/A	N/A
CCTTTATCTAGTATTCGGTGCA N/A	N/A	ATACTCATATATCGACCATATATAATAATGCTTTAGGACATATATC
CCTCTATTTAGTATTTGGTGCC N/A	N/A	N/A
CCTTTATCTAGTATTCGGTGCA N/A	N/A	ATACTCATATATCGACCATATATAATAATGCTTTAGGACATATATC
CCTCTATTTAGTATTTGGTGCC N/A	N/A	N/A
CCTTTATCTAGTATTCGGTGCA N/A	N/A	ATACTCATATATCGACCATATATAATAATGCTTTAGGACATATATC
CCTCTATTTAGTATTTGGTGCC N/A	N/A	N/A
CCTTTATCTAGTATTCGGTGCA N/A	N/A	ATACTCATATATCGACCATATATAATAATGCTTTAGGACATATATC
CCTCTATTTAGTATTTGGTGCC N/A	N/A	N/A
CCTTTATCTAGTATTCGGTGCA N/A	N/A	ATACTCATATATCGACCATATATAATAATGCTTTAGGACATATATC
CCTCTATTTAGTATTTGGTGCC N/A	N/A	N/A
CCTTTATCTAGTATTCGGTGCA N/A	N/A	ATACTCATATATCGACCATATATAATAATGCTTTAGGACATATATC
CCTCTATTTAGTATTTGGTGCC N/A	N/A	N/A
CCTTTATCTAGTATTCGGTGCA N/A	N/A	ATACTCATATATCGACCATATATAATAATGCTTTAGGACATATATC
CCTCTATTTAGTATTTGGTGCC N/A	N/A	N/A
TACGTAGTATTTGGTGCA N/A	N/A	N/A
Full barcode did not pass QC CCTCTACCTAGTATTTG	N/A	N/A
CCTGTATCTCGTATTCGGTGCA N/A	N/A	N/A
CCTTTATCTAGTATTTGGTGCC N/A	N/A	N/A
CCTCTATCTAGTATTTGGTGCT N/A	N/A	N/A
CCTCTATTTAGTATTTGGTGCC N/A	N/A	N/A
CCTCTATCTGGTATTCGGTGCC N/A	N/A	N/A
Full barcode did not pass QC CCTCTATTTAGTATTTG	N/A	N/A
CCTCTATCTGGTATTCGGTGCC N/A	N/A	N/A
CCTCTATTTAGTATTTGGTGCC N/A	N/A	N/A

CCTTTATCTAGTATTCGGTGCA N/A	ATACTCATATATCGACCATATATAATAATGCTTTAGGACATATATC
CCTCTATCTAGTATTCGGTGCA N/A	ATAACTATATTTGAAACATATATAATAATGCTTTAGGACATGTATC
CCTCTATCTAGTATTTGGTGCT N/A	N/A
Full barcode did not amplify i CCTCTACCTAGTATTTG	N/A
CCTCTATCTAGTATTCGGTGCA N/A	ATAACTATATTTCAAACATATATAATAATGCTTTAGGACATATATC
CCTCTATCTCGTATTTGGTGCC N/A	N/A
CCTTTATCTAGTATTTGGTGCT N/A	N/A
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CCTGTATCTCGTATTCGGTGCA N/A	N/A
Full barcode did not pass QC	N/A
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CCTTTATCTTGTATTTGGTGCT N/A	N/A
CCTCTATCTAGTATTTGGTGCT N/A	N/A
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CCTCTACCTAGTATTTGGTGCC N/A	N/A
CCTCTACCTAGTATTTGGTGCC N/A	N/A
CCTCTACCTAGTATTTGGTGCC N/A	N/A
Full barcode did not amplify i CCTTTATCTCGTATTTG	N/A
CCTCTATCTAGTATTTGGTGCT N/A	N/A
CCTCTACCTAGTATTTGGTGCC N/A	N/A
Full barcode did not pass QC CCTTTATCTCGTATTTG	N/A
CCTCTATCTAGTATTTGGTGCT N/A	N/A
CCTCTATCTCGTATTTGGTGCC N/A	N/A
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CCTCTACCTAGTATTTGGTGCC N/A	N/A
CCTCTACCTAGTATTTGGTGCC N/A	N/A
CCTCTATCTGGTATTCGGTGCC N/A	N/A
Full barcode did not pass QC ATTATACTTTATCTTCG	N/A
Full barcode did not pass QC ATTATACTTTATCTTCG	N/A
CCTCTATCTAGTATTTGGNTGC N/A	N/A
CCTCTACCTAGTATTTGGTGCC N/A	N/A
CCTTTATCTAGTATTCGGTGCA N/A	ATACTCATATATCGACCATATATAATAATGCTTTAGGACATATATC
CCTTTATCTAGTATTCGGTGCA N/A	CATACTCATATATCGACCATATATAATAATGCTTTAGGACATATA
Full barcode did not pass QC	CR mini-barcode did not pass QC
CCTCTATCTGGTATTCGGTGCC N/A	N/A
CCTCTATCTAGTATTCGGTGCA N/A	ATAACTATATTTAAAACATATATAATAATGCTTTAGGACATATATC
Full barcode did not pass QC	N/A
ATAACTATATTTAAAACATATATAATAATGCTTTAGGACATATATC	
CCTCTATCTAGTATTTGGTGCA N/A	ATAACTATATTTGAAACATATATAATAATGCTTTAGGACATATATC
CCTCTATCTAGTATTCGGTGCA N/A	ATACTTATATATCGACCATACATAATAATGCTTTAGGACATATATC
CCTCTATCTAGTATTCGGTGCA N/A	ATAACTATATTTTAAACATATATAATAATGCTTTAAGACATGTATC
CCTCTATCTAGTATTCGGTGCA N/A	ATACTTATATATCGACCATACATAATAATGCTTTAGGACATATATC
Full barcode did not pass QC CCTCTATCTAGTATTTG	N/A
CCTCTACCTAGTATTTGGTGCC N/A	N/A

CCTCTACCTAGTATTTGGTGCC N/A	N/A
CCTCTACCTAGTATTTGGTGCC N/A	N/A
CCTCTATCTAGTATTTGGTGCT N/A	N/A
CCTTTATCTAGTATTTGGTGCC N/A	N/A
CCTCTATCTAGTATTTGGTGCT N/A	N/A
Full barcode did not pass QC SH-E mini-barcode did	N/A
CCTCTACCTAGTATTTGGTGCC N/A	N/A
CCTCTATCTGGTATTCGGTGCC N/A	N/A
CCTCTATCTGGTATTCGGTGCC N/A	N/A
Full barcode did not amplify i CCTCTATCTGGTATTCG	N/A









































GTATTA AAA ACCATTACTAGTACTAA ACCATT CATATATCAACAAACAATGAAGACTTACATAA ACCCATACAGATATATTCC.

3TATTA AAA ACCATTACTAGTATTA ACCATT CATATGTCAACAAACAACGAAGATTTACATAA ACCCATGCAGATAAACTCC  
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3TATTA AAA ACCATTACTAGTATTA ACCATT CATATGTCAACAAACAATGAAGATTTACATAA ACCCATACAGATATATCTT/

3TATTA AAA ACCATTACTAGTACTAA ACCATT CACATGTCAACAAACAATGAAGATTTACATAA ACCCATACAGATATATCTT/

3TATTA AAA ACCATTACTAGTACTAA ACCATT CATATGTGCGAATAAACAATGAAAATTTACATAA ACCCATACAGATATATCT

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GTATTA AAA ACCATAACTAGTATTTA ACCATT CATATGTCAACATATCATGAAGACTTACATAA AGACATAACA ACCCATCTCCI



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3TATTAACCATTACTAGTATTTAACCATTCATATGTCAACAAACAACGAAGATTACATAAACCATATAGACAACTTC











































AACATTCAAGTTAAGCCAAGTAATTAACGAGATTTAAGACCTACCACAAACACTAAATCGTCTAAGCCATACCAAGTCTC

AACATTCTCTTAAATTCAAGTAACTAAACGAAATTTAAGACCTAACACAAATCTAAATCGTCTAAGCCATACCAAGTCTCC  
AACATTCTCTCAAATTCAAGTAATTAACGAGATTTAAGACCTAACACAAATCTAAATCGTCTAAGCCACACCAAGTCCCC

AACATTCTCTCGAGTTCAGGTAATTAACGAGATTTAAGACCTAACACAAACCTAAATCGTCTAAGCCATACCAAGTCCCC

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CCCCATCCCTGAAATCAAGTAAACTTAAG

TCATCTCTGAGGTCTCGCAAATTCAAG  
TCATCCCTGAAATCTGGTAAATTTAAG

TCATCCCTGAGGTCTAGTAAATTTAAG

CCTCATCCCTGAAATCAAGTAAATTTAAG

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ACCATCCCTGAAGTCTGGTAAATTTAAG  
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