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Nanosized Hydroxyapatite and Other Calcium Phosphates: Chemistry of Formation and Application as Drug and Gene Delivery Agents

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Abstract The first part of this review looks at the fundamental properties of hydroxyapatite, the basic mineral constituent of mammalian hard tissues, including the physicochemical features that govern its formation by precipitation. A special emphasis is placed on the analysis of qualities of different methods of synthesis and of the phase transformations intrinsic to the formation of hydroxyapatite following precipitation from aqueous solutions. This serves as an introduction to the second part and the main subject of this review, which relates to the discourse regarding the prospects of fabrication of ultrafine, nanosized particles based on calcium phosphate carriers with various therapeutic and/or diagnostic agents coated on and/or encapsulated within the particles. It is said that the particles could be either surface-functionalized with amphiphiles, peptides, proteins or nucleic acids or injected with therapeutic agents, magnetic ions or fluorescent molecules. Depending on the additive, they could be subsequently used for a variety of applications, including the controlled delivery and release of therapeutic agents (extracellularly or intracellularly), magnetic resonance imaging and hyperthermia therapy, cell separation, blood detoxification, peptide or oligonucleotide chromatography and ultrasensitive detection of biomolecules, and *in vivo* and *in vitro* gene transfection. Calcium phosphate nanoparticles as carriers of therapeutic agents that would enable a controlled drug release to treat a given bone infection and at the same be resorbed in the body so as to regenerate hard tissue lost to disease are emphasized hereby as one of the potentially attractive smart materials for the modern medicine.

Keywords: Bone, Calcium phosphate, Drug Delivery, Hydroxyapatite, Precipitation, Soft Chemistry, Theranostics

Introduction: Hydroxyapatite as the main constituent of bone

Hydroxyapatite (HAP) is to most people known as the mineral component of bone^{1,2,3}. Therein, stiff HAP crystals are responsible for imparting an appropriate compressive strength, whereby collagen fibers, able to dissipate energy effectively, provide superior elastic properties, thus ameliorating the brittleness of the sole HAP⁴. However, the exceptional stiffness and strength of bone come not only from the synergetic combination of material properties of its mineral and organic components, but from its hierarchical, superstructural organization as well⁵.

Bone is an organ that not only does provide a basic mechanical support to the body by generating and transferring forces that are involved in locomotion, but it also has various other functions⁶. For example, by storing minerals within, mostly calcium and phosphate, it presents the main mineral reservoir for the body. Absorption and release of salts is the mechanism by which bones buffer the blood and prevent excessive pH changes. Bones, such as skull or ribs,

also serve to physically protect vital internal organs, including brain, heart and lungs. Some bones also act as fabrics for producing red and white blood cells. Bone has formed in co-evolution with the surrounding tissues of organisms and the environment. In eukaryotic cells, calcium ions play a plethora of functions, including that of a messenger in various signal transduction pathways. Calcium-activated ATPase, $\text{Na}^+/\text{Ca}^{2+}$ exchangers, calcium channels and intracellular calcium binding proteins maintain a fine Ca^{2+} homeostasis in cells such as odontoblasts. On one hand, a controlled imbalance of internal calcium levels is a precursor for the bone-building activity, which, unlike in the case of amelogenesis and dentinogenesis, always proceeds through external accretion. On the other hand, increased amounts of intracellular Ca^{2+} may result in uncontrolled secretion and internal precipitation. Namely, Ca^{2+} is known to be a part of harmful deposits in the body, such as atherosclerotic plaque, kidney stones and dental calculi⁷. It also has a strong tendency to bond with many proteins, particularly phosphorylated ones. In case of ATP, its aggregation may be induced thereby, resulting in the loss of biological function. On the other hand, binding of Ca^{2+} to phosphorylated proteins involved in biomineralization, such as osteopontin in bone or dentin phosphophoryn, is a vital step in formation of hard tissues of bone and dentin⁸.

Aside from its mineral and protein components, bone is also populated by cells, macromolecules and blood vessels. Three types of cells are involved in maintaining a healthy bone structure: a) osteocytes involved in signal transduction of mechanical stimuli; b) osteoblasts, which are derived from mesenchymal stem cells and secrete collagenous proteins, thereby building the bone material; and c) osteoclasts, which are derived from hematopoietic marrow cells and secrete acids and proteases, thereby degrading the mineralized tissue. RANKL, a protein molecule bound to the osteoblast surface and serving to activate osteoclasts has been intensively investigated because of its role in facilitating an optimal communication between these two types of cells⁹. Overexpression of RANKL has been linked to a variety of degenerative bone diseases, including rheumatoid arthritis and osteoporosis¹⁰. Through the cooperative action of osteoblasts and osteoclasts, bone is constantly being remodeled in response to the physiological requirements. Julius Wolff was the first to propose that bone remodels itself when force is exerted on it by the mechanism according to which the internal architecture of the trabecular bone first undergoes adaptive changes when placed under load, followed by secondary changes to the external cortical portion of the bone¹¹. An evidence of an impressive remodeling capacity of bone has come from the observed modifications in bone shape and density in astronauts subjected to microgravity conditions for prolonged periods of time¹². Bone is, for example, often regarded as a living mineral due to its continual growth and dissolution, formation and degradation, renewal and remodeling, taking place during the organism's lifetime.

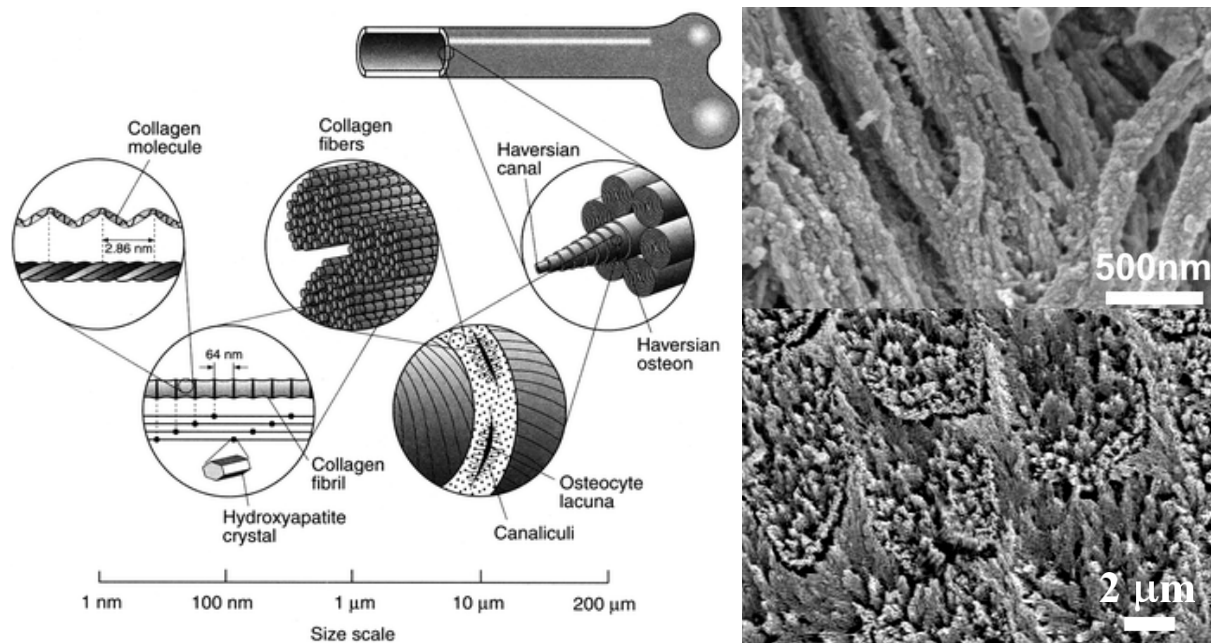


Fig.1. Bone (left) is a complex, hierarchically structured biological material in which the building components are precisely arranged at scales spanning half a dozen of orders of magnitude. The image on the left shows sketches of the structural elements of cortical/compact bone (which comprises the harder, outer layer of the cross-section of bone, surrounding the softer trabecular/spongy/cancellous bone) at different scales. The image on the upper right side shows the nanostructure of mineralized collagen fibers in bone, whereby the image below displays the fine structure of dental enamel, the hardest substance in the body. Within the former structure, HAP particles are incorporated within the organic matrix, whereas the latter structure is composed of an almost pure mineral with elongated HAP nanofibers connected into bundles and forming equally uniaxially directed enamel rods. Reprinted with permission from Refs. 13, 14, and 15.

Many mysteries, such as the role of “sacrificial” bonds that break under stress, but only to be reformed at a later time, imparting durability and resilience to hard tissues, still surround the superior functioning of this basic material of Nature¹⁶. Many new functions of bone have also been gradually revealed over time. It has now been established that bone does not only act as a reservoir for minerals, primarily calcium and phosphate, which circulate through blood in supersaturated concentrations, but also stores growth factors, fatty acids, heavy metals and other toxic elements, and is involved in buffering the blood by controlled release of alkaline salts. Be that as it may, bone presents a connective tissue and a failure of the ability to stay “connected” and properly transmit stress throughout the body has an implication of slowly bringing about dysfunction of many other, close or distant segments of skeleton and the body. In view of that, finding the ways to heal the impaired bone tissue in timely manner can be regarded as one of the most important tasks that medicine can contribute to. In view of that, bone research has a special meaning in the world of medicine. However, understanding bone is a challenging task for the modern scientists, especially because it requires an atypical inter-disciplinary element in one’s approach, explained by the fact that bone stands at the interface between many separate fields of science. The more one focuses one’s attention on finer levels of organization, the more of the biological approaches cede their place to fundamental physicochemical methods of probing bone structure and properties.

And just as usual, as scientific attention is focused to ever smaller physical details, the things do not get simplified, but become ever more complex instead¹⁷. Hence, the structural arrangement of nanosized HAP crystals within the collagen matrix is still a subject of scrutiny. It

is still not resolved whether mineralization first occurs within the 40 nm wide gap or the 27 nm wide overlap region of collagen fibers. Contrary to earlier assumptions, a recent cryo-TEM study has shown that upon mineralization of collagen the mineral first fills the overlap region and only then it proceeds to incorporate itself within the gap region¹⁸. Two forms of HAP crystals in dentin and bone can also be distinguished based on where they are found: extrafibrillar and intrafibrillar¹⁹. The former are larger nanocrystals (plate-shaped with 2 – 3 nm in thickness and a few tens of nanometers in length and width) existing in-between individual fibrils (that is, bundles of collagen fibers having ~ 1.5 – 3.5 nm in diameter, and are essentially individual elongated, triple helix molecules), whereas the latter are found to figure as links between individual fibers along their long axis. However, the role of each has not been discerned yet²⁰.

Also, just as the structural water has been added to the basic structural diagrams of proteins in recent years²¹, its role for the proper mechanical performance of hard tissues has been increasingly pointed out by researchers²². Collagenous tissues, such as bone or dentin, which contain approximately 70 wt% apatite, 20 wt% collagen and 10 wt% water (with only about 3 % of noncollagenous proteins, including some polysaccharides as well), are nowadays known to partly owe their mechanical properties to the structural water^{23,24}. Hydrated dentin is, for example, shown to dramatically degrade in toughness following its dehydration^{25,26}. Also, in case of collagen molecules wherein backbone hydrogen bonding between polypeptide chains in its triple-stranded structure does not present the major stabilizing force, unlike in the case of α -helices and β -sheets, additional enthalpic contributions are known to come from water molecules that form a “scaffold” around the surface of the triple helix, implying that water plays an intimate role in stabilizing this protein²⁷. This observation coincides with the recently observed 10-fold drop in tensile properties of single fibrils of collagen following desorption of the bound water in vacuum, even though the strength of the molecule was the same in water and air²⁸.

Hence, even though the fascinating properties of bone are products of precise and intricate arrangement of its building blocks on many different levels – from nanometer to millimeter scales (Fig.1) – the complexity of each one of these building blocks is equally complex as to deserve paying sole research attention thereto. In fact, the complexity of this material has ever since puzzled scientists involved in bone research; hence, the name of this compound, apatite, derives from Greek *απαταω*, which means “to deceive”. The respective mineral was, however, named so because it had easily been mistaken for other, more precious minerals²⁹; yet, a drop of lime juice was sufficient to dissolve it. Unlike some other similarly complex materials, such as doped manganites which exhibit an enormous set of electric and magnetic behaviors depending on the structural arrangement of the constitutive ions^{30,31}, in case of HAP the main emphasis is on the breadth of possible mechanical properties depending on different phase arrangements and the structure and morphology of the compound. Another remarkable feature of this material is a considerably low crystal growth rate even under relatively high supersaturations. The reason for this is thought to lie in the complex growth mechanism that involves precipitation of amorphous, ~ 1 nm sized solid units called Posner’s clusters in the first stage of the process, and their aggregation and ionic rearrangement followed by an increased compactness and crystallinity in the second stage³². Owing to the fact that this mechanism resembles the one of the growth of protein and viral crystals that involves chirality selection and orientation arrangements, the low crystal growth rate of HAP is often compared to that of these biological compounds³³. Low crystallinity of particles precipitated under physiological conditions and stoichiometric sensitivity to mildest changes in synthesis conditions are additional

characteristics of the formation of HAP, which will be discussed more in the following section. All of these fundamental peculiarities are currently subject of intensive scrutiny.

The main content of this review is divided to two parts: fundamentals and application. In the first part, the reader will get acquainted with the fundamental properties of HAP and the chemistry of its formation, whereas in the second part the advanced applications of this compound will be numbered and discoursed on.

1. Fundamentals

1.1. Chemical identity and basic properties

HAP is a mineral from the family of apatites, the general formula of which is $M_5(ZO_4)_3X$, where M is a rare-earth metal, such as Ca^{2+} , Cd^{2+} , Sr^{2+} , Ba^{2+} , Pb^{2+} , Zn^{2+} or Mg^{2+} , ZO_4 could be PO_4^{3-} , CO_3^{2-} or SO_4^{2-} , and X is OH^- , F^- , Cl^- or CO_3^{2-} . Whereas $Ca_5(PO_4)_3OH$ is the formula for HAP, $Ca_{10}(PO_4)_6(OH)_2$ is the formula of the unit cell thereof, and its pK_{sp} (K_{sp} = solubility product) equals 58.65 at 37 °C. HAP is, therefore, the most stable calcium orthophosphate (CAP) phase in the pH range of 4.2 – 12.4. The unit cell of stoichiometric HAP can also be represented with the following formula: $M1_4M2_6(PO_4)_6(OH)_2$, where M1 and M2 are two different crystallographic positions for ten calcium atoms. Four Ca atoms in the unit cell of HAP thus occupy M1 position where they are surrounded by nine oxygen atoms which belong to PO_4 tetrahedra. The other 6 Ca atoms occupy M2 site where they are coordinated by six O atoms of the PO_4 tetrahedra and one of the two OH^- groups^{34,35}. In biological conditions, HAP is subject to an extensive substitution of ions, so that human bone is, for example, best described as $(Ca,Z)_{10}(PO_4,Y)_6(OH,X)_2$, where $Z = Na^+$, Mg^{2+} , K^+ , Sr^{2+} , etc., $Y = CO_3^{2-}$, HPO_4^{2-} , and $X = Cl^-$, F^- .

Irrespective of whether it is found in enamel, dentin, cementum or bone, biogenic apatite is always impure and non-stoichiometric. The major impurity is CO_3^{2-} (3 – 8 wt%), whereby minor impurities include Na^+ (0.5 – 1 wt%), Mg^{2+} (0.4 – 1.2 wt%), K^+ (0.03 – 0.08 wt%), Cl^- (0.01 – 0.3 wt%), and F^- (0.01 – 0.06 wt%). Some of these substitutions, such as $OH^- \rightarrow Cl^-$, $PO_4^{3-} \rightarrow CO_3^{2-}$ or $Ca^{2+} \rightarrow Mg^{2+}$, Sr^{2+} , are known to weaken the apatite structure and make it more soluble, whereas others, such as $OH^- \rightarrow F^-$, are known to additionally strengthen it and also lessen its solubility³⁶. In fact, most of these impurities, except fluoride, contribute to increased solubility of the resulting apatite stoichiometry, which explains why fluorine is added to toothpaste formulations³⁷, although health risks concerning its unfavorable effects on the gut flora and indications that it may even act as a neurotoxin³⁸ are currently under research. Also, extensive amounts of fluoride in the apatite structure have been shown to lead to increased porosity and weakening of the material³⁹.

CO_3^{2-} is the most common dopant in biological apatite⁴⁰, and carbonated HAP is shown to have an improved bioactivity compared to pure HAP, which has been attributed to the greater solubility of the carbonated phase⁴¹. Incorporation of CO_3^{2-} ions within the HAP crystal structure has also been shown to have a retarding effect on the crystal growth⁴². The complexity of this substitution is, in fact, so high that it extends beyond the area of biomineralization and touches the most fundamental problems of crystallography and crystal growth phenomena⁴³. When apatite is synthesized by precipitation, at atmospheric conditions, B-type of carbonated HAP normally forms where CO_3^{2-} ions substitute for PO_4^{3-} . Typically, when HAP is prepared by means of an annealing treatment, A-type forms where CO_3^{2-} ions substitute for OH^- , owing to the

high mobility of OH⁻ groups⁴⁴. However, this is not necessarily the case since A-type substitution was also observed in HAP prepared by precipitation at room temperature⁴⁵. Biological apatite also normally presents a mixture of A and B types^{46,47}. In both cases, the fact that the divalent carbonate substitutes trivalent phosphates or monovalent hydroxyls implies the necessity to compensate for the imbalanced charges by modifying the stoichiometry of the compound or favoring incorporation of other impurities into the crystal lattice. On the basis of charge neutrality, the chemical formula of carbonated HAP is Ca_{10-x/2}[(PO₄)_{6-x}(CO₃)_x][(OH)_{2-2y}(CO₃)_y], where x and y are numbers of CO₃²⁻ ions substituting for PO₄³⁻ and OH⁻, respectively⁴⁸.

The most frequently encountered structure of HAP belongs to a hexagonal system with P6₃/m space group. The less common symmetry of HAP lattice is described by P2₁/b space group within the monoclinic crystal system⁴⁹. The unit cell dimensions of pure hexagonal HAP are: a = b = 0.937 nm, c = 0.6881 nm. However, each one of the substitutions has an effect on the lattice parameters, so that biological HAP deviates from the perfect crystal arrangement of ions depending on the additives and the positions that they occupy in the lattice. For example, A-type substitution upon carbonation of HAP implies larger CO₃²⁻ ions replacing smaller OH⁻ ions and thus expanding the *a* axis and contracting the *c* axis, whereas B-type substitution has an opposite effect^{50,51}. These changes in crystal lattice parameters often induce changes in crystallinity, thermal stability, morphology, solubility and other physicochemical and biological properties of the material⁵².

HAP is a ceramic material, and most of the primary chemical bonds found in ceramic materials are a mixture of ionic and covalent types, which explains for complex interplay of latent properties that HAP and ceramic materials in general are prone to exhibit. As expected, CAP can adopt numerous crystal structures depending on stoichiometry and conditions of formation. Some of the main CAP phases are listed in Table 1, whereby the crystal structure of HAP is illustrated in Fig.2.

Table 1. The main CAP phases obtainable upon precipitation from the solution.

Phase	Chemical formula	Space group	pK _{sp} at 37 °C	Ca/P molar ratio
Monocalcium phosphate anhydrous (MCPA)	Ca(H ₂ PO ₄) ₂	Triclinic P ¹	1.14	0.5
Monocalcium phosphate monohydrate (MCPM)	Ca(H ₂ PO ₄) ₂ ·H ₂ O	Triclinic P ¹	1.14	0.5
Dicalcium Phosphate (DCPA, Monetite)	CaHPO ₄	Triclinic P ¹	7.0	1
Dicalcium Phosphate Dihydrate (DCPD, Brushite)	CaHPO ₄ ·2H ₂ O	Monoclinic I _a	6.6	1
α-Tricalcium	Ca ₃ (PO ₄) ₂	Monoclinic	25.5	1.5

Phosphate (α -TCP)		$P2_1/a$		
β -Tricalcium Phosphate (β -TCP, Whitlockite)	$Ca_3(PO_4)_2$	Rhombohedral $R3cH$	29.5	1.5
Tetracalcium Phosphate (TTCP)	$Ca_4(PO_4)_2O$	Monoclinic $P2_1$	37.5	2
Octacalcium Phosphate (OCP)	$Ca_8H_2(PO_4)_6 \cdot 5H_2O$	Triclinic P^1	97.4	1.33
Hydroxyapatite (HAP)	$Ca_{10}(PO_4)_6(OH)_2$	Pseudo-Hexagonal $P6_3/m$	117.3	1.67

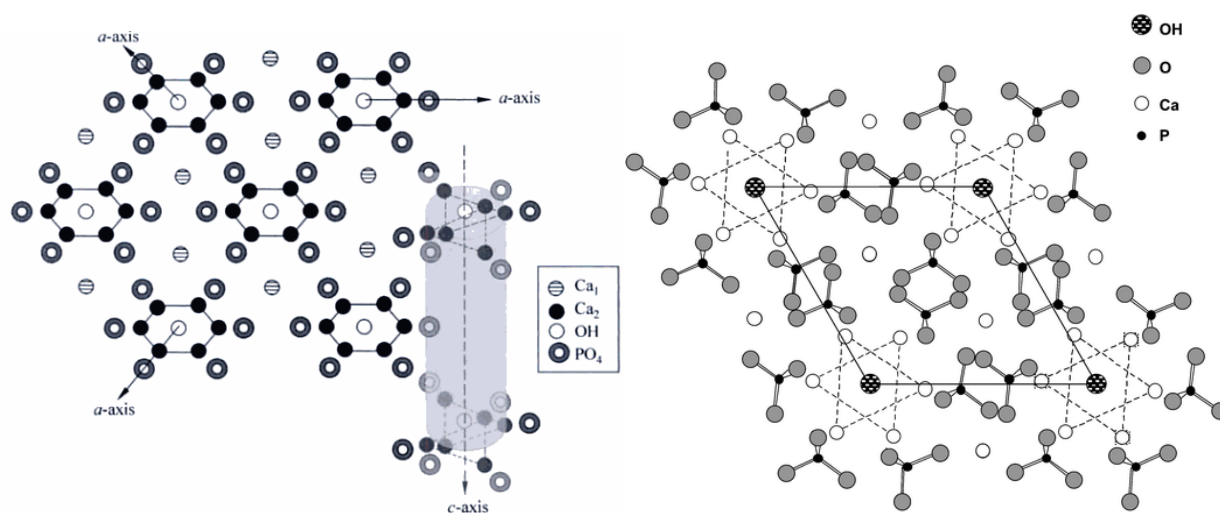


Fig.2. Crystal structure of HAP showing its c-axis perpendicular to 3 a-axes lying at 120° angles to each other (left), with projection on the (001) plane of HAP structure (right).

1.2. Typical synthetic pathways

Numerous methods have been applied for the synthesis of fine particles of HAP, including reactions in solid state^{53,54,55,56}, abrupt or continuous precipitation from solution^{57,58,59,60}, hydrothermal processing^{61,62}, electrospraying⁶³, electrospinning⁶⁴, flux cooling method⁶⁵, spray pyrolysis⁶⁶, ultrasound⁶⁷ and mechanochemical processing^{68,69}, microwave irradiation assisted precipitation^{70,71}, microemulsion- and surfactant-assisted precipitation^{72,73,74,75,76}, sol-gel syntheses based on hydrolysis of metal organic precursors^{77,78}, and chemical vapor⁷⁹ and plasma deposition⁸⁰. However, from the point of view of control over apatite structure and morphology, these methods could be roughly divided to low and high temperature ones. The latter typically involve mechanochemical homogenization of precursor compounds, such as $Ca_3(PO_4)_2$ and $Ca(OH)_2$, and their subsequent annealing at $\sim 1000^\circ C$. The advantage of this method lies in the ability to precisely set the stoichiometry of the final product, whereas long reaction times and high annealing temperatures are some of the main downsides. In general, if the molar ratio of Ca/P is not set to 1.667 during preparation, extraneous phases normally appear: α - or β -TCP at lower values and typically CaO at higher values, which is especially the case upon annealing (with α -TCP forming at higher temperatures, such as around $1200^\circ C$, and β -TCP forming at lower temperatures, such as up to $900^\circ C$). In addition to the high

levels of energy consumption, another major downside of high-temperature, solid state methods lies in the difficulty to produce uniform nanosized particles by their means. The latter are, on the other hand, known to promote a favorable biological response, including higher osteoinductivity and osteoconductivity, leading to a quicker integration of the implanted HAP-based material⁸¹. Low-temperature methods that involve precipitation from solution have a disadvantage in the frequent presence or transient and metastable phases in the final product. As we shall see in what follows, it is exactly this multitude of possible phase combinations, sensitive to subtle changes in synthesis conditions⁸², that may be linked to the aforementioned Greek origins of the word “apatite”.

HAP is a sparingly soluble salt in neutral and alkaline aqueous solutions, and forms crystals with high edge free energies. As surface/interfacial energy of a crystal is indicative of the difficulty of its forming, sparingly soluble salts as a rule have higher interfacial free energy values than soluble ones. Also, as the interfacial tension can be expressed as proportional to the size of the critical step/nucleus upon dissolution/crystallization, high values of the latter can be deduced and correlated with the experimentally observed slow dissolution/crystallization of HAP⁸³. As a direct result, synthesized HAP typically exhibits not only a wide distribution of particle sizes (partly caused by the aggregation mechanism of growth, which will be discussed in the next subsection, and partly by slow crystallization, which makes it difficult to avoid the overlap between nucleation and diffusional crystal growth phases), but highly irregular particle shapes as well. In view of that, finding conditions for a controllable synthesis of HAP nanoparticles with tunable sizes and shapes presents a worthwhile research challenge. Various additives have been applied in precipitation syntheses for the sake of attaining this aim, including cetyltrimethyl ammonium bromide (CTAB)⁸⁴, poly(acrylic acid) (PAA)⁸⁵, poly(allylamine hydrochloride)⁸⁶, and others.

Biological apatite is, moreover, known for its high content of defects, caused in part by a relatively large percentage of impurities, all of which affect the lattice parameters, crystal morphology, crystallinity, solubility and the thermal stability of the material. HAP crystal surface is thus rarely smooth, and the reason is related to its biological significance. The exceptional roughness of biological HAP, comprising surface irregularities in the order of size of single unit cells, hypothetically corresponds to the tendencies to increase protein binding in the process of biomineralization⁸⁷. Previous studies have shown that rough surface improves biocompatibility of the material and has a positive effect on inflammatory reactions, whereas the viability of monocytes seeded on flat surfaces tends to be far lesser⁸⁸. Also, cells are constantly creating and decomposing HAP in bone, and rougher surfaces provide conditions for an easier anchoring of cells thereto. An *ab initio* model has showed that amino acid side chains carrying either acidic (i.e., aspartic and glutamic acids) or basic (i.e., lysine or arginine) residues are prone to interact with HAP crystal surfaces, which is in agreement with numerous spectroscopic and other modeling data that report both carboxylic and amino residues typically found in close proximity to the HAP surface⁸⁹. In fact, the reason why phage display panning techniques on HAP surfaces have not yielded significant results so far lies exactly in the tendency of numerous peptide combinations to bind to it, which diminishes the high selectivity on which this approach inherently depends⁹⁰.

The typical models of protein-mineral interaction refer to specific binding of additives to crystal faces, which are then prevented from growing. Such is the case for the current model of amelogenesis wherein nanospherical particles of amelogenin are assumed to bind onto (hk0) faces of the growing HAP crystals and thus foster their uniaxial growth along [001] axis⁹¹.

However, one of the opposing actual explanatory trends lies in invoking the surface-controlled channeling or the diffusion-controlled transportation activity of additives for the sake of building apatite crystals using either amorphous units or individual ions and complexes as building blocks. Such an idea is supported by the successful synthesis of a few ceramic materials, previously obtainable only through high-temperature annealing treatments, by precipitation at room temperature in the presence of short peptides derived from phage display libraries^{92,93}.

Precipitation of HAP is, in fact, often quite morphologically sensitive (Fig.3) to the presence of additives^{94,95,96,97,98,99}. Electrostatic forces that govern the interaction between growing inorganic particles and given additives and tunable by controlling the ζ -potential of colloidal particles are thus regularly shown as crucial for setting the optimal conditions for the growth of nanocrystals in desirable sizes and morphologies. For example, elongated HAP particles were obtained in the presence of poly(L-lysine), whereas in the presence of more charged poly(L-glutamic acid), small nanocrystals resulted under otherwise same conditions of precipitation¹⁰⁰. This was explained by assuming that the more charged the heterogeneous nucleation surface, the more cations will be attracted thereto and the more nuclei will be formed. Consequently, the particles will be smaller and there would be no biaxial growth, unlike in the case of the less charged macromolecular surface. On the other hand, hydration effects could not be reduced to a simplistic rule of a thumb, and in reality it is difficult to predict the effects of addition of salt and pH change as they are always synergistically bound with the effects of other species present in the actual reaction system. A recent computer model¹⁰¹ has thus shown ions with low charge density adsorbed preferentially on the surface of a hydrophobic particle, leading to micelle-like clusters of particles, whereas ions with high charge density tended to be depleted from the particle surface, leading to formation of similar clusters of dispersed hydrophobic particles. Only in the intermediate case, conditions for the dispersion of individual particles and avoidance of the formation of clusters were found. Generally speaking, the effect of additives on composition, structure and properties of HAP particles is so intensive that we can over and over again recall the words of Stephen Mann of Bristol University: "Such is the lure of the organic matrix that you can attend some scientific conferences, notably those on bone mineralization, and hardly ever come across a serious mention of calcium phosphate"¹⁰².

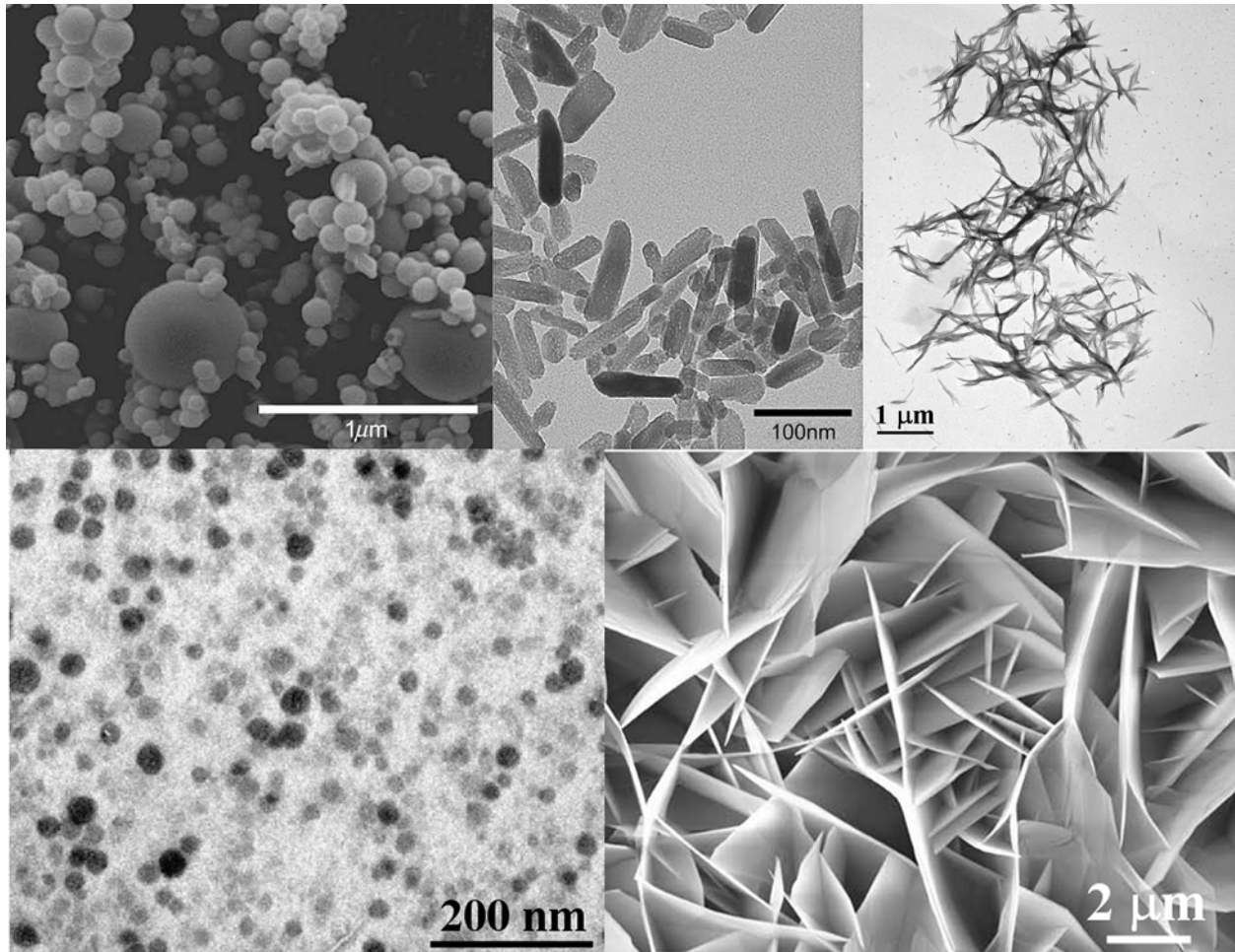


Fig.3. Owing to its structural flexibility, HAP can be prepared in a variety of morphologies, ranging from spheres to filaments to rods. However, HAP particles are on most occasions found in the forms of plate- or needle-shaped particles (bottom left), whereas their spherical nanosized morphologies are in most cases the sign of their amorphous nature (bottom right). Reprinted and adapted with permission from Refs. 103 and 104.

Methods for preparation of fine CAP particles usually follow a soft chemical route^{105,106}, which involves precipitation from solutions or suspensions of precursor salts, ideally with no subsequent high-temperature annealing treatment. Entrapment of additional composite components is carried out by precipitation of CAP in the presence of dissolved cargo molecules. Surface functionalization, on the other hand, normally requires a time-delayed introduction of the surface-anchoring molecules owing to a sufficient aging that the initial precipitate has to undergo in solution prior to the formation of HAP phase. Conjugation of proteins, peptides, polymers, cell-penetrating moieties, reporter groups and other functional ligands to the carrier surface is in the case of CAP particles expected to proceed non-covalently, that is, via adsorption governed by hydrophobic or van der Waals forces.

Amphiphilic mixtures in form of reverse micelles, other microemulsion phases or simple steric dispersants could be implemented with the purpose of preventing agglomeration of particles and stabilizing their suspensions^{107,108,109}. Spray drying enables drying of the precipitated powders either without inducing an undesired agglomeration of the particles or with forming attractive and uniform agglomerates of smaller particles¹¹⁰. Bis(2-ethylhexyl)sulfosuccinate (AOT) has thus recently been used for the purpose of assembling HAP

particles into enamel-like bundles¹¹¹. Successive modification of surface charges of the particles and of an adsorbent during the cycles of washing and rinsing can also be carried out to avoid agglomeration and yield pure and stable dispersions of CAP nanoparticles. Agitation by means of ultrasound is often used for the same purpose, although it can markedly affect not only the particle size distribution, but their morphologies too. Hence, in a particular study, ultrasound applied during the ripening of a precipitate comprising hollow CAP nanospheres led to their transformation into fibrous particles¹¹². As for the particle composition, 80 % HAP and 20 % TCP is usually pointed out as an ideal phase composition ratio from the point of view of the optimal bioresorbability of the compound¹¹³. However, by varying HAP/TCP ratio in the final compound, the optimal degradation rate thereof, defined by the intended application in the body, can be set. Porosity, usually controlled using a porogenous agent, such as poly(methyl methacrylate) (PMMA)¹¹⁴, presents another structural property of the particles that can be used to control the release rate of encapsulated compounds¹¹⁵. The particle size presents another important parameter that defines the bioresorption rate of HAP materials. As expected, smaller, nanosized particles are associated with significantly higher resorption rates compared to the bigger, microsized ones¹¹⁶. In addition to composition and size, the particle shape is also known to influence the uptake efficiency of the encapsulated drug¹¹⁷. The morphology of CAP particles may be thus used as a parameter in optimizing a favorable bioresorbability thereof. It is usually claimed that the particles produced in forms of powders or suspensions should ideally be non-agglomerated, spherical and uniform in size and morphology, which is to ensure the reliability and reproducibility of their application in the body. With respect to that, the surface of HAP particles could be modified with various additives, such as: hexanoic and decanoic acids that hydrogen-bond to the surface P-OH groups; alkanolic acids with longer alkyl chains so as to render the surface hydrophobic; oleic acid typically applied for the stabilization of magnetic fluids; sodium dodecyl sulphate or other surfactants; or through covalent bonding, the example of which may be esterification of surface P-OH groups with dodecyl alcohol, alkyl phosphates, pyrophosphoric acid, hexamethyldisilazane or various silanes which could be used as precursors for silica coating^{118,119}. Surface modifications affect the surface charge, hydrophilicity, colloidal stability of the particles and their interaction propensities, thereby affecting their biological response as well.

For the purpose of obtaining small, nanosized particles, abrupt precipitation procedures, yielding a high density of crystallization nuclei, are to be performed. Avoiding the overlap of nucleation and crystal growth phases is normally considered as the key to achieving monodispersity of synthesized particles. Low nucleation rates and fast crystal growth rates are, in contrast, the fundamental recipe for producing elongated HAP crystals. For example, when a slow and controlled decomposition of urea was used to promote changes in pH and provide conditions for precipitation, the HAP crystals formed were of plate-shaped and needle-shaped character¹²⁰. Highly strained single-crystal apatite fibers with 20 – 60 μm in length and 100 – 300 nm in diameter were also obtained in a precipitation reaction using urea as the alkaline agent¹²¹. pH and precipitation temperature as well as the molar ratio of precursor ions are often control parameters used to optimize and fine-tune the structural, morphological and thereupon the biodegradation properties of the carrier particles. As it is known that the formation of HAP follows a multi-step route permeated with complex solid-solid transformation pathways, dictated by the empirical Ostwald-Lussac's rule of kinetics of phase transformations¹²², aging time is another experimental parameter applied to optimize the synthesis conditions (namely, insufficient aging can lead to amorphous components remaining in the system, whereas an overly

extensive aging can lead to Ostwald ripening and broadening of the particle size distribution that ripening naturally entails). What follows is exactly a discussion of the critical aspects of this complex chemical mechanism.

1.3. Mechanism of formation of HAP by precipitation

Even if explored all alone, without any involvement of additives, precipitation of HAP, especially at low supersaturations, still presents an enigma. Not only is this process sensitive to the mildest influences within the experimental conditions, such as the amount and nature of impurities or the texture and chemical identity of the reaction vessel (which sometimes drastically affects the level of critical supersaturation), but the chemical pathways leading to the formation of HAP as the final phase are also subject to change depending on mildest modifications of the initial experimental conditions. Due to this reason, researchers in the biomineralization field are still in dispute over the exact chemical mechanism of formation of HAP in biological and *in vitro* conditions alike. Ostwald-Lussac's rule predicts the highest nucleation rate for the least stable phase for which the supersaturation limit is exceeded under given conditions, implying that amorphous CAP will under most circumstances be the first phase to precipitate, followed by the solid state transformation to OCP first and only then to HAP. Ca(OH)₂ or TCP may be secondary phases depending on the exact stoichiometric ratio between the precursor Ca²⁺ and H_xPO₄^{x-3} ions in the solution. However, the exact chemical pathways, the transformation mechanism (dissolution/recrystallization or bulk rearrangement of ions within the prime crystal lattice), the transient compounds and time frames for the nucleation and growth of each one of the phases are still subject to uncertainty.

In any case, the nature of formation of HAP as predicted by Ostwald-Lussac's rule presents an argument against the sole diffusional growth of these crystals. Although the latter has for a long time been the major paradigm in explaining the formation of biological crystals, what we are witnessing today is a paradigm shift wherein the growth of crystals via amorphous precursors is slowly becoming the major explanatory model in the field^{123,124,125}. A large amount of evidence has been collected recently in favor of this new paradigm which proposes the role of macromolecules in linking and organizing precursor amorphous or crystalline units within the growing crystals¹²⁶. As of today, many variations on this theme flood the literature. According to one of the models, amorphous CAP acts as a "mortar" in cementing the "bricks" that are nanosized HAP units transported by biological molecules, and is also able to subsequently crystallize, contributing to the integrity of the final structure¹²⁷. Another model proposes these building blocks to be bubbles of a phase that stands at the boundary between liquid and amorphous solid. These small liquid/amorphous-like entities are supposed to form even without a polymeric process-directing agent, although in such case they are short-lived and difficult to detect. In addition, a strict application of Ostwald-Lussac's rule implies the formation of a structured liquid phase prior to the formation of an amorphous solid (usually considered as the first one to form, subsequently transforming to more stable solid phases). D'Arcy Thompson in his book *On Growth and Form* had thus argued: "In accordance with a rule first recognized by Ostwald, when a substance begins to separate from a solution, so making its first appearance as a new phase, it always makes its appearance first as a liquid"¹²⁸. Most of the polymer phase is returned back to the solution, although on rare occasions it may get trapped between the crystalline layers, serving as an evidence of said nature of formation^{129,130}. Invoking this mechanism, even monocrystalline systems, such as the basic crystalline units of enamel, micro-

sized apatite fibers, are accepted to be, may form through aggregation of subunits. The recent TEM studies have confirmed the existence of transient amorphous CAP phases in the developing enamel¹³¹. It has also been documented that long and defect-free filaments with the aspect ratios of up to 10^4 can be obtained by the aggregation mechanism¹³², overthrowing the idea that so grown apatite fibers would be prone to fracture due to many defects. In fact, in the context of a highly anisotropic organization of apatite crystals within enamel, such a slightly imperfect crystalline nature could be even proven as favorable.

As a consequence of Ostwald-Lussac's rule (Fig.4) and the relatively large number of phase compositions, the stages of which the precipitate has to pass through prior to reaching HAP crystal symmetry, the formation of the latter compound by precipitation can be expected to proceed at a relatively slow pace. The complexity and the crucial impact of the slow formation rate are best illustrated by the fact that the whole process of formation of elongated HAP crystals in dental enamel, having aspect ratios of up to 10^4 , takes more time to complete than is needed for the entire embryo to be created *in utero*¹³³. In view of such a slow rate of crystallization, there is a question whether this process could ever occur as a slow stream of crystallization, and not as a back-and-forth stream of crystallization/dissolution/crystallization, in which "mistakes" are made, but recognized and subsequently corrected, as is otherwise typical for biological syntheses^{134,135}. Namely, it is not the perfect reproduction, but a high selectivity for the product properties that typifies biosynthetic phenomena. A similar stochastic nature can be, in fact, seen as ingrained in practically every aspect of creativity exhibited in the biological world, from human thinking and reproduction to the evolution of life¹³⁶. This may also shed light on why the simultaneous activity of two types of cells – osteoblasts that build the bone material and osteoclasts that degrade the mineralized tissue – is required to maintain the functional structure of bone.

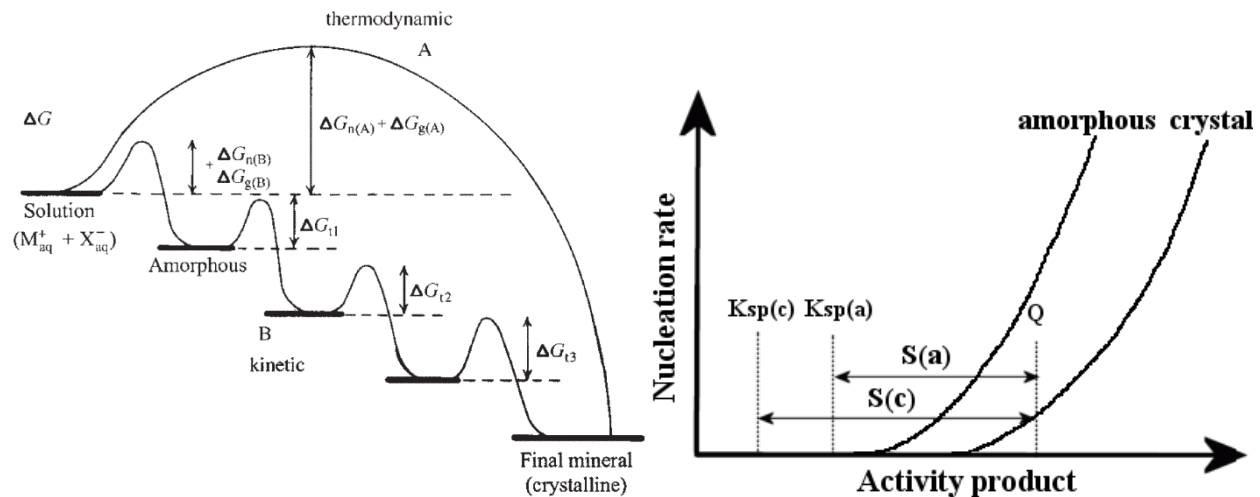


Fig.4. An inability of the system to traverse multiple energy barriers posed on its way to the final, most energetically favorable state implies its transition through a number of transient stages. As the Ostwald-Lussac rule suggests, the less stable polymorph will pose the lowest energy barrier in front of itself, so that the precipitated ions will adopt it as one of the temporary states on their way to settle into a more stable phase. The image on the left is reprinted with permission from Ref. 137. The diagram on the right demonstrates that even though supersaturation may be higher for a crystalline phase ($S(c)$) than for the amorphous ($S(a)$), the nucleation rate of the amorphous phase is higher at any given value of the product of ionic activities (Q), which is a consequence of a lower S being offset by a reduced interfacial energy.

As far as the major parameters that could be controlled so as to optimize the formation of HAP along a desired route are concerned, the following can be numbered: pH, ionic strength, temperature, concentration and identity of additives, Ca/P molar ratios and supersaturation¹³⁸. pH primarily affects precipitation of HAP with its effect on the amount of free hydroxyl groups and on the balance of phosphate species. A shift to lower pH will lower the saturation level by decreasing the concentration of free OH⁻ groups and shifting the balance of phosphate species from PO₄³⁻ to HPO₄²⁻ to H₂PO₄⁻ to H₃PO₄. Hence, the lower the pH, the more of the phosphate groups will be protonated and the precipitation will be less favored. Also, pH can shift the surface charge of the interacting particles by changing the distribution of proton and hydroxyl groups hydrating the interface. Although H₃O⁺ and OH⁻ are usually considered as charge-determining ions in case of HAP particles, ions other than these can adjust the surface charge, and in calcium-containing solutions, Ca²⁺ ions may bind to the negatively charged HAP surface at pH > IEP (isoelectric point), leaving the surface neutral rather than negative. The opposite effect can take place in phosphate-rich solutions when binding of H_xPO₄^{x-3} species at pH < IEP may result in a negatively charged HAP particle surface rather than the positive. These insights were, however, gained not by means of electrophoretic analyses but through the Brönsted isotherm¹³⁹, and are not necessarily in agreement with the electrophoretic studies¹⁴⁰. In view of the propensity of HAP to undergo dissolution/recrystallization processes, it is questionable how stable the structure and composition of the interface layer of HAP particles is as they undergo a pH change. It is known that the drastic effect of F⁻ in the suspension medium on the ζ-potential can be explained only by assuming its incorporation in the crystal lattice of the apatite¹⁴¹. Hence, an intensive exchange of ions between the solution and HAP particles is expected to take place, frequently shifting the ζ-potential in hardly predictable ways. This may explain why the surface composition of suspended or precipitated HAP particles is different compared to their bulk composition.

A hydrated layer containing relatively mobile ionic species is assumed to be present on the surface of precipitated CAP particles in the solution. The composition of this layer would be subject to change depending on pH and ionic content of the medium. Experiments carried out in our lab have, furthermore, shown that the ripening time of CAP precipitate has a marked effect on the surface charge of the particles. As can be seen from Fig.5a, ζ-potential of the particles in the pH range of 5 – 8 shifts from positive to negative values with increasing the aging time in solution from 10 min to 7 days. The high surface ion mobility is also thought to be responsible for the relatively high electrical conductivity exhibited by HAP. In addition to the high surface area of the mineral particles in bone, the ion-exchange propensity of HAP may be another factor crucial in providing the cellular environment with an access to the constituent ions for the sake of facile bone remodeling or consumption of ions for other purposes (as bone also acts as a frequently accessed mineral reservoir). One of the consequences of the maturation of bone may thus be correlated with the loss of these ion-exchange properties¹⁴². On the other hand, despite the high mobility of surface ions, HAP is typified by its sparsely soluble nature and slow crystal growth even at very high supersaturations^{143,144}. This implies facile reorganization and restructuring of ionic layers (that is, the solid surface layer of the particle and the double-layer of ions surrounding it) that contribute to surface charges following changes in the ionic environment. Indeed, ζ-potential of HAP particles has been observed to change with the immersion time for certain compositions, suggesting an intensive exchange of ions across the interface layer and its restructuring following changes in the solvent medium^{145,146}. It is natural to expect that the structure of the mobile surface layer would depend on the physical and

chemical conditions under which the particles were prepared, and this effect can be invoked to explain a large discrepancy between the IEPs and surface potential values for HAP particles reported in the literature. Consequently, it has been evidenced that methods for preparation and any changes in stoichiometry (Ca/P ratio) significantly affect IEP of HAP¹⁴⁷. Pure HAP powders precipitated in acidic conditions were, for example, shown to possess 1 – 3 pH units lower IEPs compared to those precipitated from alkaline solutions¹⁴⁸.

HAP particles in sols are also excessively prone to selective leaching of ions, which leaves different ζ -potential vs. pH curves of the same powder in different solvent media as a consequence¹⁴⁹. Such mild solubility effects are known to be responsible for the scatter in IEPs and points of zero charge (PZC) values reported in the literature. Thus, whereas some studies report negatively charged particles in the entire pH range in which HAP is the stable phase¹⁵⁰, others report IEP values at anywhere between 5 and 7.5, below which the particles should become positively charged^{151,152}. There are, however, reports¹⁵³ on IEP of HAP suspensions detected at pHs as high as 10. In addition, HAP has two types of crystal planes which carry different net charges: positive on *a* planes and negative on *c* planes, and there are speculations that the *a* planes tend to adsorb acidic proteins, whereas the *c* planes tend to attract the basic ones¹⁵⁴. Elongation of HAP particles along the *c*-axis would thus lead to a shift towards more positively charged particles with a higher specificity of adsorption onto negatively charged acidic proteins¹⁵⁵. Since biological entities are predominantly dispersed on the negative side, it comes as no surprise that positively charged HAP promotes good adhesion of cells thereto, whereas poor cellular adhesion and growth was observed on negatively charged HAP surfaces¹⁵⁶. Extracted biological HAP crystals were thus also shown to comprise a series of discrete and alternating domains of variously charged (in both magnitude and sign) surfaces (Fig.6a), independently of topography, indicating their intrinsic potential for periodic binding of matrix proteins under physiological conditions¹⁵⁷.

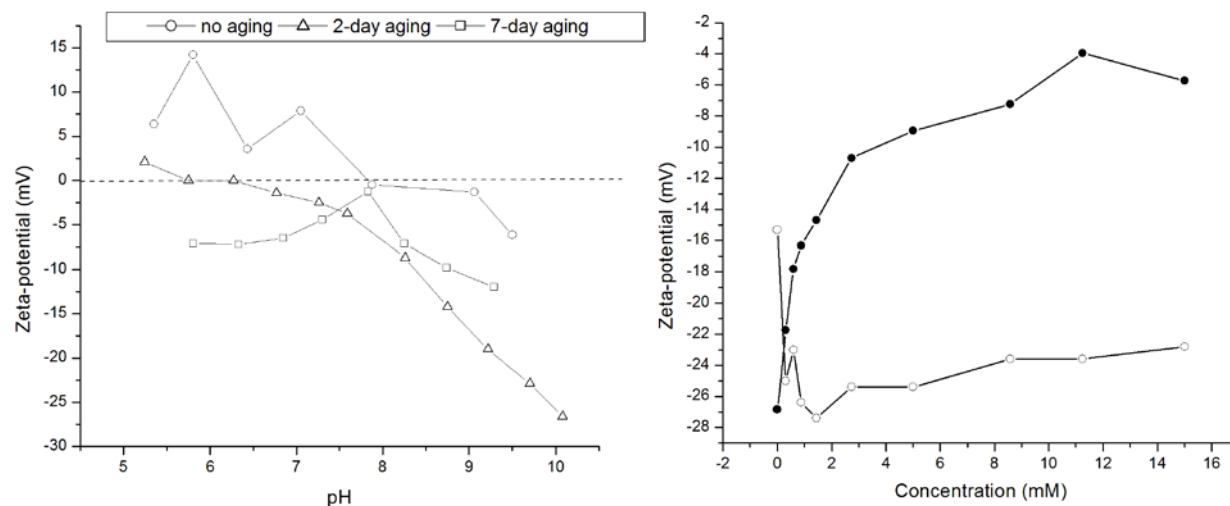


Fig.5. a) ζ -potential vs. pH curves for as-prepared, 2-day-aged and 7-day aged CAP prepared by precipitation from a solution comprising 25 mM $CaCl_2$ and 15 mM KH_2PO_4 at pH 10.5. b) ζ -potential of suspended HAP nanoparticles vs. $[Ca^{2+}]$ (-o-) and $[H_2PO_4^-/HPO_4^{2-}]$ (-●-) at pH 7.40 ± 0.02 . The graph shows that both of these ionic species adsorb on the HAP particle surface. The plateau observed in both cases signifies the adsorption of ions reaching a saturated level.

In addition to ions in the double layer surrounding the particle, HAP particles in suspension undergo practically all the mechanisms that contribute to the charged surface (that is,

adsorption, ionization and selective dissolution), which additionally increases the complexity of this physical effect. In case of hydrous metal oxide sols, in general, the surface hydroxyl groups (M–OH) become deprotonated at high pH values, transforming from –OH to –O[–] and thus contributing to the negative charge of the particle surface, whereas they become protonated, transforming from –OH to –OH₂⁺ at low pH values. As Ca²⁺ ions on the CAP particle surface are bound to OH[–] groups owing to hydration effects, protonation/deprotonation thereof can influence the particle charge. HAP particles also cannot be considered as immune to selective dissolution, which is known to contribute to imbalanced neutrality of ionic crystalline particles as wholes in many ionic crystals. For example, in the case of AgI particles, pK_{sp} = 16; however, the zero point charge does not exist at pAg 8, but is displaced to pAg 5.5 (i.e., pI 10.5) because smaller and more mobile Ag⁺ ions are held within the AgI crystal lattice less strongly than heavier and less mobile I[–] ions¹⁵⁸. A similar discrepancy in the mobility between Ca²⁺ and H_xPO₄^{x-3} ions can be inferred for the case of HAP crystal symmetry. Namely, Ca²⁺ ions are more flexibly arranged within the lattice, which is therefore more prone to display Ca²⁺ vacancies, whereas PO₄³⁻ groups are practically those that define the hexagonal structure of the crystal. The atomic arrangement of all calcium orthophosphates is built up around the network of PO₄³⁻ groups that provide stability to the structure¹⁵⁹. This may also explain why the crystallization of a few CAP phases has been shown¹⁶⁰ to be much more sensitive to the activity of phosphate species than to that of Ca²⁺ (Fig.6b), even though both ions adsorb well to the surface of HAP particles (Fig.5b). This is presumably due to weaker hydration attraction of solvated H_xPO₄^{x-3} groups to the surrounding protons than of solvated Ca²⁺ ions to the adjacent OH[–] groups. As a result, a higher activation barrier is attributed to Ca²⁺ in the solution than to H_xPO₄^{x-3}, which is also in agreement with the generally observed more pronounced solubility effect for anions of the Hofmeister series than for cations¹⁶¹, corroborating their more significant effect on the morphology of precipitated nanoparticles as well¹⁶².

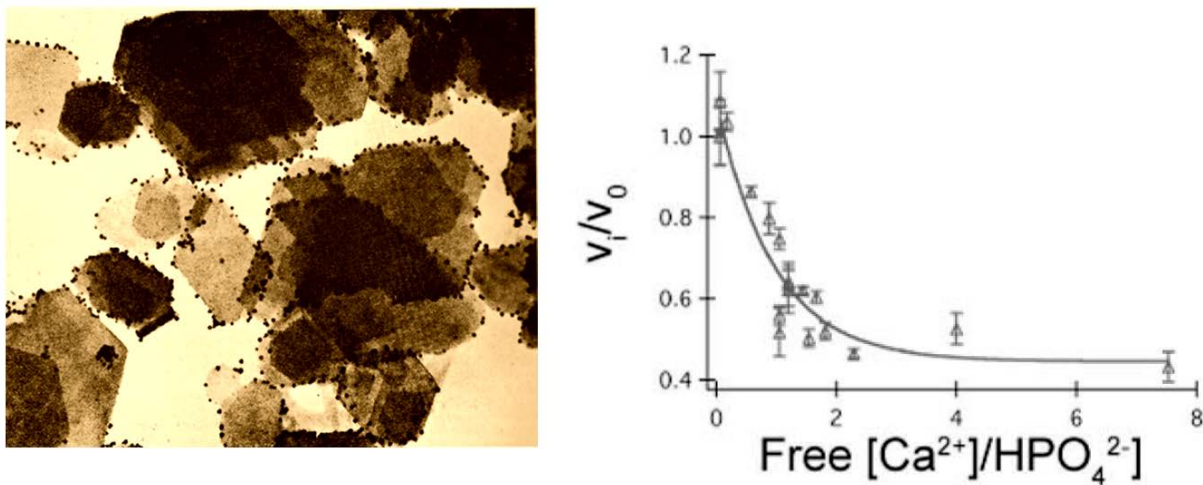


Fig.6. Both micro-sized kaolin crystals and nanosized gold particles on the left image are negatively charged as a whole. However, because edges of kaolin crystals are positively charged, gold nanoparticles effectively adsorb onto them. HAP crystals show a similar modulation of charges over their surface, which enables them to interact with an array of species relying on electrostatic attraction. Reprinted and adapted with permission from Ref.112. The image on the right shows increasing HAP crystal growth rates when the activity/abundance of H_xPO₄^{x-3} ions in the solution dominates over the activity/abundance of Ca²⁺ ions. Reprinted with permission from Ref. 163.

Ionic strength plays a role in screening ion-ion interactions (as well as the ones between charged colloidal particles) in the solution. Namely, surface charge density is equal to $\sigma = \epsilon\kappa\psi_0$, where ϵ is the dielectric constant of the medium (inversely proportional to the ionic strength), $1/\kappa$ is the length of the diffuse double layer (composed of Stern layer of adsorbed counter-ions and a diffuse layer composed of both counter-ions and co-ions, yielding as a sum the Debye length at which the electrical neutrality is again established), and ψ_0 is the surface potential. The addition of an inert electrolyte compresses the diffuse layer of charged ions and co-ions around each of the dispersed and charged particles. Namely, a higher concentration of co-ions and counter-ions implies screening of the particle surface charge at a closer distance to the particle. In other words, $1/\kappa$ decreases, which entails either an increase in σ or a decrease in ψ_0 , or both. For example, in case of AgI particles whose potential depends on the concentration of Ag^+ and I^- in the solution, addition of electrolyte and the corresponding drop in κ leads to adsorption of potential-determining Ag or I ions, which increases σ but keeps ψ_0 constant. However, for an ionogenic surface, σ stays constant, but ψ_0 drops. Simply saying, increasing ionic strength causes the layer of counter-ions around the charged particle to shrink and thus increases the propensity of the dispersed particles to agglomerate, which is known as “salting out” effect, and is known to be involved in the pathological calcifications in the body¹⁶⁴.

Now, it is well known that Nature disperses colloids almost exclusively on the negative side. Most cells and biological surfaces are thus negatively charged. The same can be said for HAP particles in physiological conditions. The effect of the ionic strength on (de)stabilization of biological colloids, including HAP, should thus be in theory primarily sensitive to the valence of the cation. Critical coagulation concentration (ccc) of the cation may be calculated using the following empirical formula: $\text{ccc} = 0.8/v^6$, where v is the valence of the cation. One can then see that for monovalent Na^+ $\text{ccc} = 0.8 \text{ M}$, for divalent Ca^{2+} $\text{ccc} = 12.5 \text{ mM}$, and for trivalent Al^{3+} $\text{ccc} = 1 \text{ mM}$. However, although CAP more efficiently nucleates on negatively charged surfaces, it can nucleate on positively charged ones as well for as long as the proper chemical identity is selected¹⁶⁵. On the other hand, the sign and intensity of the charge of the nucleation surface are known to play a determining role in defining the crystal symmetry of the resulting precipitate.

Ca/P molar ratios affect the formation of HAP firstly as the result of unequal activation barriers for cations and anions (Fig.6b). Although most crystal growth models neglect this effect by treating all the inclusive ions identically, whenever the crystal growth rate depends on the rate-limiting ion, this effect has to be taken into account in addition to other thermodynamic and kinetic factors¹⁶⁶. For example, due to extensive re-adsorption of $\text{H}_x\text{PO}_4^{x-3}$ ions onto the particle surface, Ca/P molar ratios in the equilibrium solution were often measured to be as high as 25 despite the composition of the precipitate corresponding to almost stoichiometric HAP¹⁶⁷. The ratio between Ca^{2+} and $\text{H}_x\text{PO}_4^{x-3}$ ions in the solution is also important because it may trigger precipitation of a specific phase combination and induce a particular phase transformation pathway in the solid state. Different Ca/P ratios can thus initiate different kinetic pathways for the reaction, which brings us to the discussion of the mechanism of precipitation of HAP.

The main parameter used to describe the latter is supersaturation ratio, S , defined as

$$S = Q/K_{sp}$$

Q is the product of ionic activities of precursor ions in the solution for the given stoichiometry, and for half a unit cell of HAP equals

$$Q = \{Ca^{2+}\}^5\{PO_4^{3-}\}^3\{OH^-\}$$

K_{sp} is the product of ionic activities for the given compound at the saturation level. In view of the continuous transfer of matter across the solid/liquid interface in both directions, K_{sp} could be also defined as the product of activities of dissolved ions of a solid substance in equilibrium between the dissolved ions precipitating and the precipitated ions dissolving. Ostwald-Lussac's rule dictates that the most soluble phase (that is, the least stable) for which $S > 1$ will be the first to precipitate, which will be successively followed by precipitation of less soluble phases. The reason for this is that the thermodynamic barrier posed between the state occupied by dissolved ionic species and the solid phase will be the lowest for the thermodynamically most unstable phase. An important contributor to this effect is the surface/interfacial energy, i.e., the work required to increase the surface area of a substance by one area unit, which is the hurdle that must be overcome when forming a solid phase. Amorphous phase is less ordered and will have a lower interfacial energy (and that particularly if it is hydrated – as such, it is actually most similar in chemistry to the surrounding aqueous environment) than any crystalline phase, which means that it tends to be the first to precipitate prior to subsequently transforming into a more stable, crystalline modification. In case of the precipitation of calcium phosphates, this means that HAP would be the last phase to form. The initial precipitation of the amorphous CAP is normally followed by nucleation of OCP at a certain stage. However, note that only phases with $S > 1$ are involved in this successive precipitation. Those for which $S < 1$ are assumed not to be precipitated at any stage of this process, and their appearance may only be transitory during phase transitions that involve rearrangements of ions in the solid state. This kinetic rule was empirically observed, although it can be nowadays supported by theoretical arguments. In one such calculation¹⁶⁸, it was shown that in simulated body fluid (SBF), whose S normalized per growth unit ($n = 9$ for HAP) equals 19.5 with respect to HAP, the nucleation rate of OCP is higher than the one for HAP, implying that OCP would be the preceding crystalline phase to form. DCP, however, cannot form under these conditions because the solution itself is undersaturated with respect to it. Only if $[Ca^{2+}]$ and $[H_xPO_4^{x-3}]$ in the fluid increase so that the solution becomes supersaturated with respect to DCP, the nucleation rate for this phase would become higher compared to the one for OCP and HAP. Also, as pH of SBF increases, the difference between the nucleation rates for the formation of HAP and OCP decreases, and at \sim pH 10 they become equal. In fact, as phases such as DCPD or OCP are thermodynamically less stable than HAP, they would never form in reality if it were not for their ability to grow at a comparatively high rate. HAP is a relatively hard crystal, ranking 5 on the Mohs scale, but despite that, as previously noted, it has an unusually low interfacial energy, which explains for the facility with which it nucleates in form of small particles in both biological and *in vitro* conditions. Had HAP had a high interfacial energy, a long persistence of these tiny crystals in both the biological and geological environments would not have been made possible. However, these other, thermodynamically less stable CAP phases have even lower surface energies, which is the reason why they nucleate even faster than HAP and thus often present intermediates leading to the eventual formation of HAP.

OCP crystals almost always come in plate-shaped (100) morphologies. Namely, (100) OCP faces are more hydrated than others and thus do not provide that favorable conditions for the attachment of PO_4^{3-} and Ca^{2+} ions as other phases. Consequently, the crystals grow slower along [100] axis, which results in (100) faces being the dominant in the final crystal morphologies. Also, (100) faces have the lowest interfacial energy, so that the crystal growth

proceeds in such a way to maximize their exposition on the crystal surface. This rule, which is in accordance with the Wulff construction¹⁶⁹, does not necessarily apply for biomineralized HAP for which, due to the growth regulated by organic adsorbents and matrices, the crystals often predominantly expose less thermodynamically stable faces. For example, the outer surface of the tooth enamel is composed of the smallest habit face, (001), which is more resistant to dissolution under acidic conditions than (100)¹⁷⁰. Still, since (100) faces are the principal ones on both OCP and HAP crystals, and since there are 2 of them on an OCP crystal and 6 of them on a HAP crystal, OCP should indeed have a plate-shaped or leafy morphology, whereas HAP should most optimally possess a hexagonal acicular morphology. However, when OCP acts as a precursor (an intermediate phase) for the formation of HAP, the final HAP particles retain the plate-shaped or acicular and fibrous morphologies in which (100) faces are dominant. Sometimes thus they retain forms of rosettes and fascicles with protruding blades or fibers. Intermediate phases are thus expected to play a crucial role in defining the morphology, interfacial properties and the growth mechanism of the HAP phase.

The first phase that should form upon precipitation of CAP is thus an amorphous phase with the stoichiometric formula of $\text{Ca}_9(\text{HPO}_4)_x(\text{PO}_4)_{6-x}(\text{OH})_x$ ($x < 0.5$). In contrast, notice that the stoichiometric formula of HAP is usually written as $\text{Ca}_{10-x}(\text{HPO}_4)_x(\text{PO}_4)_{6-x}(\text{OH})_{2-x}$ ($0 < x < 1$). The particles initially formed would actually be agglomerates of amorphous CAP units. Only in the following stage, the transformation of this phase into HAP and any other phase with $S > 1$ under the given conditions would take place. Under most of the conditions that resemble the physiological ones ($\text{pH} \approx 7.4$ and $T = 37^\circ\text{C}$), the identity of the precipitate changes over aging time (although OCP is accepted to be the transient phase in physiological conditions and at $\text{pH} < 9$, the mechanism is not clearly defined at higher pHs), eventually resulting in a stable and most often biphasic product (HAP/TCP). Solubility isotherms for DCPD and OCP at room temperature and for Ca/P molar ratio of 1.16 (resembling the Ca/P ratio within the initially precipitated amorphous phase) intersect at $\text{pH} 6.7$ (above this value, OCP is a more stable phase, and the trend is reversed at lower pHs), which implies that according to Ostwald-Lussac's rule¹⁷¹, the transformation of the amorphous phase to HAP should follow $\text{OCP} \rightarrow \text{DCPD} \rightarrow \text{HAP}$ route at $\text{pH} < 6.7$ and $\text{DCPD} \rightarrow \text{OCP} \rightarrow \text{HAP}$ route at $\text{pH} > 6.7$. Interestingly, magnetic field has been shown to accelerate the phase transformations along steps defined by the Ostwald-Lussac rule upon precipitation of CAP, also modifying ζ -potential of the particles¹⁷². The same effect was observed in the case of calcite^{173,174} and many other inorganic and organic diamagnetics, including proteins¹⁷⁵, where owing to the diamagnetic anisotropy of the crystalline order or the peptide bond an external magnetic field has an effect on precipitation and assembly thereof.

If the trend of pH change during this process is followed, it would be observed that the stage of formation of the amorphous phase is followed by a mild decrease in pH caused by the higher alkalinity of Ca^{2+} compared to the acidity of $\text{H}_x\text{PO}_4^{x-3}$. The subsequent formation of HAP is, however, entailed by a more significant drop in pH due to OH^- ions getting incorporated in the crystal lattice. The time span between mixing the reactants and the point when a more drastic pH drop is detected is considered as the induction time (a.k.a. nucleation lag time), τ , which is inversely proportional to S . Hence, the induction time is taken not as the time between the point of mixing the components and the formation of the first amorphous CAP particles, but as the time that passes before HAP starts forming, which is marked by the onset of the more significant pH drop (Fig.7). The third stage is typified by a pH drop rate with a similar slope as in the first phase, which is the sign of stabilization of the phase transformation. The slope eventually approaches zero, meaning that the stable phase has formed. All in all, the overall pH drop trend

is the one of sigmoidal shape. As expected, the induction period decreases with increasing the initial reagent concentrations, temperature and pH. However, pH change cannot be used to precisely estimate the rate of nucleation and crystal growth, primarily because of the inevitable presence of the buffering phosphate species. Even without their buffering activity, it is known that the nucleation rate increases with time (as $1 - \cos x$ function), from $t/\tau \approx 0.5$ to $t/\tau \approx 6$ when the constant nucleation rate is reached. This explains why the part of the pH vs. t curve where the onset of the pH drop is observed typically follows the trend of increasing its slope with time, all until the mother phase gets depleted of the growing units and the curve approaches the third stage of the process, that is, the one of mere rearrangement of the crystalline phase.

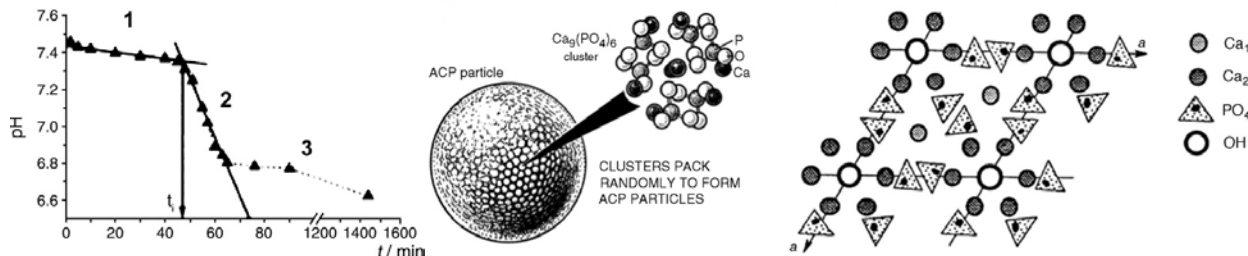


Fig.7. Three stages in the trend of pH drop following precipitation of CAP and formation of HAP (left), the presumed random packing of amorphous clusters into nanosized amorphous CAP (middle), and their rearrangement into symmetry that corresponds to HAP (right). Reprinted with permission from Refs. 176,177.

Hence, the process of formation of HAP in a precipitation reaction from a pure solution can be divided to following stages¹⁷⁸: i) homogeneous nucleation; ii) aggregation of primary amorphous CAP particles into typically spherical units; iii) aggregation of spheres into chain-like structures; iv) growth of these structures; v) secondary precipitation and phase transformation. The initially precipitated particles of the amorphous phase were observed to be round-shaped with 20-30 nm in size (although they can reach 120 nm in size)¹⁷⁹, but composed of smaller particles of 4 nm in size on average. It was also observed that an increase in the ripening time implied aggregation of spherical singlets and formation of needle-shaped CAP particles of about 20 nm in length¹⁸⁰. This aggregation model for precipitation of HAP (Fig.8) is in agreement with numerous literature reports. Matijević has pointed out that most microsized uniform colloidal particles form not through following the classical LaMer's model that refers to successive stages of nucleation and crystal growth, but by involving an aggregation of primarily precipitated units at a certain stage of the process^{181,182,183}. Microstructural investigations of fine particles obtained by precipitation thus in most cases reveal structures composed of aggregated subunits¹⁸⁴, and this mechanism of growth has been evidenced for numerous inorganic and organic systems alike^{185,186,187}. A recent high-resolution *in situ* observation of formation of fine particles has shown an intensive coalescence of nanoparticles during their growth¹⁸⁸. The structural nature of the primary units and their transformation pathways are, however, still the subject of disagreement among different research groups. The difficulty in assessing the structure of primarily precipitated units stems from the fact that owing to its transient nature, the amorphous CAP normally remains undetected unless it becomes stabilized by additives, such as rare-earth metal ions or peptides, for example.

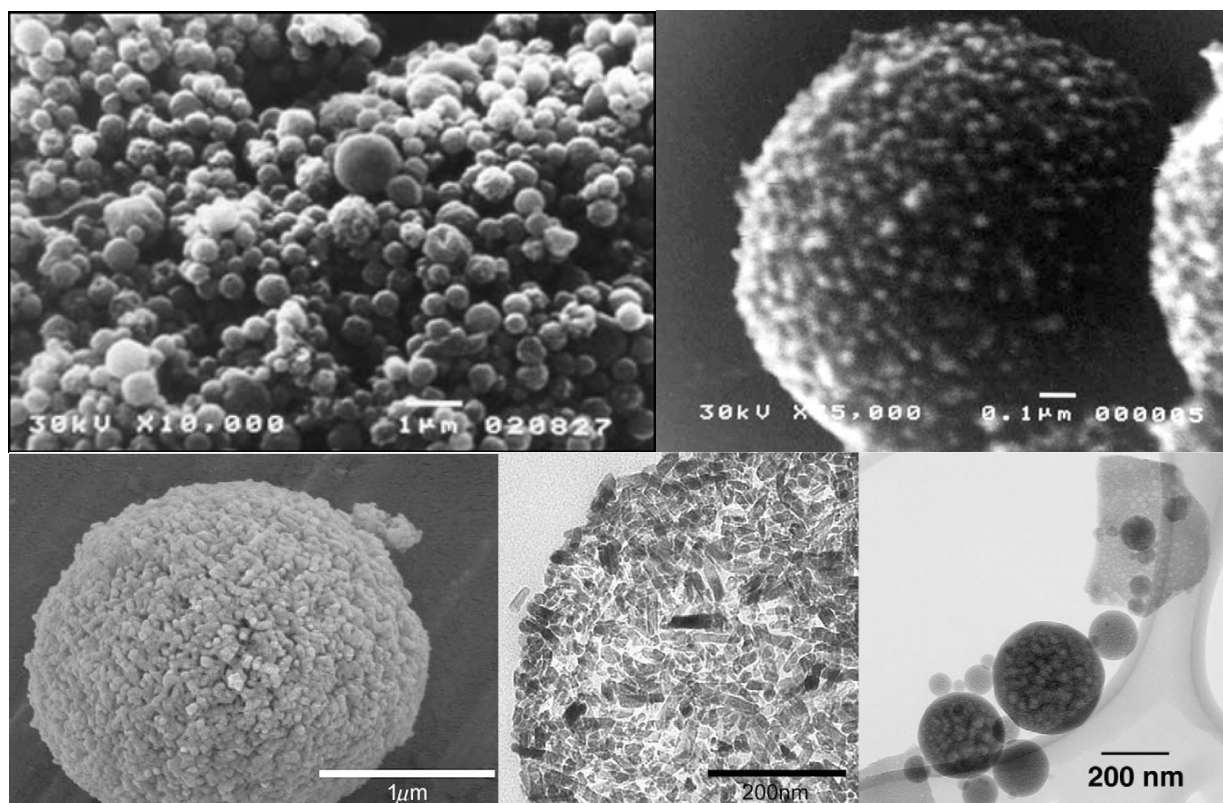


Fig.8. Uniform spherical HAP particles obtained using ultrasonic spray-pyrolytic processing (upper left), forming through aggregation of smaller spherical subunits (upper right). Reprinted with permission from Ref.189. Spray drying is another method that routinely leads to formation of microsized HAP particles through aggregation of nanosized subunits, and some of the morphologies are shown in images in the bottom row. Reprinted from Ref. 190.

However, the exact pathway of phase transformations following precipitation and leading to HAP as a final product is subject to change depending on the experimental conditions, including ionic concentrations, molar ratios of precursor ions, pH, T, the presence of additives, etc. Thus, for example, under given conditions that involved $[Ca] > 0.5$ M and pH 10 – 11, it was shown that OCP is the first phase that forms, transforming subsequently to amorphous CAP (which was observed to be the fastest step), which then transforms to DCP, which eventually transforms to HAP¹⁹¹. The process was sensitive to temperature, as 24 h was required to attain HAP at 25 °C, whereas the same process was over in only 5 min when taken place at 60 °C. When precipitation was carried out at 95 °C and at pH 10.6, the formation of HAP was so quick that all other phases were virtually undetectable. This may have been due to a sufficiently high heat content of the system, which enables rapid transcending of the stages dominated by intermediate phases. In support of this observation, (a) DSC/TGA analysis led to detection of only the adsorbed water loss peak; (b) XRD showed unmodified phase identity and no change in the lattice parameters (which would result from the loss of OH⁻ ions from the crystal lattice and the formation of vacancies during annealing) after heating up to 1100 °C; and (c) IR spectroscopy showed only bands at 3572 and 631 cm⁻¹ coming from stretching and vibrational modes of OH⁻ ions and those at 1087, 1032, 962, 601, 571 and 474 cm⁻¹ ascribed to PO₄³⁻ vibration modes, while the HPO₄²⁻-derived band at 875 cm⁻¹, CO₃²⁻ absorption at 1410 cm⁻¹ and weak shoulders at 990, 970 and 945 cm⁻¹ indicative of α- and β-TCP were absent¹⁹². In another one of the studies¹⁹³, any T < 95 °C required ripening periods extending up to 20 h to obtain pure

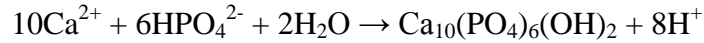
HAP. The size and crystallinity of HAP particles increased with increasing the precipitation temperature. Intuitively, this could be explained by assuming that the number of nuclei formed is proportional to S , which would be lower at a higher temperature. Smaller number of nuclei, on the other hand, implies the formation of larger particles. However, simple solubility product calculations can show that an increase in temperature corresponds to higher, not lower S , and the magnitude of change in S per $^{\circ}\text{C}$ is higher in the vicinity of the physiological conditions than in alkaline conditions with $\text{pH} > 10$ ¹⁹⁴. In fact, according to Le Châtelier's principle, an increase in temperature entails a rise in solubility in case of endothermic dissolution processes and a drop in solubility in case of exothermic ones. As the precipitation of CAP can be considered an endothermic process¹⁹⁵, an increase in temperature will yield lower solubility and, therefore, higher S . However, the fact that precipitation of HAP involves numerous transitions between phase intermediaries, some of which are exothermic and some of which are endothermic, predicting the temperature effect on the phase transformation pathways is difficult. The fact that these theoretical predictions (i.e., higher S and smaller particle size at higher temperatures) often deviates from experimental observations implies that the thermodynamic complexity of these phase transformation pathways may have a decisive effect on the morphology and size distribution of the precipitated CAP particles. The reason for failure of one such standard model may also be due to the effect of higher diffusion rates at higher heat contents of the solution offsetting the dependence of S on the temperature of the system. Namely, higher diffusion rates contribute to the ability of ions to find more energetically favorable positions in the crystal lattice, resulting in a better crystallinity.

The composition of the initial amorphous phase is known to depend on pH, and is approximately $\text{Ca}_3(\text{PO}_4)_2$ (Ca/P molar ratio = 1.5) at high pH values, whereas the Ca/P ratio approaches 1 at lower pHs¹⁹⁶. It is assumed that as the amorphous phase matures at an almost constant pH, its Ca/P molar ratio increases to values that correspond to HAP stoichiometry, which implies that the transformation process will take place faster at higher pHs. The rate of conversion of the amorphous CAP to HAP has been shown to increase with pH in the range of 7 – 10 and then to decrease at higher pH¹⁹⁷. Typically, measuring $[\text{Ca}^{2+}]$ and $[\text{H}_x\text{PO}_4^{x-3}]$ in the supernatant results in observing a drop in $[\text{Ca}^{2+}]$ that follows a sigmoidal trend, similar to the one of the aforementioned pH vs. t dependence, whereas $[\text{H}_x\text{PO}_4^{x-3}]$ increases yielding also a sigmoidal curve. The reason is that from the point of precipitation of amorphous CAP, the phase transformation proceeds with the shift in the Ca/P molar ratio of the precipitate from 1 – 1.5 to 1.667, implying a release of PO_4^{3-} ions into the solution over the course of ripening. This also implies that a decrease in the induction time can be related to a decrease in $[\text{HPO}_4^{2-}]$ and an increase in $[\text{CaPO}_4^-]$ in the solution, pointing to the former as a retarding and the latter as a promoting parameter of crystallization. It is, however, still not clear if this phase transformation takes place through an internal structural rearrangement of the metastable phase or by dissolution/re-precipitation. Ever since the dark line observed in the center of enamel crystals¹⁹⁸ was shown to be OCP phase, it has been speculated that it should be a remnant of the OCP phase composition of either the initially precipitated or the most dominant transitory phase^{199,200}. Claims that the precipitated apatite is normally a solid solution rather than a double salt of OCP and HAP may speak in favor of partial or complete transition between these CAP phases in the solid state. Brown *et al.* have thus proposed that the hydrolysis of a unit cell thick layer of OCP transforms the latter to a two unit cell thick layer of HAP, which may thus be the mechanism for $\text{OCP} \rightarrow \text{HAP}$ transformation²⁰¹. If that is so, the phase transformations would be both solution-mediated and controlled by the diffusion of ions within the lattice. A contrary argument is that

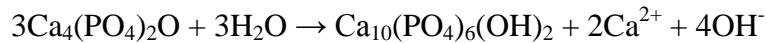
since the amorphous phase is a random network of bonds rather than a periodic structure, there could be no direct structural matching between the crystalline phases, implying that the phase transformation has to proceed via dissolution. If that is true, the process would be diffusion- or surface-reaction-controlled and consequently highly dependent on temperature and pH, which has indeed been experimentally confirmed. It was also shown that washing the precipitate at the stage where it is still amorphous increases the final Ca/P ratio due to removing the acidic layers of $H_xPO_4^{x-3}$ ions surrounding the growing particles (as already mentioned, it appears that $H_xPO_4^{x-3}$ ions are more effectively absorbed onto HAP particles than Ca^{2+} , and can be therefore used for manipulating their ζ -potential^{202,203} and thus controlling the efficiency of adsorption of biomolecules and cells thereto²⁰⁴), thereby validating the effect of exchange of ions between the solution and the precipitate on the final identity of the latter²⁰⁵. This can also be confirmed by the fact that during the ripening period, the precipitate could be isolated as a powder and analyzed for its structure without being subject to phase transitions in the solid state. If the presence of the liquid phase is required for the proper ripening, the transition has to at least partially follow a dissolution/recrystallization mechanism. That these phase transitions are solution-mediated can also be supported by another set of experiments in which it was observed that the amorphous-to-crystalline conversion starts sooner in the presence of more intensive stirring, as well as that the amorphous precipitate is prevented from undergoing the transition to crystalline CAP phases by placing it in ethanol or acetone instead of water²⁰⁶.

On the other hand, under low saturation conditions when the crystal formation occurs primarily through diffusion of ions onto already existing surfaces, the mechanism of nucleation can be substantially different in comparison with the one existing at high S when amorphous apatite is known to present the intermediate. In biological conditions, S is maintained at low levels for the purpose of letting the crystallization proceed as controlled by macromolecules and not by the thermodynamic propensities of the system. In view of that, the possibility that HAP forms directly upon a preexisting template cannot be discarded. Epitaxial effects were many times proven as essential in self-assembly procedures, and many biomineralization mechanisms (e.g., crystallization of thin flakes of nacre in the mollusk shells) inherently depend on the interfacial structural matching between an organic substrate and an inorganic phase. The driving force for the epitaxial growth is the tendency of the growing phase to approach the state of a minimal interfacial energy, which occurs when the two lattices precisely match each other. It is a general rule that a certain lattice mismatch in the overgrown phase (as compared with the ideal lattice parameters) occurs irrespective of the nature of the substrate (organic or inorganic), so that a certain flexibility in the overgrown phase is required for the epitaxial growth to be favored. HAP certainly presents one such phase as it can form even at Ca/P molar ratios as low as 1.33 (although Ca/P of biological HAP is normally in the range of 1.5 - 1.67) owing to a high structural flexibility and tolerance to defects, primarily in terms of calcium vacancies in the crystal lattice of HAP. HAP at such a low Ca/P molar ratio would require a removal of two calcium atoms from the stoichiometric formula and protonation of two PO_4^{3-} groups (i.e., $Ca_8(HPO_4)_2(PO_4)_4$ with completely empty halide and hydroxyl columns and the hexagonal structure preserved, which is required to maintain the charge neutrality)²⁰⁷. Biomineralization processes are, in fact, based on using only a few inorganic minerals, but endowed with exceptional structural flexibilities, such as calcite, silica or HAP, so that they could be shaped into materials with a wide variety of different properties²⁰⁸. However, there is one problem tied with the hypothesized immediate formation of HAP during its precipitation in biological

environments, and it is a large local drop in pH following its formation. Namely, for each unit cell of HAP formed, 8 protons are released into the solution:



This number may, however, be lower depending on the amount of carbonate (or other ions present in the solution) incorporated in the lattice and whether hydrogen or dihydrogen phosphates, as dominant in the physiological pH range of biomineralization, are consumed in the process as precursor phosphate ions. Also, phosphate and carbonate species as well as proteins in the extracellular matrix in which the mineralization proceeds may act as adequate buffers to mitigate the release of protons following the formation of HAP if the process proceeds at a slow pace. Nevertheless, HAP is the second most basic CAP phase (TTCP being the only more basic phase thereof), and the intensive pH drop following its direct formation still may be harmful for the surrounding tissues. Brown *et al.* have thus claimed that TTCP, a rarely mentioned CAP phase, may present an intermediate in the formation of biological HAP²⁰⁹. However, they have also observed that its immediate transformation to HAP would produce a significant increase in pH due to release of OH⁻ groups. Namely, the TTCP→HAP transition via hydrolysis of TTCP is entailed by a rise in pH:



The formation of TTCP as one of the intermediary phases, however, implies that pH does not necessarily need to drop during the formation of HAP. For example, if TTCP happens to be mixed with some of the more acidic CAP phases, such as DCPD, DCPA or MCPM, the transformation reaction may proceed without drastic changes in pH of the medium and, therefore, without imposing chemical stress on the adjacent tissues. The latter mechanism could be supported by the fact that solubility curves for TTCP and DCPA closely match at around pH 7.6. From this observation, it can be concluded that the complexity of phase transformations during the formation of HAP is enormously high and still largely open to exploration, scrutiny and dispute.

During the TTCP→HAP transformation, pH can climb up to almost 12 before eventually descending down to close to neutral or even acidic values. The magnitude of this increase was shown to depend on whether stoichiometric or calcium-deficient HAP forms, with the pH increase being more pronounced in the former case. The trend is, as expected, dependent on the temperature, owing to large activation energies intrinsic to these reactions. It is interesting that concentrations of Ca²⁺ and H_xPO₄^{x-3} ions in the solution are also prone to exhibit humps following complex transformations for which it is still not clear whether they take place in the solid state or entail dissolution/recrystallization pathways²¹⁰. Incorporation of CO₃²⁻ into HAP crystal lattice is another factor contributing to the decrease in pH, and that especially when the reaction is carried out under atmospheric conditions. This clearly brings us to the task of discerning the influence of dissolved gases on the precipitation processes. It is known that precipitation of numerous ceramics crucially depends on the presence of oxygen and the oxidation reactions brought about in its presence²¹¹. In view of that, it is also worth recalling that atomic aggregates do normally present crystallization nuclei (and 1.2 nm is usually considered as the critical size after which an unstable embryo becomes able to transcend the free energy barrier and continue to grow by diffusion and/or ripening), this is not necessarily the case. This role is

sometimes fulfilled by individual molecules in the excited or ionized state, such as in the case of formation of water droplets from supersaturated vapor. Likewise, the nucleation process during precipitation of ferrites from an alkaline solution is supposed to be centered around the Fe(II) → Fe(III) oxidation process^{212,213}. Gas bubbles are proposed as acting as electrostatic carriers of the incompletely understood long-range attraction forces between hydrophobic surfaces^{214,215}, and their full range of influence, similarly to that of ions in Hofmeister series in terms of their effect on protein aggregation and assembly, is still unknown^{216,217}. The fact that small differences in the gaseous content can trigger significant ones in the experimental outcomes is quite intriguing, and yet often omitted from serious chemical analyses²¹⁸. The effect of the nucleation surface, which is often disguised in terms of the reaction vessel walls, is another frequently neglected factor that, however, sometimes crucially defines the morphological propensities of the particles. In that sense, there is the example of BaSO₄/BaCrO₄ nanofilamentous superstructures formed after their self-assembly either on glass walls or TEM grids, but when the identical procedure was attempted in plastic bottles, the crystallization experiments failed²¹⁹. Many similar examples wherein changes in the vessel geometry or batch size significantly altered the experimental outcomes may be found in the literature²²⁰. Minor changes in the rate at which the reactants are mixed or introduced to the mixture can often have a drastic effect on the identity of the precipitate. Namely, upon abrupt mixing, even though S may in theory correspond to a metastable supersaturated state, locally produced high reactant concentrations may lead to irreversible formation of nuclei and a premature precipitation under uncontrolled local conditions (wherein pH and ionic activities may significantly differ from measured or calculated bulk values²²¹). It is known that texture and porosity of the external substance are important as they can heavily influence the HAP nucleation efficiency²²². This probably explains why Tadashi Kokubo, the inventor of the method for evaluating the bioactivity of a compound by immersing it in SBF and awaiting the formation of HAP, suggested discarding any plastic bottles with visible scratches and never reusing them for storing SBF²²³. Namely, SBF is a metastable solution with S with respect to HAP normalized per growth unit equivalent to 19.5, and to avoid premature crashing of one such solution storing it in a vessel with smooth wall surface and without any particulate remnants of formerly formed precipitates therein stands forth as of vital importance for reliable bioactivity analyses. A metastable solution, which could be in theory stable throughout geological times, may often turn into an immediately crashing solution if the slightest amount of impurities is introduced to it, as in accordance with the more energetically favorable heterogeneous nucleation compared to the homogeneous. In fact, it is claimed that CAP is sensitive to impurities of less than 10⁻⁶ % (0.01 ppm); that impurity molecules present condensation centers for the formation of amorphous CAP; and that they as such present the basis for the primary particle formation during crystallization of HAP²²⁴. The kinetics of enamel dissolution has also been found to be strongly influenced by the presence of impurities released to the solution during the reaction²²⁵. In view of this, depending on minor effects of impurities or changes in the experimental conditions, an unexpected pH vs. t trend can be triggered, signifying deviations from the standard precipitation pathways. Solutions that are particularly prone to exhibit this sensitivity are those that lie towards the middle of the metastability zone, between the levels of saturation (S = 1) and critical supersaturation (S > 19.5).

One of the ways to assess the metastability of CAP solutions and the extent of the nucleation promoting/inhibiting effect of additives is to measure the aforementioned nucleation induction time of the solutions at different saturations and different concentrations of additives.

Since a direct measurement of the nucleation rate is difficult, one of the most common ways to assess the kinetics of nucleation is to measure the induction time at different supersaturations.

Expressions for the inverse of the induction time for nucleation, τ , and for the nucleation density, N , are given by classical nucleation theory as:

$$1/\tau = \Omega \exp(\Delta G^*/kT) = A \exp(-\beta v^2 \gamma^3 / (kT)^3 (\ln S)^2)$$

$$N = B \exp(-\beta v^2 \gamma^3 / (kT)^3 (\ln S)^2)$$

ΔG^* is the activation energy for nucleation, Ω is a preexponential factor, γ is the interfacial tension for the formation of the critical nucleus (typically comprising 10 – 100 atoms), S is the supersaturation ratio of the solution, β is a shape factor, v is the molecular volume, k is the Boltzmann constant, T is temperature, and A and B are constants. Induction time is the consequence of the activation energy required for nucleation, ΔG^* , which depends on the interfacial tension for formation of the critical nucleus, γ , and S . From the equations above, we could see that the induction time is reduced and the nucleation density is increased by lowering the interfacial tension, γ , of a CAP solution. For example, promotion of heterogeneous nucleation of CAP in the presence of another solid surface indicates that this other phase reduces the CAP solution interfacial tension compared to homogeneous nucleation of CAP in the absence of the additive. The equations above also predict that nucleation is promoted by increasing S . They also indicate that systems with higher nucleation densities, leading to smaller particles, should also have shorter induction times. The amount and nature of precipitated CAP also varies with the concentration of the additive. Higher amounts of a peptide additive may result in higher degrees of binding of Ca^{2+} and $\text{H}_x\text{PO}_4^{x-3}$ ions, causing depletion of species from the solution, lowering S and reducing the driving force for nucleation. Certain additives may also have opposite effects on nucleation depending on their concentration. It was thus pointed out that the concentration of amelogenin, the main protein of the developing enamel matrix, has a directly proportional effect on the amount of CAP precipitated and is inversely proportional to the induction time at low concentrations²²⁶. There is, however, evidence that this trend becomes reversed at high concentrations, when amelogenin starts acting as a nucleation inhibitor²²⁷.

Furthermore, in case of the classical heterogeneous nucleation, the nucleation rate is equal to:

$$J = A \exp(-\Delta G/kT) = A \exp(-16\pi v^2 \gamma^3 f(\theta) / 3k^3 T^3 (\ln S)^2)$$

Molecular volumes defined by the crystal structure, v , for HAP, OCP and DCP equal 263.24, 310.59 and 126.53 \AA^3 , respectively; γ was estimated as $\sim 40 \text{ mJ/m}^2$ at the interface between a growing HAP phase and a metastable supersaturated solution²²⁸; $f(\theta)$ is the contact angle function for a nucleus on a substrate; $(16\pi/3)$ is the geometric factor, corresponding to spherical nucleus, and is different for other nucleus shapes (32 for cubic, e.g.).

So we see that since $\exp(-1/\ln S^2) = S^2$, both the nucleation density and the nucleation rate are proportional to S^2 , whereas the crystal growth rate, J , is proportional to S^x . It can be represented as equal to kS^x , where k is the rate constant and x is the reaction order. Different reaction orders correspond to different mechanisms of crystal growth with 4 main scenarios: (a) mass transport and diffusion-limited growth at very high levels of S ($x = 1$); (b) polynucleation of surface growth islands at high S ($x > 2$); (c) layer-by-layer growth at moderate S ($x = 1$); and

(d) screw dislocation growth at low S ($x = 2$). Except for the case of $x > 2$, all these mechanisms are valid for both growth and dissolution. For diffusion-limited growth, typically taking place at high S , $x = 1$. This implies that in a relatively high S range, an increase in S would lead to the nucleation rate increasing more than the crystal growth rate does. This explains why the smaller the particles one tends to obtain, the conditions of higher S one should produce. More nuclei are then produced, and consequently the final particles will be smaller in size. In contrast, in order to obtain a monocrystal, a single nucleation event has to be produced in the solution, followed by the crystal growth; consequently, conditions with a low S need to be set.

Crystallization of HAP at low S , with or without the presence of an organic phase, is shown to be a second-order reaction, corresponding to a surface-diffusion-controlled spiral growth mechanism²²⁹. The fact that stirring of the reaction mixture does not typically produce any effect on the crystal growth rate excludes the possibility that the rate-determining step under low S could be the bulk diffusion of ions. At very low concentrations, additive molecules cannot influence the volume transport processes and their effect, therefore, must be due to their participation in the surface reactions in the adsorption layer. However, sometimes at low concentrations and that particularly when polynucleation mechanism of growth takes place, additives may promote nucleation by increasing its rate. On those occasions, additive molecules may be adsorbed specifically on certain sites of the crystal surface (in general, their binding will be strongest at kinks, moderate at steps, and weakest on crystal terraces), so that bridges are formed, wherein the additive molecules may transport the growth units more efficiently to the crystal surface. This mechanism bears resemblance to the one describing the formation of silicon nanowires in the vapor-liquid-solid (VLS) process, during which nanodroplets of gold deposited on top of silicon wafers attract silicon atoms from the vapor, and after becoming supersaturated with respect to silicon begin to precipitate it, building well-aligned nanowires oriented perpendicular to the underlying surface^{230,231}. That the same additive may promote nucleation at low surface coverage and inhibit it at higher concentrations has been explained by hypothesizing lowering of the surface energy following absorption whenever the nucleation-promoting effect takes place. However, as pointed out by Zhang and Nancollas, "Since the surface underneath covered by additive molecules can no longer participate in the formation of surface nuclei, this explanation for rate promotion is merely phenomenological"²³². On the other hand, at higher concentrations, the adsorbent layers are expected to hinder the delivery of growing units from the solution onto the growing sites on the crystal surface²³³.

Finally, it is worth keeping in mind that the conceptual framework for analyzing CAP phase transitions based on supersaturation is an approach that possesses fundamental limitations. Related to free energy of the dissolution reaction ($\Delta G_s = -RT\ln K_{sp}$), K_{sp} is a thermodynamic property. ΔG of the precipitation reaction is equal to: $\Delta G = -RT\ln S$, which implies that the larger the S , the greater the driving force for precipitation. This implies that solution speciation in general is a thermodynamic consideration. Consequently, whenever kinetic effects take place, solubility product fails to provide a good basis for describing the precipitation process. The effects of the interaction of ions and small particles with organic molecules present such cases. As the result, the effects of ion association and complexation in the solution may significantly lower the free ion concentrations compared to the calculated values.

In addition, calculating ionic activities that figure in equations for S and K_{sp} is always an approximation owing to the following effects. Firstly, all the additional species (ions, aliphatic additives and macromolecules, etc.) can influence the solubility of the given compound, even though they may not participate in the reactions. Common ion effect is the consequence of Le

Châtelier's principle, and according to it, a salt will be less soluble if one of its constituent ions is already present in the solution. Uncommon ions tend to increase solubility by increasing ionic strength of the solution, which is usually referred to as the salt effect or the diverse ion effect (Fig.3). Complexation can also modify the amount of free ions in the solution (that is, their activities). This is particularly the case for biological fluids where complexation reactions involving many organic species in the solution may make ionic concentrations markedly deviate from the resulting ionic activities. Also, in case of purely inorganic systems, the substantial complex formation introduces additional limitations in the application of the concept of solubility product.

The assumption that all the oppositely charged ions in solution are thoroughly hydrated and separate is not valid. The effects of ion association and complexation in the solution may significantly lower the free ion concentrations compared to the calculated values. Also, the equilibrium described by K_{sp} refers not to all chemical compositions of the solid phase, but to a specific phase. Hence, calcite, aragonite and vaterite are all described with the same chemical formula (CaCO_3) and yet have different K_{sp} . Even so, it is only one phase that in most cases enters the equilibrium. Then, K_{sp} is defined for large monocrystals for which surface area effects are negligible. However, solubility increases with decreasing the particle size and once particles become smaller than 1 μm in diameter, this effect has to be accounted for. Specific structuring of water molecules around solutes and colloidal species also affects the thermodynamics of phase transitions in solution. For example, the release or trapping of water following the formation of a solid has been shown to modify the energy landscape of initial and final free energy states of most precipitation reactions²³⁴. On the other hand, just as each self-assembly event implies a co-assembly of the immediate environment surrounding the assembling system²³⁵, each crystallization event can be seen as a process of a simultaneous rearrangement and restructuring of the interfacial water molecules. From one such point of view, the kinetic attributes of the latter process can be considered as a crucial factor in determining the crystal formation rate.

Note also that in reality every phase transition follows a non-classical model. Whereas the classical nucleation model predicts that the nucleation rate should continuously increase with S , the non-classical model is built on the assumption that there are two antagonistic effects taking place. Namely, the higher the S , the larger will be the thermodynamic driving force for nucleation, inducing the latter to proceed at a higher rate (although this rate normally decreases as nucleation proceeds due to depletion of growing units from the mother phase). But on the other hand, moving the system state away from equilibrium modifies the transfer of matter between the growing embryos and the mother phase, and since this transfer has to be ensured for the nucleation to proceed, this shift can significantly delay nucleation in some cases. In case of the glass formation, for example, an ultrafast cooling promotes the latter effect, leaving the system in the metastable state with an extremely slow transformation to the stable, crystalline form. If one would make an attempt to condense water vapor at the temperature of liquid nitrogen, at first it may seem that such a transition would proceed momentarily. But it is not so²³⁶. Namely, at such a low temperature, the equilibrium pressure of water vapor (p_0) is so low that one collision between its molecules (which is the first step to the formation of an embryo that then has to advance forming a stable nucleus or to simply dissipate) occurs every 10^{16} years. To induce nucleation, one would have to increase supersaturation ratio (p/p_0) to $\sim 10^{12}$ by compressing the system and increasing its concentration. In contrast, condensation of water vapor at atmospheric conditions occurs in the supersaturation range $p/p_0 = 5 - 8$. As we see, this antagonistic effect implied by the fact that the mother phase has to ensure a reasonable supply of

matter to the embryo for nucleation to proceed becomes most critical in far-from-equilibrium conditions, although complex kinetic conditions within a system may sometimes predispose it to exhibit such far-from-equilibrium characteristics even though its state lies close to equilibrium. The fact that nucleation rate is subject to variations, both before and after the critical point is reached, and depends on the system in question, contributes to difficult predictions of nucleation and crystal growth rates in any real systems.

1.4. Characterization of structural properties

As complementary to various microscopic techniques used to visualize morphology and structure of CAP particles at micro and nano scales, spectroscopic techniques, such as X-ray diffraction (XRD), are routinely applied to study the phase composition of CAP precipitates. Rietveld analysis can be used to detect the shift of diffraction peaks along the 2θ axis compared with the stoichiometric values. This shift is proportional to the lattice distortion (that is, a change in the unit cell constant), and can be correlated with Ca deficiency, for example. Also, the intensity ratio between different peaks can be used to study the crystallographic orientation for a given morphology. For example, if (100), (200) and (300) reflections are more intense than those of a standard HAP, it signifies that the particles (fibers in this case) grew along the c -axis, that is, with the preferred orientation along the $\{h00\}$ planes. Even the morphological difference between needles and bars of HAP could be thus clearly read on the corresponding XRD patterns²³⁷. The specificity of orientation of crystals on a given substrate, particularly interesting for HAP growth experiments that aspire to result in enamel-like structures ($f = 0.86$), can also be estimated from an XRD analysis by calculating the Lotgering orientation factor, $f = (P - P_o)/(1 - P_o)$, where $P = I(00l)/I(hkl)$ of the sample and $P_o = I(00l)/I(hkl)$ of the standard²³⁸. The crystallite size can be calculated from the broadening of (002), (222) and (300) diffraction peaks using the Debye-Scherrer equation, assuming that HAP crystals are prism-shaped with the height equal to the crystallite size along (002) plane (c plane) and the length corresponding to the crystallite size along (300) plane (a plane)²³⁹. The main diffraction peaks for DCPA, α - and β -TCP, DCPD, CAP, and OCP are listed in Table 2, whereas those for stoichiometric HAP are shown in Table 3.

Table 2. The main X-ray diffraction peaks for the selected CAP phases. Adapted and partially reproduced from Ref. 240.

Most Important Peaks in the X-Ray Patterns of DCPA, α and β -TCP, DCPD, CAP, and OCP																	
DCPA ^a			α -TCP ^b			β -TCP ^c			DCPD ^d			CAP ^e			OCP ^f		
d (Å)	I/I_1 %	Miller Indices	d (Å)	I/I_1 %	Miller Indices	d (Å)	I/I_1 %	Miller Indices	d (Å)	I/I_1 %	Miller Indices	d (Å)	I/I_1 %	Miller Indices	d (Å)	I/I_1 %	Miller Indices
6.74	13	(010), (1 $\bar{1}$ 0)	7.31	25	(111)	6.49	15	(006)	7.57	100	(020)	3.46	25	(002)	18.68	300	(100)
3.48	13	(1 $\bar{2}$ 1)	4.00	20	(150)	5.21	20	(110)	4.24	100	(021)	2.78	100	(211), (112)	9.36	45	(200)
3.37	70	020	3.91	40	(202)	4.05	15	(204)	3.05	75	(111), (041)	2.68	40	(300)	9.05	40	
3.35	75	(110), (2 $\bar{2}$ 0)	3.88	40	(241)	3.45	25	(1.1.10)	2.93	50	(22 $\bar{1}$)	2.23	16	(310)	5.52	25	
3.33	17	(2 $\bar{1}$ 0)	3.69	40	(132)	3.21	55	(214)	2.62	50	(220), (15 $\bar{1}$)	1.93	16	(232), (401)	3.66	30	
3.13	20	(11 $\bar{1}$)	3.66	17	(151), (222)	3.01	15	(300)	2.60	30	(20 $\bar{2}$)	1.84	16	(213)	3.44	60	
2.96	100	(1 $\bar{2}$ 2), (1 $\bar{1}$ 2)	3.01	20	(510)	2.88	100	(2.1.10), (217)	2.17	20	(151)				3.42	50	
2.94	35	(10 $\bar{2}$)	2.95	20	(113)	2.76	20	(218)	2.15	17	(24 $\bar{2}$)				3.21	25	
2.75	20	(2 $\bar{3}$ 0)	2.92	35	(402), (023)	2.61	65	(220)	1.82	20	(241)				2.87	30	
2.72	35	(200), (102)	2.91	100	(441), (170)	1.93	20	(4.1.10), (327)							2.83	100	(260)
2.25	15	(030)	2.86	30	(511)	1.90	15	(328)							2.82	95	(320), (241)
2.50	15	(2 $\bar{3}$ 1), (02 $\bar{2}$)	2.62	50	(043), (352)	1.73	25	(2.2.20)							2.78	45	
2.20	13	(003)	2.59	30	(080)										2.75	35	
															2.71	25	
															2.67	50	(700)
															2.64	35	

Table 3. The main X-ray diffraction peaks for stoichiometric HAP, according to ASTM Card No. 9-432.

List of the Peaks Observed in the X-Ray Powder Diffraction Pattern of HAP and the Respective Data for Stoichiometric HAP

A.S.T.M. Card No. 9-432			
$2 \cdot \theta$ (°)	I/I_1 %	d (Å)	Miller Index of the Corresponding Reflection
16.84	5	5.250	(101)
18.78	3	4.720	(110)
21.60	9	4.070	(200)
22.97	9	3.880	(111)
	1	3.510	(201)
25.90	40	3.440	(002)
28.22	11	3.170	(102)
29.14	17	3.080	(210)
31.86	100	2.814	(211)
32.20	60	2.778	(112)
32.90	60	2.720	(300)
34.22	25	2.631	(202)
35.51	5	2.528	(301)
39.26	7	2.296	(212)
39.86	20	2.262	(310)
	1	2.228	(221)
42.10	9	2.148	(311)
	3	2.134	(302)
43.80	7	2.065	(113)
	1	2.040	(400)
45.32	5	2.000	(203)
46.69	30	1.943	(222)
48.16	15	1.890	(312)
48.59	5	1.871	(320)
49.51	40	1.841	(213)
50.53	20	1.806	(321)
51.33	11	1.780	(410)
52.24	15	1.754	(402), (303)
53.27	20	1.722	(004), (411)
54.45	3	1.684	(104)
55.87	9	1.644	(322), (223)
57.15	7	1.611	(313)
58.17	3	1.587	(501), (204)
60.01	5	1.542	(420)
60.45	5	1.530	(331)
61.66	9	1.503	(214), (421)
63.07	11	1.474	(502)
	3	1.465	(510)

Infrared (IR) and Raman spectroscopies are often used to qualitatively detect and discern CAP phases, although with much less precision than XRD (Fig.9). IR and Raman spectroscopies are, on the other hand, particularly useful for studying the structure of inorganic/organic composites, owing to a facile detection of bonds within organic molecules as well as a high sensitivity of the position of the bands depending on the local environment surrounding the given vibration. However, unlike IR spectroscopy where a large absorbance of water poses obstacles in probing precipitates in their aqueous environments, Raman spectroscopy does not suffer from this problem. As in XRD analyses, the bandwidth can be used as an indicator of crystallinity and the amount of defects of the material in the sense that wider peaks indicate lower short-range order. Since vibrational frequencies of stretching vibrations are correlated to bond length and bond strength, frequency shifts of IR/Raman bands can be used to estimate changes in the local environment surrounding the active species²⁴¹. As a rule, hydrogen bonding decreases the frequency of stretching vibrations, as it lowers the restoring force, but increases the frequency of bending vibrations.

The main Raman band for HAP is $\nu_1(\text{PO}_4)$ at 960 cm^{-1} , deriving from a totally symmetric, non-degenerated stretching mode of the “free” tetrahedral phosphate ion. The other three phosphate peaks are found at 400-500 (ν_2), 550-650 (ν_4), and 995-1120 cm^{-1} (ν_3). The latter band is particularly interesting because it presents the key for discerning HAP from OCP, the two CAP phases that show a tremendous resemblance in their IR and Raman spectra^{242,243}. Namely, unlike HAP, OCP exhibits a peak at $\sim 1015\text{ cm}^{-1}$ positioned between $\nu_1(\text{PO}_4)$ and the main doublet of ν_3 bands with peaks at 1045 and 1075 cm^{-1} . The main CO_3^{2-} peak is at $\sim 1080\text{ cm}^{-1}$ or $\sim 1110\text{ cm}^{-1}$, depending on whether the fully or partially carbonated HAP is of B-type or A-type, respectively²⁴⁴. This peak, can, however, overlap with the main triply degenerated asymmetric stretching mode vibration²⁴⁵ of the P-O bond, ν_3 , at 1087 cm^{-1} . The other two components of this vibration are said to appear at 1046 and 1032 cm^{-1} , whereas the weak peak at 472 cm^{-1} with the shoulder at 462 cm^{-1} belongs to the doubly degenerated bending mode of PO_4 group, ν_2 , and the peaks at 601, 575 and 561 cm^{-1} are assigned to a triply degenerated bending mode, ν_4 , of the O-P-O bond. In addition, IR spectroscopy could be used to detect CO_3^{2-} ions in the apatite structure. The CO_3^{2-} bands appear at 755 cm^{-1} for the ν_4 stretching mode, 872 cm^{-1} with the shoulder at 880 cm^{-1} for the ν_2 stretching mode, and at 1418, 1456, 1506 and 1558 cm^{-1} for the ν_3 stretching mode. The bands at 872, 1418 and 1456 cm^{-1} indicate the B-type of carbonated HAP, while those at 880 and 1558 cm^{-1} are typical of the A-type^{246,247}. The intensity ratio between 880 and 872 cm^{-1} bands (I_{880}/I_{872}) can be used to determine the ratio between CO_3^{2-} substitutions of OH (A-type) and PO_4^{3-} (B-type) groups in carbonated HAP²⁴⁸. On the other hand, alterations in the HAP lattice parameters could be used to detect the presence of carbonate and other additives from XRD. Precisely, the incorporation of the CO_3^{2-} ions in the hexagonal channel of the apatite structure (A-type HAP) leads to an increase in the lattice parameter a and a markedly lesser decrease in the parameter c in comparison with the stoichiometric HAP. In contrast, the parameter a decreases while c increases with a partial substitution of PO_4^{3-} by smaller CO_3^{2-} ions (B-type HAP).²⁴⁹ XRD is thus ideally used to detect phase purity, whereas X-ray fluorescence proves convenient for detecting Ca/P molar ratio and content of other additives or impurities. Incorporation of CO_3^{2-} ions in the crystal lattice of HAP leads to reduction of symmetry, a decrease in the crystallite size and increased degree of distortion, thereby inducing an increase of the band width and a shift of the $\nu_1(\text{PO}_4)$ band to lower wavenumbers. If $\nu_1(\text{PO}_4)$ peak at 960 cm^{-1} is shifted to $\sim 965\text{ cm}^{-1}$, it signifies the presence of non-carbonated, pure HAP, whereas a large shoulder at 950 cm^{-1} signifies amorphous CAP. Hence, broadening of this spectral line can be calculated to estimate the proportion of carbonated apatite in a HAP sample. For β -TCP, the symmetric stretching band is split to a peak at 950 and a peak at 970 cm^{-1} . This splitting that reflects the difference in the intratetrahedral P-O bond lengths for the different nonequivalent PO_4^{3-} ions of β -TCP structure is much smaller for HAP and FAP. Lattice modes for HAP are present at $< 350\text{ cm}^{-1}$, and $\nu_1(\text{OH})$ is present at 3575 cm^{-1} .

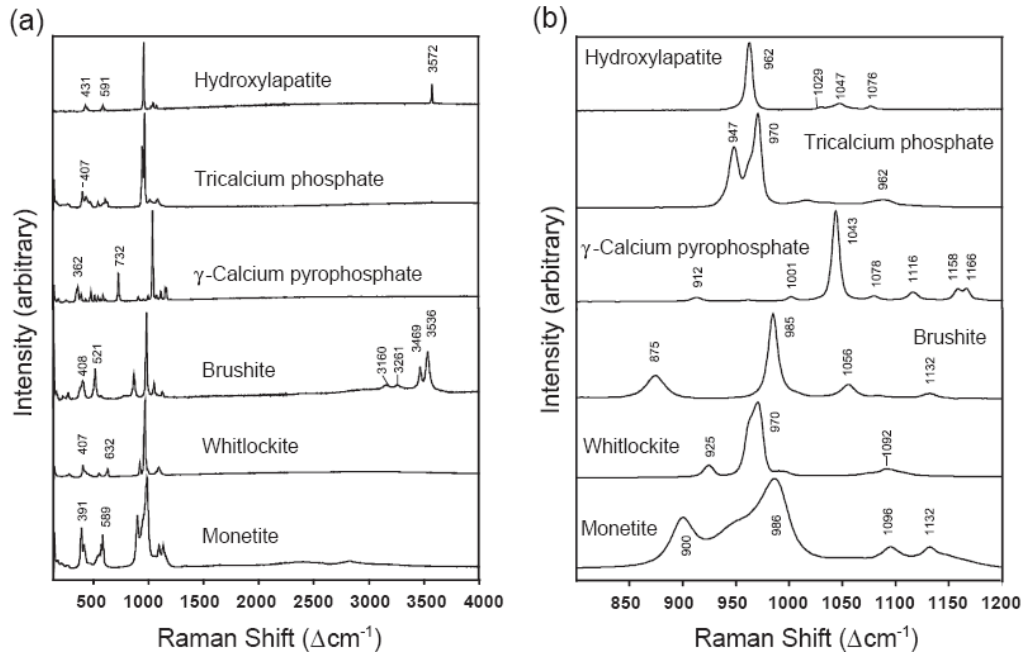


Fig.9. Typical Raman spectra for the main CAP phases. All visible peaks are caused by P-O stretching and bending modes of vibration. The 800 – 1200 cm^{-1} wavenumber area magnified on the right shows only stretching P-O modes. As a consequence of different crystal symmetries of the different CAP phases, the P-O vibrations in each one of them are surrounded by a specific atomic environment, which causes a specific shift of the corresponding bands along y-axis. This shift can be used to identify the given phases. Reprinted with permission from Ref.250.

2. Application

Owing to their being the elementary constituent of hard tissues, CAP materials have been so far mostly used for biomedical purposes, that is, for the substitution of damaged hard tissues. The application of HAP, however, extends beyond the scope of biomedicine. For example, HAP has been applied as an adsorbent in chromatography for separation of proteins and DNA^{251,252}. In nanosized form, it has been applied as a stimulus-responsive stabilizer in Pickering emulsions²⁵³, following the observation that colloidal CAP in milk is involved in stabilizing casein micelles²⁵⁴. It has also been used as a nutritional supplement and a raising agent in the food industry. In the field of fertilizers, HAP has been used as the essential ingredient of dialysis pouch systems for maintaining concentration of phosphates in the plant root environment²⁵⁵. In the context of water purification processes, both synthetic and biological HAPs have been applied for the removal of heavy ions²⁵⁶. However, although CAP materials have been used for a variety of purposes, the main area of their application is still medicine, and this is why this part of our discussion will focus on this particular field.

2.1. The promises of nanomedicine

It has been shown that an increasing ability of humans to control physical phenomena at ever finer scales implies the trend of continual miniaturization of electronic devices and the eventual “descent” of human technologies into the nano range²⁵⁷. Consequently, nanoscale tools and devices are expected to increasingly find usage in numerous aspects of the modern society. The modern medicine is thus only one among the fields that is currently undergoing a transition

towards ever finer and more sensitive methodologies in terms of finely targeted and superiorly efficient delivery of pharmaceutical agents that minimizes the side effects of medical treatments²⁵⁸.

Many eyes are thus focused on the promises of nanomedicine (Fig.10), which can be roughly defined as the application of physical chemistry of nanoscale phenomena for biomedical purposes²⁵⁹. Although only a few nanoparticle-based drugs have so far been approved by the FDA, it is estimated that the impingement of nanomedicines on the modern society will only increase in the coming years²⁶⁰. Also, with approaching the patent expiration times, the biotech industries are intensively seeking new competitive business strategies, and nanomedicine is often mentioned as one of the key focuses. In view of their offering solutions to fundamental pharmaceutical problems ranging from poor solubility of drugs to the lack of target specificity, nanotechnologies are viewed as a potential revolution in the world of medicine.

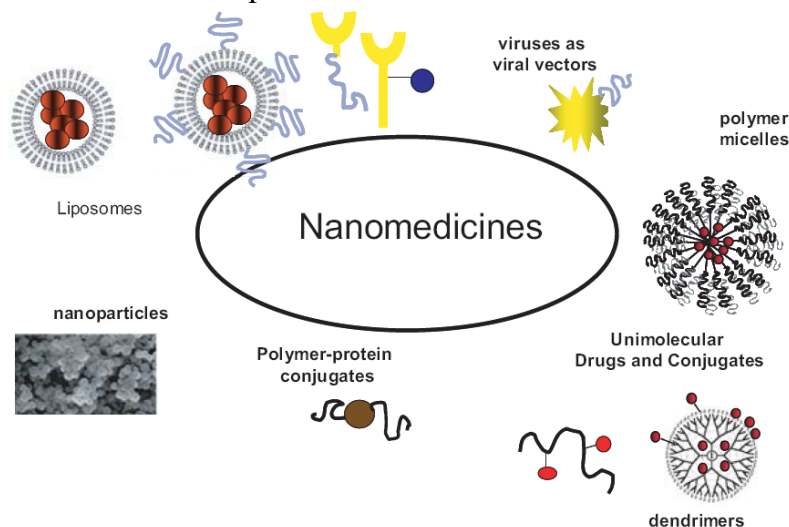


Fig.10. A few of the major contemporary candidates for drug and gene delivery carriers. Reprinted with permission from Ref.181.

A particular emphasis in biomedical research is, therefore, placed on nanosized drug delivery carriers for either local or systemic use. Many biopharmaceuticals have limited long-term structural stability when formulated at high concentrations, which limits their shelf-life. In addition, many are susceptible to biodegradation when delivered, which in addition to their frequently extensive hydrophobicity, reduces their bioavailability. For localized treatments, the delivery vehicle is meant to be retained at the delivery site, ensuring the local and rate-controlled administration of the therapeutic. For therapeutics delivered systemically, through the vasculature, the nanoparticulate carrier is expected to: a) prolong the circulation time in plasma; b) enable carrying of hydrophobic drugs to their destination in the body, thus preventing their clumping in the blood and increasing their bioavailability; and c) through a proper mechanism (such as surface functionalization or guiding via an external field), it may direct the therapeutic to a desired tissue.

The majority of nanomedicines on the market are still merely pharmaceuticals that are formulated into nanosized structures in order to manipulate the pharmacodynamics, biodistribution and overall effectiveness of the drug²⁶¹. The main purpose of decreasing the particle size of poorly water-soluble drugs down to nanoscale is an accelerated absorption and a higher efficacy produced thereby. This results from the fact that the solubility and dissolution

rate of a drug can be increased by reducing the particle size to increase the interfacial surface area, in accordance with the Noyes-Whitney equation²⁶². However, using nanoparticles as “smart” drug carriers for both a controlled drug release and ultrasensitive diagnostics would present the next step. Irrespective of the delivery method (local or systemic), the nanomaterial would in this case act as a depot and a carrier for therapeutic agents, providing a convenient solubilizing and protective environment. Both the shelf-life of the therapeutic prior to administration and the delivery efficacy upon administration are thus expected to be increased. The efficacy of delivery of many other drugs, including human growth hormone, leuprolide, insulin, as well as highly water insoluble drugs, such as paclitaxel (small hydrophobic drug used for the treatment of cancer), levonorgestrel (used for female birth control), and morphine was increased upon their incorporation within specific drug delivery vehicles, whereby pharmaceutical compounds having been delivered so far by nanocarriers include mostly small molecules, peptides, and DNA²⁶³.

Many different systems are nowadays investigated for their usage as drug delivery carriers, including micelles, vesicles, polymers, microspheres, hydrogels, and solid implantable devices, such as microchip-based drug delivery systems consisting of microfluidics combined with sensors²⁶⁴. However, as we shall see, inorganic nanoparticles offer many advantages in comparison with other concurrent systems. In addition, owing to their high surface-to-volume ratios, nanoparticles offer enhanced reaction sites compared to their bulk counterparts. As smaller, they also more easily penetrate the extracellular space and are more efficiently absorbed by the cells, thereby increasing the efficiency of the therapeutic action.

2.2. Why calcium phosphates?

CAP particles with tunable phase composition and thus the resorption rate offer an advantage of increasing the efficiency and local character of the drug delivery as well as a controlled release over time. A reduced dosage and frequency of administration, and consequently fewer side effects, would result thereby. By means of fine CAP particles that are stable prior to injection and then comparatively slowly resorbed in the body, the active compound is protected until the particles are injected and delivered to the targeted site, after which a controlled release, tunable to anywhere between a few days to a few weeks, may be set to take place.

CAP particles are also able to permeate the cell membrane and dissolve in the cell, which makes them an attractive candidate for gene delivery agents too^{265,266,267}. The encapsulation of DNA into CAP nanoparticles protects the nucleic acid from cytoplasmatic environment and enables its efficient delivery into the cell nucleus. Non-viral gene delivery systems have been, in fact, intensively investigated as possible alternatives to viral vectors for the transfection purposes. Although non-viral gene delivery vectors typically possess lower transfection efficiencies compared to viral ones, their excellent safety profiles are appealing for gene therapy²⁶⁸. Several nanomaterial-based vectors have been developed using functionalized multi-wall and single-wall carbon nanotubes, metallic, bimetallic, TiO₂ and silica nanoparticles, magnesium phosphate, manganous phosphate, and various dendrimers²⁶⁹. However, unlike many of these options, including poly-L-lactic acid (PLLA) and a few other polymeric particles proposed as drug/gene delivery carriers (PLLA, e.g., is typified by a very slow dissolution rate in the body, which disables a prompt release of an encapsulated active ingredient, whereas in the later stages of dissolution it undergoes a self-catalytic degradation reaction owing to an increased

release of acidic products harmful for the surrounding tissues²⁷⁰), the degradation of CAP does not produce any toxic chemicals apart from the release of Ca^{2+} (which may in the worst case scenario initiate protein aggregation), and these particles are also scalable in size and imply low manufacturing costs. In that sense, nanosized CAP particles present one of the most viable options for gene delivery systems, and for that reason they have been extensively used as an *in vitro* gene delivery agent for over 30 years and are currently being investigated for *in vivo* purposes as well^{271,272}.

Structural and chemical features of CAP particles may provide all the essential requirements for their efficient usage as genetic transfection agents, including: the binding affinity of DNA molecules onto HAP crystals²⁷³; the composite particle stability in extracellular space; an efficient cellular uptake and resorbability of the particles followed by a gradual release of the DNA and its escape from the endosomal network into the cytoplasmatic intracellular space, and eventual cytosolic transport and nuclear localization for transcription²⁷⁴.

Compared to other drug delivery carriers, CAPs possess the following advantages:

- Favorable biodegradability and biocompatibility properties in general.
- Soluble and less toxic than silica, quantum dots, carbon nanotubes or magnetic particles²⁷⁵.
- More stable/robust than liposomes, which predisposes them for a more controlled and reliable drug delivery. Contrary to liposomes and other micelle-based carriers, which are subject to dissipation below specific critical concentrations (which presents a clear obstacle upon injecting them into the bloodstream), CAP-based systems and particularly those with Ca/P molar ratio close to the one of HAP, are negligibly soluble in blood, which is by itself supersaturated with respect to HAP.
- Higher biocompatibility and pH-dependent dissolution compared to polymers²⁷⁶. Namely, the dissolution of CAP is accelerated at low pH media, which are typically found in endolysosomes and in the vicinity of tumors, providing an advantage in the delivery of drugs into malign zones or cell organelles. Also, unlike most metallic and oxide nanoparticles, including Au, Ag, Co, Cr, Cd, Se, Te, TiO₂, CuO and ZnO, which have all been shown to induce damage to DNA, produce oxidative lesions, increase mutation frequency and decrease cell viability, nanosized CAP belongs to the class of the safest nanomaterials evaluated so far (together with SiO₂ and most likely Fe₂O₃ and C₆₀)²⁷⁷.
- Low production costs and excellent storage abilities (not easily subjected to microbial degradation).
- Preparation of nanosized CAP is not a complicated task, provided we recall that with the exception of enamel, CAP in the biological domain almost exclusively exists in form of nanoparticles. This is explained by the free energies of nucleation and of crystal growth being approximately the same for a wide range of conditions, implying that the formation of elongated HAP crystals is not a strictly favorable process unless it occurs through aggregation of smaller, nanosized units²⁷⁸. A multihierarchical model of naturally highly

defective HAP crystals has thus been proposed²⁷⁹, the lowest element of which corresponds to the so-called Posner's clusters of approximately 1 nm in size. In that sense, CAP nanoparticles require less effort and are less time-consuming to prepare compared to quantum dots and dendrimers, respectively.

- Unlike other alternatives, both Ca^{2+} and PO_4^{3-} are naturally found in the body in form of amorphous and crystalline solids in hard tissues as well as in blood (in concentrations of 1 – 5 mM). Ideally, CAP particles would safely be distributed throughout the body, with the dissolved material being regulated by the action of kidneys, avoiding their precipitation as pathological deposits.
- Unlike most other ceramics, nanosized CAP can be prepared *in situ*, under ambient conditions, in a wide array of morphologies, from spheres²⁸⁰ to platelets²⁸¹ to rods²⁸² to fibers²⁸³. In one particular set of experiments, combinations of 2 different temperatures (25, 40, 60, and 80 °C), 4 pH values (8, 9, 10, and 11), and 3 drying methods (atmospheric, vacuum and freeze drying) yielded a variety of HAP particle morphologies, ranging from spheres to rods to wires and needles to “bamboo-leaf-like” ones²⁸⁴. The particle morphologies could be optimized for the most favorable dissolution and adhesive properties in the body. For example, whereas nanospheres would exhibit less friction while moving through plasma, platelets and needle-shaped particles would have a higher propensity to adhere^{285,286}, which may be desirable in situations where clearance from the mucosal tissues by the mucosal layer disfavors an effective uptake of the drug.
- CAPs can also be prepared with a variety of phase compositions (Table 1), thereby enabling fine tuning of the dissolution properties *in vivo* at the structural level as well. For example, the higher the content of TCP on the account of the content of HAP in biphasic CAP, the higher the dissolution and resorption rates of the material^{287,288}. The optimal drug release rates could thus be tuned by changing the HAP/TCP molar ratio within the particles aside from modifying their size and/or morphology. Calcination time and temperature can also be used to control the particle size and composition, and have consequently been shown to affect the drug delivery performance of HAP nanoparticles²⁸⁹. In general, in case of biphasic CAP, tiny differences in chemical composition may lead to significant differences in material properties²⁹⁰. Control over a single synthesis parameter can thus be used to prepare CAP materials with a wide range of intrinsic properties²⁹¹.

The main potential downside of injecting CAP nanoparticles in the body, which needs yet to be assessed prior to reaching clinical application stages, certainly comes from the possible atherosclerotic complications associated with their undesirable deposition along the arterial walls. Namely, it is known that cholesterol and its derivatives, altogether with various CAP phases, primarily HAP, comprise the largest percentage of atherosclerotic plaque²⁹² (Fig.11). Structural matching between crystal lattices of cholesterol and HAP and their consequent epitaxial co-precipitation are largely assumed to be responsible for their entwined presence in these pathological cardiovascular deposits²⁹³. Firstly, crystals of both HAP and cholesterol monohydrate typically develop (001) faces. Furthermore, OH^- ions of HAP lie in planes parallel to (001) face and are thus facily engaged in hydrogen bonding across the exposed (001) crystal

planes, especially since a fit of the (001) planes of the two crystal structures shows a close superposition of the hydrogen bonding groups²⁹⁴. A crystalline layer of either of the two structures can thus serve as a quasi-epitaxial nucleation surface for the growth of the other.

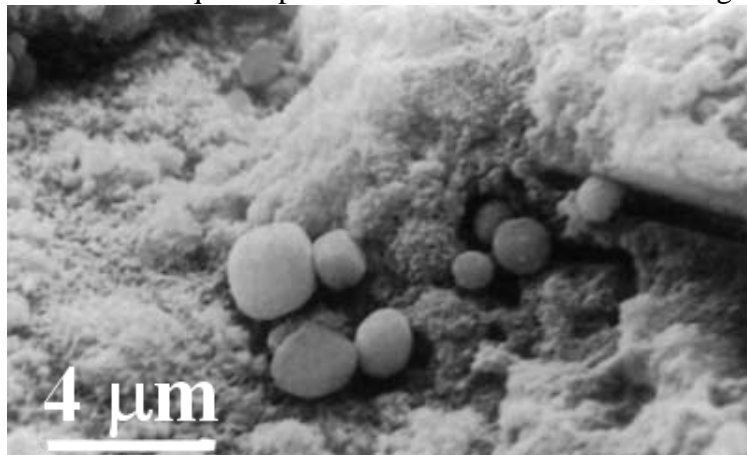


Fig.11. Spherical CAP particles found in segments of atherosclerotic deposits. Reproduced with permission from Ref.295.

To be truly innovative and advanced, drug delivery carriers should be made multifunctional, that is, allow for more than one function upon their injection in the body. Theranostic systems that possess the capacity for a simultaneous controlled drug delivery and imaging/diagnostics thus hold great prospect in this sense. Smart delivery rates and amounts depending on the diagnostic response are meant to be integrated within these particulate systems. As recently observed by Sumer *et al.*, “Successful development of theranostic nanomedicine requires significant advances in materials science and nanocomposite materials; ideal nanomedicine platforms should be small in size, provide high drug-loading densities, be efficient in targeting to the diseased tissues with minimal nonspecific uptake, provide responsive release mechanisms to improve drug bioavailability and also imaging ultrasensitivity to pre-validate and monitor therapy”²⁹⁶. By combining several useful properties within a single particle, multifunctional pharmaceutical nanocarriers may significantly enhance the efficacy of many therapeutic and diagnostic protocols, and it is them that could be considered as the most exciting synthetic ideal in the field of controlled drug delivery, as of today.

2.3. Prospective applications

The first and still the foremost application of nanosized HAP is as components of scaffolds for the reparation of impaired hard tissues^{297,298}. In surgical procedures, allografts and autografts as parts of hard tissues taken from donors or the patient itself make up for 95 % of materials used as bone grafts, whereas only 5 % are synthetic materials²⁹⁹. Whereby in case of autografts, two separate surgical procedures are required followed by an increased risk of infection and failure, a risk of implant rejection and disease transmission, including the need for an extensive sterilization procedure during which the properties of the substitute material are often degraded, are present in the case of allografts. Hence, there is a great impetus on the research of novel and reliable synthetic materials for substitution of hard tissues. Although metals including Fe, Co, and Ti are often used to fill bone defects and provide the internal fixation, fatigue, corrosion, tissue infection and poor implant-tissue interface result on many

occasions^{300,301}. On the other hand, there are problems that prevent the wide usage of ceramic-based materials for reparation of hard tissues. For example, HAP is all by itself an exceptionally brittle material, similarly to most ceramics. Hence, it does not provide a sufficient mechanical support for the surrounding bone/skeleton. HAP is the most osteogenic CAP phase, which can be attributed to its being the major mineral phase of bone, dentin and enamel; however, its rate of resorption is quite low. The rate of resorption smaller than the rate at which new bone forms is a discrepancy that impedes proper healing of the bone defect. It has to be set to precisely match the rate of expected bone regeneration in order to maximize the efficacy of the healing process³⁰². Henceforth, much research has been done on exploring the mixtures of HAP and TCP, known as biphasic calcium phosphates (BCP), for the most optimal restoration of hard tissues^{303,304}. Namely, TCP and, as of recently CaSO_4 ³⁰⁵, present more rapidly resorbable osteogenic phases than HAP, and are used to balance the low resorption rate of the latter. A medical interest in amorphous CAP has likewise been sparked after the discovery of its higher bioresorbability compared to HAP and other CAP phases³⁰⁶. However, the main drawbacks associated with the application of amorphous CAP in the body include its low mechanical strength and difficult injectability. On the other hand, CAP phases with Ca/P ratio of less than 1 are not suitable for biological implantation due to their overly high solubility. The design of a material that is both mechanically strong enough for bone replacement and biodegradable is an especially difficult challenge for scaffold-based strategies. One of the strategies used to design a biomaterial that is both biodegradable and mechanically strong enough to serve as a bone substitute is to couple particle dimensions in the nano range with a soft component that contributes tensile properties to the stiff ceramic component. Polymer matrices have thus been reinforced with HAP nanocrystals to improve the mechanical properties of these polymers^{307,308}. Poly-L-lactide³⁰⁹, poly(lactic-co-glycolic acid) (PLGA)^{310,311,312} and collagen³¹³ all present prospective choices as polymeric phases that promote cellular adhesion, proliferation and growth. In fact, inorganic nanoparticle fillers in general have been shown to add tensile strength, stiffness, abrasion resistance, crack resistance, and stability to polymer networks. Depending on the time scale of the desired biodegradation of the polymer phase, one may simply vary its chemical nature; PLLA thus requires approximately a year to be fully resorbed, polycaprolactone has a biodegradation period of about 6 months, whereas PLGA degrades in the body in a month or so. Another step in developing advanced regenerative biomaterials comprises introducing growth factors (BMP-2 and PDGF), cells, antibiotics and additional ions into injectable composites based on CAP and various polymers or proteins³¹⁴.

However, the main application that this review will focus on will be the usage of HAP/CAP nanoparticles as theranostic drug delivery and gene therapy carriers. In view of that, the primary interest may be given to CAP nanoparticles with encapsulated antibiotics. Examples of such compounds may be gatifloxacin³¹⁵ (a fluoroquinolone that has bactericidal property for both anaerobic and aerobic bacteria) and ciproflaxacin³¹⁶, both of which are used for treating osteomyelitis. In this case, the inflammation of bone tissue destroys vascular channels, leaving a portion of dead and infected bone (sequestrum) detached from the adjoining healthy bone. Owing to the impaired vascularity, antibiotics may not be delivered adequately to the lesion by the intravenous route. It is usually recommended that the administration of antibiotics lasts 4 – 6 weeks in the treatment of chronic osteomyelitis; however, through a drug delivery with sustained and controlled release rate the necessity of repetitive, daily injections of adequate doses of the drug would be eliminated. The thing of main concern here is the ability to design a material that would enable a high enough initial burst of the drug release, which would surpass the minimal

inhibitory concentration for the given infective agent, such as a bacterial colony. If, on the other hand, the amount of the released drug is continuously lower than this critical level, it may result in developing antibiotic resistant organisms, rather than keeping their population under control, through such a controlled release of the antibiotic.

By means of antibiotic-impregnated CAP nanoparticles, a local delivery of antibiotics would also be coupled to remineralization of the lost portion of bone. Such an approach would overcome difficulties that implantable antibiotic delivery systems currently face; namely, the need for the removal of the implant at the end of the treatment. CAP/HAP particles carrying gentamicin and vancomycin were thus shown not only to exhibit an antibiotic effect on the healed tissue, but to stimulate the growth of the new bone owing to the bioactivity of the carrier and its similarity to the natural composition of bone³¹⁷. CAP particles are in this case also expected to be more effective than polymers (e.g., polycaprolactone)³¹⁸ in view of the latter not being able to provide support for bone tissue reconstruction at the targeted space. Another antibiotic effective in treating osteomyelitis, recently injected within PLLA nanospheres coated with CAP³¹⁹, is tigecycline, which possesses activity against both gram-positive and gram-negative pathogens, binding to the 30S ribosomal subunit and inhibiting protein synthesis³²⁰. Cefuroxime was also impregnated within porous HAP/TCP and successfully used to treat osteomyelitis, with the implants inducing the highest concentration of the antibiotic in bone and serum 21 days after the implantation³²¹. Amoxicillin, one of the most common antibiotics prescribed to children has also recently been injected into PLGA microspheres coated with HAP³²², whereby the same research group also managed to encapsulate indomethacin, a hydrophobic drug, within HAP-coated liposomes based on 1,2-dimyristoyl-sn-glycero-3-phosphate (DMPA) and 1,2-dimyristoyl-sn-glycero-3-phosphocholine (DMPC)³²³. Doxycycline, an antibiotic used to treat periodontitis, has also been encapsulated within HAP nanoparticles, exhibiting sustained and single or multiple stage release profiles depending on the Ca/P ratio³²⁴. Other compounds, such as vitamins^{325,326,327,328} or Coenzyme Q10 (i.e., ubiquinone, a lipophilic antioxidant)³²⁹, have been prepared using this method (Fig.12). Using CTAB as an additive, uniform hollow CAP nanospheres have recently been prepared with a potentially improved drug uptake efficiency³³⁰. Nanosized hollow ellipsoidal HAP/CaSiO₃ composites with the improved drug loading capacity were also prepared in a multistage process involving a selective dissolution of the initially precipitated CaCO₃ core³³¹. CAP particles could be also coupled with stem cells^{332,333,334,335} or bone marrow mesenchymal stromal cells³³⁶, both of which are known to facilitate the regenerative therapy of connective and hard tissues. Various elements of extracellular matrix (ECM) could also be incorporated with the purpose of improving the biocompatibility and bioactivity of the particles or accelerating the osteoinductive response of the surrounding hard tissues³³⁷. Individual amino acids and their combinations have also been anchored on the surface of HAP nanorods³³⁸. Hydrophilic organic coatings, such as albumin, dextran or hydroxymethylmethacrylate could also be used to enhance biocompatibility of the particles³³⁹. With respect to regeneration of hard tissues, all of such systems could be applied for treating their loss occurring due to infection or even treat soft tissues such as pulp³⁴⁰ or cementum-gingiva-alveolar bone interface damaged by a periodontal disease in dentistry.

As CAP and bioactive glasses are still considered as compounds that the next generation of biomaterials will be based on^{341,342}, synthesis of nanosized CAP particles coated with Bioglass or blended within nanophase composites³⁴³ would present another prospective research direction. Such a material could be used both as a filler of bone defects and an efficient drug delivery carrier owing to facile surface functionalization through the silanol groups. If the layers of

bioactive glass are thin enough (a few atomic layers), it would predispose the particles for a relatively quick dissolution of the coating (bioactive glass has a significantly higher rate of resorption compared to CAP), right after the attached drug has fulfilled its role, after which the CAP core can act as a bone filling or biodegradable remineralization agent. In such a way, both the ideals of an efficient drug delivery and bioremineralization would be satisfied. Furthermore, by controlling the thickness of the coating (achievable through varying the control parameters of the synthesis), the time control over the dissolution of the bioactive glass and the consequent release of the drug could be possessed. Coating with silica glass is typically carried out via a hydrolysis of given alkoxides (i.e., tetraethyl orthosilicate (TEOS) in most cases) and localized polycondensation on the surface of the core particles^{344,345,346}. LaSr-manganite particles were thus coated with silica³⁴⁷, whereas cholesterol platelets were coated using sodium silicate solution and a procedure involving a reaction with an ion-exchange resin³⁴⁸. A challenge would be to coat CAP particles with Au (metal-ceramic coatings are difficult to achieve owing to different thermal expansion coefficients between the two components among other factors), but if succeeded, such particles shelled with Au could be used for photothermal therapies and diagnostics, such as optical coherence tomography³⁴⁹. The absorption efficiency of Au is a few orders of magnitude higher than that of conventional photothermal dyes, and Au surfaces can be facilely functionalized with antibodies and other bioactive molecules. On the other hand, instead of being the core component of drug delivery nanoparticles, CAP phase can be coated around another phase. Such an approach was used to prepare CAP-coated PLLA microspheres with a sustained 30-day release profile of attached bovine serum albumin (BSA) in SBF compared to non-coated particles typified with an initial burst in the drug release³⁵⁰. An opposite approach, that is, co-precipitation of PLGA and a drug, tige cycline, in the presence of precursor HAP nanoparticles was recently implemented, resulting in HAP/PLGA/drug composite particles³⁵¹. They turn out to be particularly appealing not only because of their biocompatible and biodegradable nature, but because HAP releases hydroxyl groups upon dissolution, whereas PLGA releases acidic products of degradation. In combination, the two components thus balance each other's potentially harmful pH changes. Notice, however, that the higher the amorphous content in CAP/HAP particles, the less intensive the corresponding increase in pH is. A convenient circumstance is that such phases with a lesser number of hydroxyl groups in the stoichiometric formula dissolve faster than HAP. As a result, the slow dissolution of HAP in biological conditions does not necessarily produce pH changes that could not be buffered by the organism. To achieve this simultaneous dissolution of the polymer and mineral components and avoid the two-step drug release profile, dispersion of nanosized CAP particles within polymeric particles as matrices may have to be ensured instead of the classical core-shell model where the drug-containing polymer would thoroughly coat the CAP core. Another favorable feature of both HAP and PLLA/PLGA³⁵² is their negatively charged surface under the physiological conditions, which naturally fosters the attachment of cells thereto.

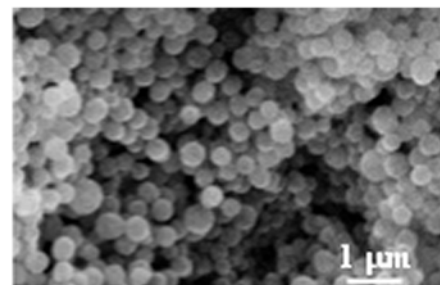
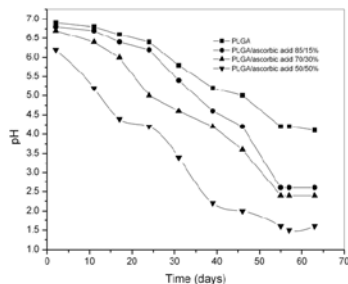
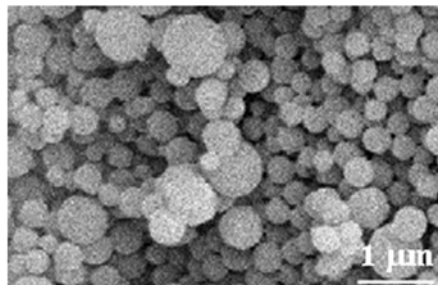


Fig.12. Uniform polylactide-co-glycolide spheres injected with ascorbic (left) and folic acid (right). The composition and size of the particles could be modified so as to optimize the release rate of the encapsulated model compound (middle). The same approach could be applied for the fabrication of drug-carrying CAP/HAP nanoparticles with controlled drug release rates. Reprinted with permission from Ref.353.

For the theranostic purposes, particles need to satisfy the requirements for simultaneous imaging/diagnostics and therapeutic effect. For the imaging purposes, the particles could be doped with specific ions or compounds, which would enable their detection using an appropriate external device. Some of the methods for imaging include fluorescence imaging (in which case fluorescent probes need to be incorporated within the particles), MRI (in which case, doping the particles with magnetic ions is carried out), gammascintigraphy (in which case, doping the particles with heavy atoms such as ^{111}In , $^{99\text{m}}\text{Tc}$, Mn, and Gd is required³⁵⁴), X-ray computed tomography and ultrasonography. At a finer scale, one could perform molecular imaging, which is defined as visually representing, characterizing and quantifying (sub)cellular biological processes within the organism. These processes include gene expression, protein-protein interaction, signal transduction, cellular metabolism, and both intracellular and intercellular trafficking. For optical imaging, the particles are normally doped with organic fluorophores, such as Cascade Blue (dark blue), 10-(3-sulfopropyl) acridinium betaine (SAB; light blue), fluorescein (green), rhodamine WT (orange), and Cy3 amidite (magenta), which were all recently encapsulated within CAP nanoparticles³⁵⁵. However, CAP particles could be equally doped with Eu^{3+} , Tb^{3+} , Gd^{3+} or other lanthanides in order to be made fluorescent^{356,357}. Nanocrystals actually offer the advantage of structurally integrating the fluorescent entities within the crystal lattice by doping, thereby transcending numerous drawbacks of using organic dyes, fluorescent proteins, and lanthanide chelates, such as broad spectrum profiles, low photobleaching thresholds, and poor photochemical stability³⁵⁸. The pathways of such nanoparticles were successively followed inside pancreatic cells³⁵⁹. Accumulation of DNA-loaded CAP nanoparticles which also contained red-fluorescing tetramethylrhodamine isothiocyanate (TRITC) BSA inside a cell and its nucleus was observed by fluorescence microscopy³⁶⁰. Combining CAP with semiconductor quantum dots^{361,362} at the particle level could also be used to enable monitoring of the route that the particles follow in the body.

It has been evidenced that the addition of ions that substitute Ca^{2+} in HAP crystal lattice can induce a more favorable capturing of the carried biocomponent, such as DNA³⁶³. Evaluating the effect of additives, such as Mg^{2+} , CO_3^{2-} , K^+ and Al^{3+} , most of which are known to be present in biological apatite, may thus be useful. The effects of many external ions on precipitation of HAP have previously been established, and it is known, for example, that citrate, pyrophosphate and molybdate ions reduce the crystallinity of HAP, whereas Mg^{2+} , Al^{3+} and Sn^{2+} at high concentrations inhibit the hydrolysis of the initially precipitated amorphous CAP phase³⁶⁴. At low concentrations, however, Mg^{2+} , Al^{3+} and Zn^{2+} promote the formation of substituted β -TCP. MgO and SrO have been shown to suppress β - to α -TCP transition and also promote attachment, proliferation and differentiation of osteoblast cells³⁶⁵. Strontium is present in tiny concentrations in bone and is, like Si ions, supposed to play a role in early bone formation, whereby 0.01 % of the human body weight is amorphous silica. Similar to other ions, such as carbonate, sodium and magnesium, these two also occur as lattice substitutions at the atomic level. Quite rarely, as in the case of strontium-based toothpastes, strontium precipitates in form of $\text{Sr}_3(\text{PO}_4)_2$ with SrCl_2 being the intermediate. In any case, the mechanism by which Si increases *in vivo* bioactivity is still unresolved. Mechanisms proposed to explain this effect can be divided to: a) passive, including the Si-induced effect of decreased apatite grain size, a change in the protein

conformation on the biomaterial surface or modifications of the surface topography and therefore the cellular response; and b) active, referring to the release of Si ions, its cellular uptake and promotion of gene expression in the direction of an increased bioactivity³⁶⁶. In general, understanding the precise biological mechanisms with respect to implantation of HAP in the body is still required, which is why an enormous shift of interest towards looking at genetic expression pathways following the implementation of HAP and bioactive glasses is witnessed today^{367,368}. There are ongoing efforts to follow the genetic expression pathways in osteoblast and fibroblast cell cultures and thus design the most optimal ionic composition of the implantable material, which would presumably lead to the so-called “third generation” of biomaterials³⁶⁹. In that sense, exploration of additives that may not even be present in the body and evaluating their bioactivity responses, may present a reasonable choice.

Another interesting direction is doping CAP with magnetic ions, such as Co^{370,371} or Fe^{372,373}, or even co-precipitation of magnetite³⁷⁴. There are indications that magnetic HAP and electrically polarized one (owing to OH groups in HAP crystal lattice) possess a more pronounced osteogenic response^{375,376}. The structure and composition of HAP have been shown to be stable upon substitution with exogenous metallic ions in the lattice^{377,378}. Such particles could be guided to the targeted area in the organism by means of an external magnetic field. They could be used either as contrast agents for MRI, for cell separation or as a heat mediator in hyperthermia therapies³⁷⁹. Magnetic HAP synthesized by co-precipitation in the presence of FeCl₂, thereby partially substituting Ca²⁺ with Fe²⁺ in the resulting crystal lattice, was recently successfully applied in treating murine colon cancer by hyperthermia, following a subcutaneous injection of the particles³⁸⁰. Owing to their small sizes, the particles would be superparamagnetic and would thus resist agglomeration caused by magnetic dipole-dipole interactions outside of the external field³⁸¹. Also, heavy metal atoms, such as ¹¹¹In, ^{99m}Tc, Gd or Mn, loaded into the nanocarrier via a carrier-incorporated chelating moiety can lead to particles for gamma- or MR imaging applications. Hetero-epitaxial growth of magnetite on HAP resulted in 300 nm sized magnetic composite particles with saturation magnetization of 30 emu/g³⁸². The particles showed an increased transfection of stem cells with the gene for glial cell line-derived neurotrophic factor in the magnetic field compared to the transfection efficiency reached in the absence of the external field. Superparamagnetic CAP composite nanospheres could also be applied for the blood detoxification. Functionalized with target-specific antitoxin receptors and let freely circulate within the bloodstream after simple intravenous injection, the particles would sequester the blood-borne toxins using their surface receptors. The blood is then circulated via a catheter through a portable high gradient magnetic separator device where the magnetic toxin complexes are retained and the detoxified blood is returned back to the bloodstream³⁸³. Unlike sole drugs, 98 % of which cannot transverse the blood-brain barrier (BBB) nor penetrate the skin, some nanoparticles can³⁸⁴, which makes them attractive for acting as carriers for drugs that are to be delivered across the BBB and the epithelial junctions of the skin that normally impede the delivery of drugs to the desired target³⁸⁵.

In fact, because the type and efficiency of the transport of nanoparticles across the cellular membrane largely depends on their size, developing synthesis methods for the size-controlled fabrication of HAP nanoparticles is essential for fine-tuning their application as drug and gene delivery agents. The transport mechanism may also predispose the particles to exert a specific effect on the cell. Particles smaller than 20 nm typically either pass the cell membrane through the membrane channels or by following vesicular or caveolae-based endocytosis routes³⁸⁶. If the particle size fits the size of the active part of receptor molecules docked on the

cell membrane, a receptor-induced transport with a possible synergetic effect on the cell may be triggered. A few studies have shown that nanoparticles of ~ 50 nm in size cross the cell membrane faster than those smaller than 15 nm or bigger than 200 nm on average, and that smaller is not necessarily the better when it comes to choosing the most optimal size for a given drug/gene delivery application^{387,388}. In fact, different application purposes require different optimal particle sizes³⁸⁹. Whenever an external control of the particles in the body is to be established, a compromise needs to be made regarding their size, since particles that are too small would tend to difficultly oppose the direction of the bloodstream and would respond to the external field with a less intensive signal, whereas particles that are too big would be less movable owing to their tendency to get interlocked between macromolecular complexes, cells and tissues³⁹⁰.

Surface modification by means of adding cell-binding peptides onto the surface or in the bulk of biomaterials has been used to attain control over cell-biomaterial interactions and promote a more efficient healing process upon the injection of the biomaterial in the body³⁹¹. By the action of surface-functionalized nanoparticulate carriers, the drug can be locally focused within the diseased area, so that a controlled and extended on-site release, with lesser doses required, can be achieved, with a greater control over toxicity and bioavailability of the drug. As for the intracellular delivery, surface functionalization can be used to optimize the charge of the particles, keeping in mind that positively charged particles are endocytosed at a higher rate compared to negatively charged ones (as cell membrane is negatively charged due to the surface-exposed phospholipids)³⁹². Good efficiency of CAP nanoparticles carrying a photoactive dye against rabbit synoviocytes and murine macrophages was thus attributed to their positive charge achieved by means of functionalizing the particle surface with poly(ethyleneimine) (PEI)³⁹³. The most common peptide for biomimetic surface modification is Arg-Gly-Asp (RGD), a ubiquitous peptide present on ECM proteins and shown to promote cell adhesion in multiple cell types³⁹⁴. Specific targeting ligands, usually monoclonal antibodies, may be attached to the carrier surface, thus promoting a diagnostic molecular recognition and the immunocarrier character of the particles. PEG as a protecting polymer or BSA can be attached to the surface of the particles so as to increase the circulation time of the nanocarrier in the blood^{395,396}. Such a strategy was employed by Abraxis Bioscience, Inc., to increase the uptake efficiency of the anticancer drug, paclitaxel, and prevent the side effects associated with using toxic solvents. Cell-penetrating peptides attached to the carrier surface would also allow for an enhanced uptake by the cells. CAP nanoparticles could also be functionalized³⁹⁷ or encapsulated³⁹⁸ with chitosan, a naturally derived polysaccharide, which is known to have apoptotic effects on tumor cells *per se*^{399,400,401}. A method for injecting doxorubicin, a cancer therapeutic chemical into degradable CAP non-agglomerating particles with 20 – 40 nm in size, with the result of having the same efficacy but significantly less expressed side effects, has recently been reported⁴⁰². Methotrexate, another anticancer drug, was also successfully loaded into 250 nm sized CAP particles prepared by reverse micelle technique⁴⁰³. CAP nanoparticles conjugated with cis-diamminedichloro-platinum (CDDP, cisplatin) were also prepared, exhibiting twice higher IC₅₀ values when compared to the free drug in an *in vitro* cell proliferation assay using a human ovarian cancer cell line⁴⁰⁴. Folic acid with its high affinity for folate receptors that are often overexpressed in cancer cells presents another compound that may be added to CAP nanoparticle carriers⁴⁰⁵. Also, photosensitizers, such as 2-devinyl-2-(1-hexyloxyethyl)pyropheophorbide (HPPH), could be incorporated in or coated on the surface of CAP particles, which would thus render the particles capable of being applied in cancer photodynamic therapy⁴⁰⁶. Raman labels, such as basic fuchsin, or nanoscopic

domains of silver or gold to initiate surface-enhanced Raman scattering (SERS) could also be attached to the core particles, which would make their antibody-conjugated versions detectable upon binding to cells or specific biomolecules by means of Raman spectroscopy⁴⁰⁷. Finally, all of these functional surface additives could be simultaneously applied in the role of emulsifiers, stabilizing the nanoparticulate suspensions and preventing agglomeration of the particles⁴⁰⁸.

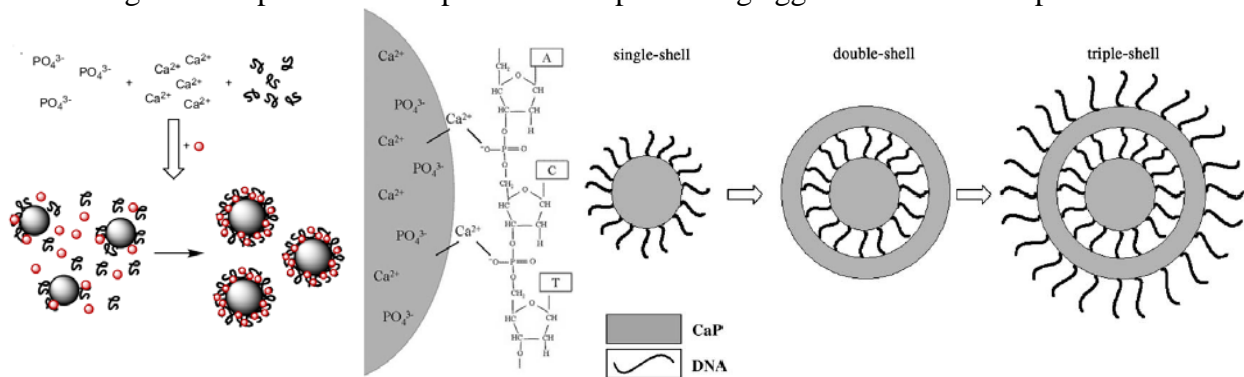


Fig.13. Precipitation of CAP in the presence of steric agents and other active molecules or particles (red dots), such as fluorescent probes, superparamagnetic entities or drugs, may lead to stable dispersions of medically functional CAP nanoparticles. The model for the attachment of nucleic acids onto the surface of these particles is shown in the middle, whereas different types of multilayered DNA-CAP composite particles prepared by the co-precipitation technique are shown on the right. Reprinted and adapted with permission from Ref. 409.

Owing to the affinity of Ca²⁺ ions on the surface of CAP particles to the helical PO₄³⁻ groups of DNA⁴¹⁰, HAP has been applied as an adsorbent in chromatography for separation of DNA⁴¹⁰. As Ca²⁺ ions can interact with carboxylate residues on the protein surface, whereas PO₄³⁻ functional groups can interact with basic protein residues, HAP has also been used for the separation of proteins^{411,412}. Functionalizing the surface of HAP particles with polymers, peptides, proteins, cell-penetrating moieties, DNA or oligonucleotides can thus be readily carried out (Fig.13). Simple co-precipitation procedures were reported for the synthesis of DNA/CAP composites, with the biocomponent either encapsulated or anchored on the surface⁴¹³. DNA could be attached to the particle surface, thus enabling their use as cell transfection agents, i.e., non-viral vectors for the gene therapy⁴¹⁴. The usage of CAP particles for this application dates back to the works of Graham *et al.* in 1973⁴¹⁵, and is nowadays routinely used for *in vitro* transfection procedures. As far as the state-of-the-art is concerned, although non-viral delivery systems show a lesser efficiency compared to the viral ones, they are markedly safer. Producing particles that would possess transfection efficiencies comparable to those of viral carriers thus remains a challenge, particularly in view of the fact that FDA has not approved any gene therapy carriers yet⁴¹⁶. However, DNA can also be injected within the particles, thus preventing its detachment in the blood stream and ensuring its secure intracellular delivery⁴¹⁷. In addition, by sequential precipitation of CAP and adsorption of DNA, core-shell particles with multiple layers of therapeutic organic components separated by walls of CAP and thus released at different time points and rates could be prepared⁴¹⁸. The same technique of layer-by-layer precipitation could be applied for producing other types of multilayered polymeric/peptide/CAP particles⁴¹⁹. Functionally graded HAP coatings encapsulating a drug can also be synthesized to enable tunable time-dependent drug release profiles⁴²⁰. As for the gene transfection a delivery of DNA into the cell nucleus has to be ensured, antisense strategies using siRNA (gene silencing) require a simpler approach, as in this case it is sufficient to deliver siRNA into the cytoplasm⁴²¹. To ensure a more effective size control and serum tolerability and, therefore, a more effective

genetic knockdown with CAP nanoparticles as siRNA-carriers, they were used as components of hybrid micelles together with PEGylated polyanion block copolymers, such as PEG-b-poly(aspartic acid) and PEG-b-poly(methacrylic acid)⁴²². Also, attaching oligonucleotides to nanoparticles has been shown to increase the resistance of the DNA fragments with respect to enzymatic degradation⁴²³. Coating CAP particles with the protective layer of oligonucleotides in addition to conjugating other functional compounds thereto (adsorbed, covalently bound or entrapped in the crystal structure) is thus an interesting research direction to explore.

Osteoinductive and cytotoxic behavior of CAP nanoparticles has been evaluated both on cell cultures and *in vivo*^{424,425,426,427}, with the purpose of providing insights into their biocompatibility (biosafety as much as biofunctionality) characteristics. However, these tests have so far been proven as sufficient only to permit the usage of CAP nanoparticles as filler components in scaffolds implementable in reparation of hard tissues. For the purpose of applying these particles as drug delivery vehicles, significantly more extensive and qualitatively more complex evaluations need to be performed, particularly in view of the fact that toxicity of a compound can be different depending on size and/or morphology of the particulate form in which it is applied, and the fact that circulating nanoparticles can get into contact with practically any tissue in the organism, producing unforeseeable effects thereon. One report thus shows how polytetrafluoroethylene (PTFE) particles with ~20 nm in size are harmful when inhaled, whereas 130 nm-sized particles with the same composition do not produce any toxic effects⁴²⁸. On the other hand, there are nanoparticulate compositions that are prone to agglomeration upon their intravenous administration, and are only in such form evidenced as damaging for the body⁴²⁹. Sometimes, also, it is only a specific window of particle sizes that produces a given, desired or undesired biological response, as it has, for example, recently been shown how 2 and 12 nm sized particles did not induce a specific biological response that 7 nm sized particles did⁴³⁰. As already noticed, a possible side effect of introducing extensive amounts of CAP into the body may be induced local atherosclerosis; hence, possible methods for assessing this effect include analyzing the cross-sections of arteries or looking at the deviations from the normal body weight gain in mice following administration of the drug delivery agent. Serum analyses can be used to follow bone metabolism (via levels of Ca^{2+} , $\text{H}_2\text{PO}_4^-/\text{HPO}_4^{2-}$ and alkaline phosphatase), kidney function (via BUN-to-creatinine ratio), and nutrition (via albumin and total levels of protein). Although intracellular injection of CAP particles may pose concerns in terms of the possibly harmful increase in levels of Ca^{2+} in the cytoplasm, such concerns are significantly minimized during the extracellular drug administration. Also, following the injection of typically a few hundreds of milligrams per kg of the body weight, histological and weight analyses of the dissected vital organs as well as hematological analyses (haematocyte and hemoglobin content and thrombocyte count) are often performed to evaluate the possible pathological effects of the drug delivery procedure.

Finally, the ideals of multifunctionality⁴³¹ dictate using multiple structural and surface modifications of CAP nanoparticles mentioned hereby in parallel, and exploring their synergy. One example of such a multifunctional composite particle is shown in Fig.14. Composite nanoparticles composed of magnetite nanodomains interspersed within a polystyrene core, and coated with amine-functionalized quantum dots have recently been synthesized as a material for simultaneous fluorescent imaging and hyperthermia therapy⁴³². Another type of particles based on BSA-stabilized manganese-doped magnetite nanoparticles comprising green fluorescence protein (GFP) and Arg-Gly-Asp peptide (known to bind specifically to $\alpha_v\beta_3$ integrin, which is expressed in tumor cells) bound to Cy5-dye-labeled thiolated siRNA (inhibiting the expression

of GFP), for the purpose of simultaneous molecular imaging and gene silencing, was also recently produced⁴³³. Theranostic HAP nanocrystals doped with Eu^{3+} and Gd^{3+} were engineered to enable simultaneous contrast enhancement for three different imaging techniques (XRD, MRI and near-infrared fluorescence imaging) as well as to act as therapeutic agents by being conjugated with the cell-membrane receptor ligand folic acid and by having their surface aminized with a dendrigraft polymer, PEI⁴³⁴. Also, particles comprising 100-nm-sized silica core and Au shell with finely dispersed 10-nm-sized particles of Fe_3O_4 , functionalized with anti-HER2 antibody with the purpose of specifically targeting epidermal growth factor receptors, were prepared and successfully applied, displaying an optical resonance peak, magnetic response, an intensive surface plasmon-enhanced scattering in targeted cells, and the ability to selectively destroy the penetrated cells depending on the frequency of the light source⁴³⁵. A similar approach may be performed relying on CAP as the core material. The first step in producing multifunctional composite CAP-based nanoparticles would be to establish convenient methods for fabrication of CAP nanoparticles with tunable structure and properties. Subsequently, the nanoparticles could be doped and surface functionalized with a wide array of atomic and molecular entities. To succeed in this, it is certain that inventing efficient and elegant methods of synthesis and conceiving innovative ideas on how to probe the fundamental understanding of the formation of complex composite and surface-functionalized nanoparticles relying on soft, bottom-up chemistry approaches, is essential, just as the availability of ultramodern imaging and other analytical techniques is. In other words, it is only a coordinated effort of soft, self-assembly, bottom-up synthetic techniques and top-down methods of manipulating the material structures at the fine scale that could yield desired results along this line of research.

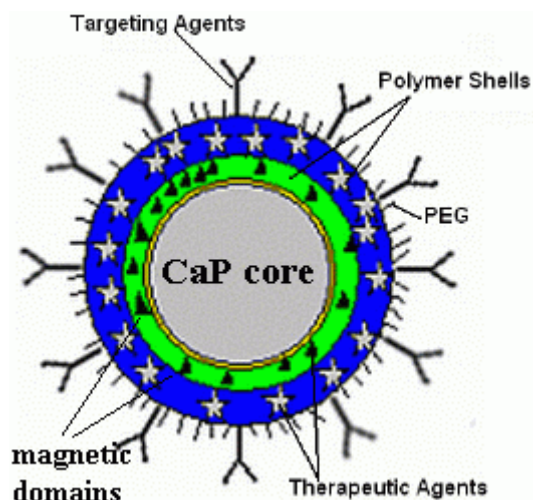


Fig.14. An example of a multifunctional drug delivery particle composed of CAP core surrounded by a polymeric shell in which magnetic domains are dispersed for the purpose of enabling the guidance of the particle in an external magnetic field and potentially producing localized heat under alternating magnetic field in hyperthermia or thermoblastic therapies. The external polymeric shell contains dispersed therapeutic agents, whereby its surface is covered by a targeting agent and PEG molecules so as to increase the circulation time of the carrier in the blood.

Conclusion

After everything said, we can conclude that nanomedicine holds immense promises for the medical treatments of the future, whereby a research focused on multifunctional theranostic CAP nanoparticles capable of diagnosing the disease and being activated as a therapeutic at a

chosen site, despite the fact that there is still a long way to go before the understanding of the chemical mechanisms of their formation is made complete, presents a prospective path for the development of this field that is still, as of today, in its embryonic stage. In order to be successful in these endeavors, the gap between life scientists and materials scientists sustained by different methods, terminologies and subjects of scrutiny existing on each one of the separate research “coasts”, has to be bridged. As of today, however, the fruitful reconciliation of these two scientific areas can be seen as nothing but enormously challenging. The other side is often hardly intelligible to most scientists, which fosters them to stick to their own fields and narrow their views into ever more specialized cognitive panoramas. Organic and biochemical entities often disobey facile analytical reproducibility and require different, oftentimes more pertinacious strategies, quite unlike more robust and macro-scale technologies implementable within materials science study concepts. Yet, the development of nanomedical methods and tools will crucially depend on the ability of natural scientists to stretch their curiosity to the other side and be open to communicate and form cross-disciplinary collaborations between the two. As in any other materials science “design” approach, a detailed understanding of the correlation between the material structure, properties and function is required for the development of CAP nanoparticles as viable drug and/or gene delivery vehicles. Hence, the basic material properties, such as particle size, morphology, specific surface area, polydispersity and composition, including control over them have to be correlated with their corresponding performance in the biomedical settings. With its strong physicochemical roots, materials science can fundamentally supplement the biomedical approach which is typically limited to investigating specific molecular recognition properties of biomolecules in elaborate statistical settings. With its engineering aspect, materials science can also provide a more reliable technological platform for biochemical studies. Life sciences can, on the other hand, provide the basis for the assessment of the real-life application potential of biomaterials designed in the lab. They can also enrich materials science with knowledge on soft matter and biochemical interactions that are critical for our ability to manipulate therewith in the design of complex multifunctional materials, such as the one that has been the topic of this review. This is all to say that to succeed in bringing CAP/HAP as advanced nanocarriers to the frontier of medicine will require a range of well coordinated interdisciplinary efforts. Bridging the gap between Materials Science and Life Science as two main contemporary research streams can be seen as a necessary precondition for satisfying this aim.



References:

- ¹ J. S. Temenoff, A. G Mikos – “Biomaterials: The Intersection of Biology and Materials Science”, Prentice Hall, Upper Saddle River, NJ (2008).
- ² J. Park, R. S. Lakes – “Biomaterials: An Introduction”, Springer, Berlin (2007).
- ³ L. L. Hench – “Science, Faith and Ethics”, Imperial College Press, London (2001).
- ⁴ I. Balać, P. S. Uskoković, R. Aleksić, D. Uskoković - “Predictive Modeling of the Mechanical Properties of Particulate Hydroxyapatite Reinforced Polymer Composites”, *Journal of Biomedical Materials Research B: Applied Biomaterials* 63, 793 – 799 (2002).
- ⁵ P. Fratzl, R. Weinkamer – “Nature’s Hierarchical Materials”, *Progress in Materials Science* 52, 1263 – 1334 (2007).
- ⁶ B. D. Ratner, A. S. Hoffman, F. J. Schoen, J. E. Lemons – “Biomaterials Science: An Introduction to Materials in Medicine”, 2nd Edition, Elsevier Academic Press, San Diego, CA (2004)
- ⁷ K. Onuma – “Recent Research on Pseudobiological Hydroxyapatite Crystal Growth and Phase Transition Mechanisms”, *Progress in Crystal Growth and Characterization of Materials* 52, 223 – 245 (2006).
- ⁸ J. D. Hartgerink, E. Beniash, S. I. Stupp – “Self-Assembly and Mineralization of Peptide-Amphiphile Nanofibers”, *Science* 294, 1684 – 1687 (2001).
- ⁹ B. F. Boyce, L. Xing – “Functions of RANKL/RANK/OPG in Bone Modeling and Remodeling”, *Archives in Biochemistry and Biophysics* 473 (2) 139 – 146 (2008).
- ¹⁰ B. F. Boyce, L. Xing – “Biology of RANK, RANKL, and Osteoprotegerin”, *Arthritis Research Therapy* 9 (Suppl 1): S1 (2007).
- ¹¹ J. Wolff – “The Law of Bone Remodeling”, Springer, Berlin (1892).
- ¹² F. Barrere, T. A. Mahmood, K. de Groot, C. A. van Blitterswijk – “Advanced Biomaterials for Skeletal Tissue Regeneration: Instructive and Smart Functions”, *Materials Science and Engineering R* 59 (1-6), 38 - 71 (2008).
- ¹³ T. Asefa, C. Yoshina-Ishii, M. J. MacLachlan, G. A. Ozin – “New Nanocomposites: Putting Organic Function Inside the Channel Walls of Periodic Mesoporous Silica”, *Journal of Materials Chemistry* 10 (8) 1751 – 1755 (2000).
- ¹⁴ G. E. Fantner, T. Hassenkam, J. H. Kindt, J. C. Weaver, H. Birkedal, L. Pechenik, J. A. Cutroni, G. A. G. Cidade, G. D. Stucky, D. E. Morse, P. K. Hansma – “Sacrificial Bonds and Hidden Length Dissipate Energy as Mineralized Fibrils Separate During Bone Fracture”, *Nature Material* 4, 612 - 616 (2005).
- ¹⁵ F.-Z. Cui, J. Ge – “New Observations of the Hierarchical Structure of Human Enamel, from Nanoscale to Microscale”, *Journal of Tissue Engineering and Regenerative Medicine* 1, 185 – 191 (2007).
- ¹⁶ G. E. Fantner, E. Oroudjev, G. Schitter, L. S. Golde, P. Thurner, M. M. Finch, P. Turner, T. Gutschmann, D. E. Morse, H. Hansma, P. K. Hansma – “Sacrificial Bonds and Hidden length: Unraveling Molecular Mesostructures in Tough Materials”, *Biophysical Journal* 90, 1411 – 1418 (2006).
- ¹⁷ V. Uskoković – “Nanomaterials and Nanotechnologies: Approaching the Crest of this Big Wave”, *Current Nanoscience* 4, 119 – 129 (2008).

-
- ¹⁸ F. Nudelman, P. Bomans, K. Pieterse, L. Brylka, G. de With, N. Sommerdijk – “Early Stages of Collagen Mineralization Studied by Cryo-TEM: Starting at the Overlap Region?”, Presented at the Materials Research Society Spring Meeting, San Francisco, CA (2009).
- ¹⁹ W. J. Landis – “Mineral characterization in calcifying tissues: atomic, molecular and macromolecular perspectives”, *Connective Tissue Research* 34 (4) 239 - 46 (1996).
- ²⁰ V. Uskoković, L. E. Bertassoni – “Nanotechnology in Dental Sciences: Moving towards a Finer Way of Doing Dentistry”, *Materials* 3 (3) 1674 – 1691 (2010).
- ²¹ P. Ball – “Water as an Active Constituent in Cell Biology”, *Chemical Reviews* 108 (1) 74 – 108 (2008).
- ²² L. Bertassoni, S. Habelitz, J. Kinney, S. Marshall, G. Marshall – “Biomechanistic Perspective on the Remineralization of Dentin”, *Caries Research* 43, 70 - 77 (2009).
- ²³ C. E. Hoffler, X. E. Guo, P. K. Zysset, S. A. Goldstein – “An application of nanoindentation technique to measure bone tissue lamellae properties”, *Journal of Biomechanical Engineering* 127, 1046-53 (2005).
- ²⁴ Bembey, A.K.; Bushby, A.J.; Boyde, A.; Ferguson, V.L. Oyen, M.L. Hydration effects on the micro-mechanical properties of bone. *Journal of Materials Research* 21, 1962-1968 (2006).
- ²⁵ A. Kishen, S. Vedantam – “Hydromechanics in Dentine: Role of Dentinal Tubules and Hydrostatic Pressures on Mechanical Stress-Strain Distribution”, *Dental Materials* 23 (10) 1296 – 1306 (2007).
- ²⁶ Fawzy A.S.; Farghaly, A.M. Probing nano-scale adhesion force between AFM and acid demineralized intertubular dentin: Moist versus dry dentin. *Journal of Dentistry* 37, 963-9 (2009).
- ²⁷ A. Cooper- »Thermodynamics of protein folding and stability«, In: *Protein: A Comprehensive Treatise* Vol. 2, pp. 217 – 270, edited by Geoffrey Allen, JAI Press, Stamford, CN (1999).
- ²⁸ Z. L. Shen, M. R. Dodge, H. Kahn, R. Ballarini, S. J. Eppel – “In Vitro Mechanical Testing of Collagen Nanofibrils”, Society for Biomaterials 2010 Annual Meeting & Exposition, Seattle, WA (2010), presentation #220.
- ²⁹ S. V. Dorozhkin – “Calcium Orthophosphates”, *Journal of Materials Science* 42, 1061 – 1095 (2007).
- ³⁰ V. Uskoković, M. Drogenik – “Synthesis of Lanthanum-Strontium Manganites by Oxalate-Precursor Co-Precipitation Methods in Solution and in Reverse Micellar Microemulsion”, *Journal of Magnetism and Magnetic Materials* 303 (1) 214 – 220 (2006).
- ³¹ V. Uskoković – “Theoretical and Practical Aspects of Colloid Science and Self-Assembly Phenomena Revisited”, *Reviews in Chemical Engineering* 23 (5) 301 - 372 (2007).
- ³² K. Onuma, A. Ito – “Cluster Growth Model for Hydroxyapatite”, *Chemistry of Materials* 10 (11) 3346 – 3351 (1998).
- ³³ K. Onuma – “Recent research on pseudobiological hydroxyapatite crystal growth and phase transition mechanisms”, *Progress in Crystal Growth and Characterization of Materials* 52 (3) 223 – 245 (2006).
- ³⁴ T. J. White, Z. L. Dong – “Structural Derivation and Crystal Chemistry of Apatites”, *Acta Crystallographia* B59, 1 – 16 (2003).
- ³⁵ N. H. De Leeuw – “Local Ordering of Hydroxyl Groups in Hydroxyapatite”, *Chemistry Communication* 17, 1646 – 1647 (2001).

-
- ³⁶ R. Z. LeGeros – “Calcium Phosphate-Based Osteoinductive Materials”, *Chemical Society Reviews* 108 (11) 4742 – 4753 (2008).
- ³⁷ J. D. Featherstone – “Modeling the caries-inhibitory effects of dental materials”, *Dental Materials* 12 (3) 194 – 197 (1996).
- ³⁸ R. L. Isaacson, J. A. Varner, K. F. Jensen – “Toxin-induced blood vessel inclusions caused by the chronic administration of aluminum and sodium fluoride and their implications for dementia”, *Annals of the New York Academy of Sciences* 15, 152 – 166 (1997).
- ³⁹ P. R. Garant – “Oral Cells and Tissues”, Quintessence, Carol Stream, IL (2003).
- ⁴⁰ F. Yao, J. LeGeros, R. Z. LeGeros – “Simultaneous Incorporation of Carbonate and Fluoride in Synthetic Apatites: Effect on Crystallographic and Physico-Chemical Properties”, *Acta Biomaterialia* 5, 2169 – 2184 (2009).
- ⁴¹ A. Porter, N. Patel, R. Brooks, S. Best, N. Rushton, W. Bonfield – “Effect of Carbonate Substitution on the Ultrastructural Characteristics of Hydroxyapatite Implants”, *Journal of Materials Science: Materials in Medicine* 16 (10) 899 – 907 (2005).
- ⁴² T. Kanno, J. I. Horiuchi, M. Kobayashi, Y. Motogami, T. Akazawa – “Characteristics of the Carbonate Ions Incorporated into Calcium-Partially-Strontium-Substituted and Strontium Apatites”, *Journal of Materials Science Letters* 18, 1343 – 1345 (1999).
- ⁴³ F. C. M. Driessens, R. M. H. Verbeeck, L. P. Kiekens – “Mechanism of Substitution in carbonated Apatites”, *Zeitschrift für Anorganische und Allgemeine Chemie* 504 (9) 195 – 200 (1983).
- ⁴⁴ P. Somasundaran – “Encyclopedia of Surface and Colloid Science”, CRC Press, Boca Raton, FL (2004), pp. 567.
- ⁴⁵ J. Barralet, S. Best, W. Bonfield – “Carbonate Substitution in Precipitated Hydroxyapatite: An Investigation into the Effects of Reaction Temperature and Bicarbonate Ion Concentration”, *Journal of Biomedical Materials Research* 41 (1) 79 – 86 (1998).
- ⁴⁶ C. Rey, B. Collins, T. Goehl, I. R. Dickson, M. J. Glimcher – “The Carbonate Environment in Bone Mineral: A Resolution-Enhanced Fourier Transform Infrared Spectroscopic Study”, *Calcified Tissue International* 45, 157 – 164 (1989).
- ⁴⁷ W. H. Emerson, E. D. Fischer – “The Infrared Absorption Spectra of Carbonate in Calcified Tissue”, *Archives in Oral Biology* 7, 671 – 683 (1962).
- ⁴⁸ Y. Suetsugu, Y. Takanashi, F. P. Okamura, J. Tanaka – “Structure Analysis of A-Type Carbonate Apatite by a Single-Crystal X-Ray Diffraction Method”, *Journal of Solid State Chemistry* 155, 292 – 304 (2000).
- ⁴⁹ M. Corno, A. Rimola, V. Bolis, P. Ugliengo – “Hydroxyapatite as a Key Biomaterial: Quantum-Mechanical Simulation of its Surfaces in Interaction with Biomolecules”, *Physical Chemistry Chemical Physics* 12, 6309 – 6329 (2010).
- ⁵⁰ T. Tonegawa, T. Ikoma, T. Yoshioka, N. Hanagata, J. Tanaka – “Crystal Structure Refinement of A-Type Carbonate Apatite by X-Ray Powder Diffraction”, *Journal of Materials Science* 45, 2419 – 2426 (2010).
- ⁵¹ M. E. Fleet, X. Lui, P. King – “Accommodation of the Carbonate Ion in Apatite: An FTIR and X-Ray Structure Study of Crystals Synthesized at 2-4 GPa”, *American Mineralogy* 89, 1422 – 1434 (2004).
- ⁵² D. Shi – “Introduction to Biomaterials”, World Scientific Press, Singapore (2006).

-
- ⁵³ S. Brown, I. Clarke, P. Williams – “Synthesis and Characterization of Hydroxyapatite and Fluoroapatite”, *Key Engineering Materials* 218-2, 35 – 38 (2002).
- ⁵⁴ N. Ignjatović, E. Suljovrujić, Z. Stojanović, D. Uskoković - “Structure and characteristics of the hot pressed hydroxyapatite/poly-L-lactide composite”, *Science of Sintering* 34, 79-93 (2002).
- ⁵⁵ Y. Fang, D. K. Agrawal, D. M. Roy – “Thermal Stability of Synthetic Hydroxyapatite”, In: *Hydroxyapatite and Related Materials*, edited by P. W. Brown & B. Constantz, pp. 269 – 282, CRC Press, Boca Raton, FL (1994).
- ⁵⁶ N. Ignjatović, E. Suljovrujić, J. Budinski, I. Krakovsky, D. Uskoković - “Evaluation of Hot Pressed Hydroxyapatite/Poly-l-lactide Composite Biomaterial Characteristics”, *Journal of Biomedical Materials Research Part B: Applied Biomaterials* 71B, 284-294 (2004).
- ⁵⁷ T. Welzel, W. Meyer-Zaika, M. Epple – “Continuous preparation of functionalised calcium phosphate nanoparticles with adjustable crystallinity”, *Chemical Communication* 1204 – 1205 (2004).
- ⁵⁸ M. Pretto, A. L. Costa, E. Landi, A. Tampieri, C. Galassi – “Dispersing Behavior of Hydroxyapatite Powders Produced by Wet-Chemical Synthesis”, *Journal of the American Ceramic Society* 86 (9) 1534 – 1539 (2003).
- ⁵⁹ J. Gomez-Morales, J. Torrent-Burgues, T. Boix, J. Fraile, R. Rodriguez-Clemente – “Precipitation of Stoichiometric Hydroxyapatite by a Continuous Method”, *Crystal Research and Technology* 36 (1) 15 – 26 (2001).
- ⁶⁰ A. C. Tas, F. Korkusuz, M. Timucin, N. Akkas – “An Investigation of the Chemical Synthesis and High-Temperature Sintering Behaviour of Calcium Hydroxyapatite (HA) and Tricalcium Phosphate (TCP) Bioceramics”, *Journal of Materials Science – Materials in Medicine* 8 (2) 91 – 96 (1997).
- ⁶¹ M. Ashok, S. N. Kalkura, N. M. Sundaram, D. Arivuoli – “Growth and Characterization of Hydroxyapatite Crystals by Hydrothermal Method”, *Journal of Materials Science: Materials in Medicine* 18, 895 – 898 (2007).
- ⁶² Dj. Janačković, I. Janković, R. Petrović, Lj. Kostić-Gvozdrenović, S. Milonjić, D. Uskoković - “Surface properties of HAP particles obtained by hydrothermal decomposition of urea and calcium-edta chelates”, *Key Engineering Materials* 240-242, 437-440 (2003).
- ⁶³ E. S. Thian, Z. Ahmad, J. Huang, M. J. Edirisinghe, S. N. Jayasinghe, D. C. Ireland, R. A. Brooks, N. Rushton, W. Bonfield, S. M. Best – “The role of electrosprayed apatite nanocrystals in guiding osteoblast behavior”, *Biomaterials* 29 (12) 1833 – 1843 (2008).
- ⁶⁴ Y. Wu, L. L. Hench, J. Du. K.-L. Choy, J. Guo – “Preparation of Hydroxyapatite Fibers by Electrospinning Technique”, *Journal of the American Ceramic Society* 87 (10) 1988 – 1991 (2005).
- ⁶⁵ K. Teshima, S. Lee, M. Sakurai, Y. Kamenno, K. Yubuta, T. Suzuki, T. Shishido, M. Endo, S. Oishi – “Well-Formed One-Dimensional Hydroxyapatite Crystals Grown by an Environmentally Friendly Flux Method”, *Crystal Growth & Design* 9 (6) 2937 – 2940 (2009).
- ⁶⁶ V. Jokanović, D. Uskoković - “Calcium Hydroxyapatite Thin Films on Titanium Substrates Prepared by Ultrasonic Spray Pyrolysis”, *Materials Transactions* 46, 228-235 (2005).
- ⁶⁷ Y. Fang, D. K. Agrawal, D. M. Roy, R. Roy, P. W. Brown – “Ultrasonically Accelerated Synthesis of Hydroxyapatite”, *Journal of Materials Research* 7 (8) 2294 - 2298 (1992).
- ⁶⁸ I. Nikčević, V. Jokanović, M. Mitrić, Z. Nedić, D. Makovec, D. Uskoković - “Mechanochemical Synthesis of Nanostructured Fluorapatite/Fluorhydroxyapatite and Carbonated Fluorapatite/ Fluorhydroxyapatite”, *Journal of Solid State Chemistry* 177, 2565-2574 (2004).

-
- ⁶⁹ S.-H. Rhee – “Synthesis of Hydroxyapatite via Mechanochemical Treatment”, *Biomaterials* 23 (4) 1147 – 1152 (2002).
- ⁷⁰ A. Lak, M. Mazloumi, M. S. Mohajerani, S. Zanganeh, M. R. Shayegh, A. Kajbafvala, H. Arami, S. K. Sadrnezhaad – “Rapid Formation of Mono-Dispersed Hydroxyapatite Nanorods with Narrow-Size Distribution via Microwave Irradiation”, *Journal of the American Ceramic Society* 91 (11) 3580 – 3584 (2008).
- ⁷¹ J. Liu, K. Li, H. Wang, M. Zhu, H. Yan – “Rapid formation of hydroxyapatite nanostructures by microwave irradiation”, *Chemical Physics Letters* 396, 429 – 432 (2004).
- ⁷² H. G. Zhang, Q. Zhu – “Surfactant-Assisted Preparation of Fluoride-Substituted Hydroxyapatite Nanorods”, *Materials Letters* 59, 3054 – 3058 (2005).
- ⁷³ S. Bose, S. K. Saha – »Synthesis and Characterization of Hydroxyapatite Nanopowders by Emulsion Technique«, *Chemistry of Materials* 15, 4464 – 9 (2003).
- ⁷⁴ J. Z. Liu, B. D. Gou, S. J. Xu, K. Wang – »Formation of Calcium Phosphate Mineral Material Controlled by Microemulsion«, *Progress in Natural Science* 12 (8) 615 – 7 (2002).
- ⁷⁵ S. Sarda, M. Heughebaert, A. Lebugle – »Influence of the Type of Surfactant on the Formation of Calcium Phosphate in Organized Molecular Systems«, *Chemistry of Materials* 11 (10) 2722 – 7 (1999).
- ⁷⁶ A. Banerjee, A. Bandyopadhyay, S. Bose – “Hydroxyapatite Nanopowders: Synthesis, Densification and Cell-Materials Interaction”, *Materials Science & Engineering C* 27 (4) 729 – 735 (2007).
- ⁷⁷ K. Cheng, W. Weng, G. Han, P. Du, G. Shen, J. Yang, J. M. F. Ferreira – “The Effect of Triethanolamine on the Formation of Sol-Gel Derived Fluoroapatite/Hydroxyapatite Solid Solution”, *Materials Chemistry and Physics* 78 (3) 767 – 771 (2003).
- ⁷⁸ C. L. Chu, P. H. Lin, S. Dong, D. Y. Guo – “Influences of citric acid as a chelating reagent on the characteristics of nanophase hydroxyapatite powders fabricated by a sol-gel method”, *Journal of Materials Science Letters* 21, 1793 – 1795 (2002).
- ⁷⁹ Y. Gao – “Synthesis and Characterization of Calcium Phosphate Coatings by metalorganic Chemical vapor Deposition”, Presented at the Materials Research Society Meeting, Boston, MA (1998).
- ⁸⁰ S. Xu, J. Long, L. Sim, C. H. Diong, K. Ostrikov – “RF Plasma Sputtering Deposition of Hydroxyapatite Bioceramics: Synthesis, Performance, and Biocompatibility”, *Plasma Processing and Polymers* 2 (5) 373 – 390 (2005).
- ⁸¹ F. Watari, A. Yokoyama, M. Gelinsky, W. Pompe – “Conversion of functions by nanosizing - from osteoconductivity to bone substitutional properties in apatite”, In: *Interface Oral Health Science 2007, Proceedings of the 2nd International Symposium for Interface Oral Health Science, Sendai, Japan*, pp. 139 – 148, Springer, Berlin (2007).
- ⁸² V. Uskoković – “Challenges for the Modern Science in its Descend towards Nano Scale”, *Current Nanoscience* 5 (3) 372 – 389 (2009).
- ⁸³ Y. Cai, R. Tang – “Calcium phosphate nanoparticles in biomineralization and biomaterials”, *Journal of Materials Chemistry* 18, 3775 – 3787 (2008).
- ⁸⁴ Y. Cai, Y. Liu, W. Yan, Q. Hu, J. Tao, M. Zhang, Z. Shi, R. Tang – “Role of hydroxyapatite nanoparticle size in bone cell proliferation”, *Journal of Materials Chemistry* 17, 3780 – 3787 (2007).

-
- ⁸⁵ K. K. Perkin, J. L. Turner, K. L. Wooley and S. Mann – “Fabrication of Hybrid Nanocapsules by Calcium Phosphate Mineralization of Shell Cross-Linked Polymer Micelles and Nanocages”, *Nano Letters* 5 (7) 1457 – 1461 (2005).
- ⁸⁶ D. G. Shchukin, G. B. Sukhorukov and H. Mohwald - “Biomimetic Fabrication of Nanoengineered Hydroxyapatite/Polyelectrolyte Composite Shell”, *Chemistry of Materials* 15 (20) 3947 – 3950 (2003).
- ⁸⁷ J. Kirkham, S. J. Brookes, R. C. Shore, S. R. Wood, D. A. Smith, J. Zhang, H. Chen, C. Robinson – “Physico-Chemical Properties of Crystal Surfaces in Matrix-Mineral Interactions During Mammalian Biomineralisation”, *Current Opinion in Colloid & Interface Science* 7, 124 – 132 (2002).
- ⁸⁸ B. Müller – “Tailoring Biocompatibility: Benefitting Patients”, *Materials Today* 13 (4) 58 (2010).
- ⁸⁹ A. Rimola, M. Corno, C. M. Zicovich-Wilson, P. Ugliengo – “*Ab Initio* Modeling of Protein/Biomaterial Interactions: Glycine Adsorption at Hydroxyapatite Surfaces”, *Journal of the American Chemical Society* 130 (48) 16181 – 16183 (2008).
- ⁹⁰ C. Tamerler, M. Sarikaya – “Molecular Biomimetics – Peptide-Based Materials for Nanotechnology and Medicine”, Society for Biomaterials 2010 Annual Meeting & Exposition, Seattle, WA (2010), presentation #182.
- ⁹¹ V. Uskoković, M.-K. Kim, W. Li, S. Habelitz – “Enzymatic Processing of Amelogenin during Continuous Crystallization of Apatite”, *Journal of Materials Research* 32, 3184 – 3195 (2008).
- ⁹² G. Ahmad, M. B. Dickerson, B. C. Church, Y. Cai, S. E. Jones, R. R. Naik, J. S. King, C. J. Summers, N. Kröger, K. H. Sandhage – “Rapid, Room-Temperature Formation of Crystalline Calcium Molybdate Phosphor Microparticles via Peptide-Induced Precipitation”, *Advanced Materials* 18, 1759 – 1763 (2006).
- ⁹³ M. B. Dickerson, R. N. Naik, M. O. Stone, Y. Cai, K. H. Sandhage – “Identification of Peptides that Promote the Rapid Precipitation of Germania Nanoparticle Networks via Use of a Peptide Display Library”, *Chemical Communications (Camb)* 15, 1776 – 1777 (2004).
- ⁹⁴ J. Zhan, Y.-H. Tseng, J. C. C. Chan, C.-Y. Mou – “Biomimetic Formation of Hydroxyapatite Nanorods by a Single-Crystal-to-Single-Crystal Transformation”, *Advanced Functional Materials* 15 (12) 2005 – 2010 (2005).
- ⁹⁵ F. Huang, Y. Shen, A. Xie, J. Zhu, C. Zhang, S. Li, J. Zhu – “Study on Synthesis and Properties of Hydroxyapatite Nanorods and its Complex Containing Biopolymer”, *Journal of Materials Science* 42 (20) 8599 – 8605 (2007).
- ⁹⁶ F. Zhang, Z.-H. Zhou, S.-P. Yang, L.-H. Mao, H.-M. Chen, X.-B. Yu – “Hydrothermal synthesis of hydroxyapatite nanorods in the presence of anionic starburst dendrimer”, *Materials Letters* 59 (11) 1422 – 1425 (2005).
- ⁹⁷ L. Yan, Y. Li, Z.-X. Deng, J. Zhuang, X. Sun – “Surfactant-Assisted Hydrothermal Synthesis of Hydroxyapatite Nanorods”, *International Journal of Inorganic Materials* 3 (7) 633 – 637 (2001).
- ⁹⁸ A. Wang, D. Liu, H. Yin, H. Wu, Y. Wada, M. Ren, T. Jiang, X. Cheng, Y. Xu – “Size-controlled synthesis of hydroxyapatite nanorods by chemical precipitation in the presence of organic modifiers”, *Materials Science and Engineering C* 27, 865 – 869 (2007).
- ⁹⁹ D. Walsh, J. L. Kingston, B. R. Heywood, S. Mann – “Influence of Monosaccharides and Related Molecules on the Morphology of Hydroxyapatite”, *Journal of Crystal Growth* 133, 1 – 12 (1993).
- ¹⁰⁰ S. I. Stupp, P. V. Braun – “Molecular Manipulation of Microstructures: Biomaterials, Ceramics, and Semiconductors”, *Science* 277, 1242 – 1248 (1997).

-
- ¹⁰¹ R. Zangi, B. J. Berne – “Aggregation and Dispersion of Small Hydrophobic Particles in Aqueous Electrolyte Solutions”, *Journal of Physical Chemistry B* 110, 22736 – 22741 (2006)
- ¹⁰² S. Mann – “Biomineralization: Principles and Concepts in Bioinorganic Materials Chemistry”, Oxford University Press, Oxford, UK (2001).
- ¹⁰³ M. Motskin, D.M. Wright, K. Muller, N. Kyle, T.G. Gard, A.E. Porter, J.N. Skepper – “Hydroxyapatite nano and microparticles: Correlation of particle properties with cytotoxicity and biostability”, *Biomaterials* 30 (19) 3307 – 3317 (2009).
- ¹⁰⁴ Y. Fan, Z. Sun, J. Moradian-Oldak – “Controlled remineralization of enamel in the presence of amelogenin and fluoride”, *Biomaterials* 30 (4) 478 – 483 (2009).
- ¹⁰⁵ D. P. Uskoković - “Fine Particles in Advanced Technology”, International Symposium on Advanced Dielectric Materials and Electronic Devices, MS&T 2006 Conference Proceedings, Warrendale, PA, Materials & Systems 1, 599-610 (2006).
- ¹⁰⁶ Y. Tanaka, Y. Hirata, R. Yoshinaka – “Synthesis and Characterization of Ultrafine Hydroxyapatite Particles”, *Journal of Ceramic Processing Research* 4 (4) 197 – 201 (2003).
- ¹⁰⁷ V. Uskoković, M. Drogenik – “Synthesis of Materials within Reverse Micelles”, *Surface Review and Letters* 12 (2) 239 – 277 (2005).
- ¹⁰⁸ V. Uskoković, M. Drogenik – “Reverse Micelles: Inert Nano-Reactors or Physico-Chemically Active Guides of the Capped Reactions”, *Advances in Colloid and Interface Science* 133 (1) 23 – 34 (2007).
- ¹⁰⁹ F. Ye, H. Guo, H. Zhang – “Biomimetic Synthesis of Oriented Hydroxyapatite Mediated by Nonionic Surfactants”, *Nanotechnology* 19, 245605 (2008).
- ¹¹⁰ C. Santos, C. F. Rovath, R.-P. Franke, M. M. Almeida, M. E. V. Costa – “Spray-dried hydroxyapatite-5-Fluorouracil granules as a chemotherapeutic delivery system”, *Ceramics International* 35, 509 – 513 (2009).
- ¹¹¹ H. Chen, B. H. Clarkson, K. Sun and J. F. Mansfield – “Self-Assembly of Synthetic Hydroxyapatite Nanorods into an Enamel prism-Like Structure”, *Journal of Colloid and Interface Science* 288 (2) 423 – 431 (2005).
- ¹¹² Y. Cai, H. Pan, X. Xu, Q. Hu, L. Li, R. Tang - “Ultrasonic Controlled Morphology Transformation of Hollow Calcium Phosphate Nanospheres. A Smart and Biocompatible Drug Release System”, *Chemistry of Materials* 19, 3081 – 3083 (2007).
- ¹¹³ N. Ignjatović, P. Ninkov, Z. Ajduković, D. Vasiljević-Radović, D. Uskoković - “Biphasic calcium phosphate coated with poly-d,l-lactide-co-glycolide biomaterial as a bone substitute”, *Journal of the European Ceramic Society* 27 (2-3) 1589-1594 (2007).
- ¹¹⁴ J. C. Hornez, F. Chai, F. Monchau, N. Blanchemain, M. Descamps, H. F. Hilderbrand – “Biological and Physico-Chemical Assessment of Hydroxyapatite (HA) with Different Porosity”, *Biomolecular Engineering* 24 (5) 505 – 509 (2007).
- ¹¹⁵ S. Hesaraki, R. Nemati – “Caphalexin-Loaded Injectable Macroporous Calcium Phosphate Bone Cement”, *Journal of Biomedical Materials Research Part B – Applied Biomaterials* 89B (2) 342 – 352 (2009).
- ¹¹⁶ N. L. Ignjatović, Z. R. Ajduković, V. P. Savić, D. P. Uskoković – “Size effect of calcium phosphate coated with poly-DL-lactide- co-glycolide on healing processes in bone reconstruction”, *Journal of Biomedical Materials Research Part B – Applied Biomaterials* 94 (1) 108 – 117 (2010).

-
- ¹¹⁷ F. Watari, N. Takashi, A. Yokoyama, M. Uo, T. Akasaka, Y. Sato, A. Abe, Y. Totsuoka, K. Tohji – “Material nanosizing effect on living organisms: non-specific, biointeractive, physical size effects”, *Journal of the Royal Society Interface* 6, S371 – S388 (2009).
- ¹¹⁸ M. Šupova – “Problem of Hydroxyapatite Dispersion in Polymer Matrices: A Review”, *Journal of Materials Science: Materials in Medicine* 20, 1201 – 1213 (2009).
- ¹¹⁹ M. Jevtić, A. Radulović, N. Ignjatović, M. Mitrić, D. Uskoković - "Controlled assembly of poly(D,L-lactide-co-glycolide)/hydroxyapatite core-shell nanospheres under ultrasonic irradiation", *Acta Biomaterialia* 5, 208-218 (2009).
- ¹²⁰ M. Jevtić, D. Uskoković - “Influence of Urea as a Homogenous Precipitation Agent on Sonochemical Hydroxyapatite Synthesis”, *Materials Science Forum* 555, 285 – 290 (2007).
- ¹²¹ M. Aizawa, A. E. Porter, S. M. Best, W. Bonfield – “Ultrastructural Observation of Single-Crystal Apatite Fibres”, *Biomaterials* 26, 3427 – 3433 (2005).
- ¹²² S. Y. Chung, Y. M. Kim, J. G. Kim, Y. J. Kim – “Multiphase Transformation and Ostwald’s Rule of Stages During Crystallization of a Metal Phosphate”, *Nature Physics* 5 (1) 68 – 73 (2009).
- ¹²³ E. Beniash, R. A. Metzler, R. S. K. Lam, P. U. P. A. Gilbert – “Transient Amorphous Calcium Phosphate in Forming Enamel”, *Journal of Structural Biology* 166, 133 – 143 (2009).
- ¹²⁴ A. George – “Protein Template is Required for Nucleation and Orientation of Hydroxyapatite in Bones and Teeth”, Presented at the 12th Annual Conference of the Institute of Science and Technology of Ceramics ISTEC-CNR, Faenza (2009).
- ¹²⁵ G. He, T. Dahl, A. Veis, A. George – “Nucleation of Apatite Crystals in vitro by Self-Assembled Dentin Matrix Protein 1.”, *Nature Materials* 2 (8) 552 – 558 (2003).
- ¹²⁶ L. B. Gower – “Biomimetic Model Systems for Investigating the Amorphous Precursor Pathway and its Role in Biomineralization”, *Chemical Reviews* 108 (11) 4551 – 4627 (2008).
- ¹²⁷ J. Tao, H. Pan, Y. Zeng, X. Xu and R. Tang – “Roles of Amorphous Calcium Phosphate and Biological Additives in the Assembly of Hydroxyapatite Nanoparticles”, *Journal of Physical Chemistry B* 111 (47) 13410 - 13418 (2007).
- ¹²⁸ D. W. Thompson – “On Growth and Form”, Dover, London, UK (1917).
- ¹²⁹ Y. C. Chien, D. L. Masica, J. J. Gray, S. Nguyen, H. Vali, M. D. McKee – “Modulation of calcium oxalate dihydrate growth by selective crystal-face binding of phosphorylated osteopontin and polyaspartate peptide showing occlusion by sectoral (compositional) zoning”, *Journal of Biological Chemistry* 284 (35) 23491 – 23501 (2009).
- ¹³⁰ L. H. He, M. V. Swain – “Understanding the mechanical behaviour of human enamel from its structural and compositional characteristics”. *Journal of Mechanical Behavior of Biomedical Materials* 1, 18 – 29 (2008).
- ¹³¹ E. Beniash, R.A. Metzler, R.S. Lam, P.U. Gilbert – “Transient amorphous calcium phosphate in forming enamel”, *Journal of Structural Biology* 166 (2009) 133-143.
- ¹³² M. Cao, Y. Wang, C. Guo, Y. Qi, C. Hu – “Preparation of Ultrahigh-Aspect-Ratio Hydroxyapatite Nanofibers in Reverse Micelles under Hydrothermal Conditions”, *Langmuir* 20, 4784 – 4786 (2004).
- ¹³³ V. Uskoković – “Prospects and Pits on the Path of Biomimetics: The Case of Tooth Enamel”, *International Journal of Design Engineering, Special Issue: Design in Nature* (in press, 2010).

-
- ¹³⁴ C. Viney – “Self-Assembly as a Route to Fibrous Materials: Concepts, Opportunities and Challenges”, *Current Opinion in Solid State and Materials Science* 8, 95 – 101 (2004).
- ¹³⁵ C. Viney, F. I. Bell – “Inspiration versus Duplication with Biomolecular Fibrous Materials: Learning Nature's Lessons without Copying Nature's Limitations”, *Current Opinion in Solid State and Materials Science* 8, 165 – 171 (2004).
- ¹³⁶ G. Bateson – “Mind and Nature”, Hampton Press, Cresskill, NJ (1979).
- ¹³⁷ A.-W. Xu, Y. Ma, H. Cölfen – “Biomimetic Mineralization”, *Journal of Materials Chemistry* 17, 415 – 449 (2007).
- ¹³⁸ C. A. Orme, J. L. Giocondi – “Model Systems for Formation and Dissolution of Calcium Phosphate Minerals”, In: *Handbook of Biomineralization Vol.2*, edited by P. Behrens & E. Bäuerlein, Wiley, Weinheim (2007).
- ¹³⁹ I. S. Harding, N. Rashid, K. A. Hing – “Surface charge and the effect of excess calcium ions on the hydroxyapatite surface”, *Biomaterials* 26, 6818–6826 (2005).
- ¹⁴⁰ P. Somasundaran, Y. H. C. Wang – “Surface Chemical Characteristics and Adsorption Properties of Apatite”, In: *Adsorption on and Surface Chemistry of Apatite*, edited by D. N. Misra, Springer, Berlin (1984).
- ¹⁴¹ A. Bengtsson, A. Shchukarev, P. Persson, S. Sjöberg – “Phase Transformations, Ion-Exchange, Adsorption, and Dissolution Processes in Aquatic Fluoroapatite Systems”, *Langmuir* 25 (4) 2355 – 2362 (2009).
- ¹⁴² S. Cazalbou, C. Combes, D. Eichert, C. Rey – “Adaptative Physico-Chemistry of Bio-Related Calcium Phosphates”, *Journal of Materials Chemistry* 14 (14) 2148 – 2153 (2004).
- ¹⁴³ A. A. Campbell, M. LoRe, G. H. Nancollas – “The Influence of Carbonate and Magnesium Ions on the Growth of Hydroxyapatite, Carbonated Apatite and Human Powdered Enamel”, *Colloids and Surfaces* 54, 25 – 31 (1991).
- ¹⁴⁴ N. Kanzaki, K. Onuma, A. Ito, K. Teraoka, T. Tateishi, S. Tsutsumi – “Direct Growth Rate Measurement of Hydroxyapatite Single Crystal by Moire Phase Shift Interferometry”, *Journal of Physical Chemistry* 102 (34) 6471 – 6476 (1998).
- ¹⁴⁵ M. Rouahi, E. Champion, O. Gallet, A. Jada, K. Anselme – “Physico-chemical characteristics and protein adsorption potential of hydroxyapatite particles: Influence on in vitro biocompatibility of ceramics after sintering”, *Colloids and Surfaces B: Biointerfaces* 47 (1) 10 – 19 (2006).
- ¹⁴⁶ H.-M. Kim, T. Himeno, T. Kokubo, T. Nakamura – “Process and kinetics of bonelike apatite formation on sintered hydroxyapatite in a simulated body fluid”, *Biomaterials* 26 (21) 4366 – 4373 (2005).
- ¹⁴⁷ E. V. Roseeva, O. A. Golovanova, O. V. Frank-Kamenetskaya – “The Influence of Amino Acids on the Formation of Nanocrystalline Hydroxyapatite”, *Glass Physics and Chemistry* 33 (3) 283 – 286.
- ¹⁴⁸ A. Barroug, J. Lemaitre, P. G. Rouxhet – “Influence of Crystallite Size on the Surface-Properties of Calcium-Deficient Hydroxyapatites”, *Journal of Alloys and Compounds* 188 (1-2) 152 – 156 (1992).
- ¹⁴⁹ M. Kosmulski – “Surface Charging and Points of Zero Charge”, *Surfactant Science Series* 145, CRC Press, Francis & Taylor, Boca Raton, FL (2009).
- ¹⁵⁰ Y. Zhang, Y. Yokogawa – “Effect of Drying Conditions During Synthesis on the Properties of Hydroxyapatite Powders”, *Journal of Materials Science: Materials in Medicine* 19, 623 – 628 (2008).
- ¹⁵¹ X. Yao, S. Tan, D. Jiang – “Fabrication of hydroxyapatite ceramics with controlled pore characteristics by slip casting”, *Journal of Materials Science: Materials in Medicine* 16, 161 – 165 (2005).

-
- ¹⁵² J. Ma, C. H. Liang, L. B. Kong, C. Wang – “Colloidal Characterization and Electrophoretic Deposition of Hydroxyapatite on Titanium Substrate”, *Journal of Materials Science: Materials in Medicine* 14, 797 – 801 (2003).
- ¹⁵³ Z. Sadeghian, J. G. Heinrich, F. Moztarzadeh – “Preparation of highly concentrated aqueous hydroxyapatite suspensions for slip casting”, *Journal of Materials Science* 40 (17) 4619 – 4623 (2005).
- ¹⁵⁴ T. Kawasaki, M. Niikura, Y. Kobayashi – “Fundamental study of hydroxyapatite high-performance liquid chromatography: II. Experimental analysis on the basis of the general theory of gradient chromatography”, *Journal of Chromatography* 515, 91 – 123 (1990).
- ¹⁵⁵ A. Kandori, S. Oda, M. Fukusumi, Y. Morisada – “Synthesis of positively charged calcium hydroxyapatite nanocrystals and their adsorption behavior of proteins”, *Colloids and Surfaces B: Biointerfaces* 73, 140 – 145 (2009).
- ¹⁵⁶ S. Boghak, S. Bose, A. Bandyopadhyay – “Electro-Thermally Polarized Hydroxyapatite (HAp) Ceramics: Influence of MgO, SrO, and ZnO Dopants”, Society for Biomaterials 2010 Annual Meeting & Exposition, Seattle, WA (2010), presentation #474.
- ¹⁵⁷ Y. Cai, J. Yao – “Effect of Proteins on the Synthesis and Assembly of Calcium Phosphate Nanomaterials”, *Nanoscale* (in press, 2010).
- ¹⁵⁸ D. J. Shaw – “Introduction to Colloid and Surface Chemistry”, Butterworth Heinemann, Oxford, UK (1992).
- ¹⁵⁹ S. V. Dorozhkin – “Calcium Orthophosphates in Nature, Biology and Medicine”, *Materials* 2 (2) 399 – 498 (2009).
- ¹⁶⁰ J. L. Giocondi, G. H. Nancollas, A. A. Chernov, C. Orme – “Does the Incorporation of Calcium or Phosphate Control the Rate of Brushite Mineralization?”, Presented at the Materials Research Society Spring Meeting, San Francisco, CA (2009).
- ¹⁶¹ E. Leontidis – “Hofmeister Anion Effects on Surfactant Self-Assembly and the Formation of Mesoporous Solids”, *Current Opinion in Colloid and Interface Science* 7, 81 – 91 (2002).
- ¹⁶² A. Filankembo, S. Giorgio, I. Lisiecki, M. P. Pileni – “Is the Anion the Major Parameter in the Shape Control of Nanocrystals”, *Journal of Physical Chemistry B* 107 (30), 7492 – 7500 (2003).
- ¹⁶³ C. A. Orme, J. L. Giocondi – “The Use of Scanning Probe Microscopy to Investigate Crystal-Fluid Interfaces”, In: *Perspectives on Inorganic, Organic, and Biological Crystal Growth: From Fundamentals to Applications*, edited by M. Skowronski, J. J. DeYoreo, C. A. Wang, Springer, Berlin (2007).
- ¹⁶⁴ T. M. Riddick – “Control of Colloid Stability through Zeta Potential and its Relationship to Cardiovascular Disease”, Livingston Publishing, Wynnewood, PA (1968).
- ¹⁶⁵ P. Ngankam, Ph. Lavalle, J. C. Voegel, L. Szyk, G. Decher, P. Schaaf, F. J. G. Cuisinier – “Influence of Polyelectrolyte Multilayer Films on Calcium Phosphate Nucleation”, *Journal of the American Chemical Society* 122, 8998 – 9005 (2000).
- ¹⁶⁶ S. R. Qiu, C. A. Orme – “Dynamics of Biomineral Formation at the Near-Molecular Level”, *Chemical Review* 108 (11) 4784 – 4822 (2008).
- ¹⁶⁷ A. Bengtsson, A. Shchukarev, P. Persson, S. Sjoberg – “A Solubility and Surface Complexation Study of a Non-Stoichiometric Hydroxyapatite”, *Geochimica et Cosmochimica Acta* 73 (2) 257 – 267 (2009).
- ¹⁶⁸ X. Lu, Y. Leng – “Theoretical Analysis of Calcium Phosphate Precipitation in Simulated Body Fluid”, *Biomaterials* 26 (10) 1097 - 1108 (2005).

-
- ¹⁶⁹ J. H. Harding, D. M. Duffy, M. L. Sushko, P. M. Rodger, D. Quigley, J. A. Elliott – “Computational Techniques at the Organic-Inorganic Interface in Biomineralization”, *Chemical Review* 108, 4823 – 4854 (2008).
- ¹⁷⁰ H. Pan, J. Tao, X. Yu, L. Fu, J. Zhang, X. Zeng, G. Xu, R. Tang – “Anisotropic Demineralization and Oriented Assembly of Hydroxyapatite Crystals in Enamel: Smart Structures of Biominerals”, *Journal of Physical Chemistry B* 112 (24) 7162 – 7165 (2008).
- ¹⁷¹ M. J. J. M. van Kemenade, P. L. de Bruyn – “A Kinetic Study of Precipitation from Supersaturated Calcium Phosphate Solutions”, *Journal of Colloid and Interface Science* 118 (2) 564 – 585 (2001).
- ¹⁷² A. Szczes, L. Holysz, E. Chibowski – “Influence of magnetic field on the properties of freshly precipitated calcium phosphate: implication in adhesion to glass surface”, *Journal of Adhesion Science and Technology* 20 (4) 345 – 358 (2006).
- ¹⁷³ C. Y. Tai, C. K. Wu, M. C. Chang – “Effects of Magnetic Field on the Crystallization of CaCO₃ using Permanent Magnets”, *Chemical Engineering Science* 63 (23) 5606 – 5612 (2008).
- ¹⁷⁴ C. Y. Tai, M. C. Chang, R. J. Shieh, T. G. Chen – “Magnetic Effects on Crystal Growth Rate of Calcite in a Constant-Composition Environment”, *Journal of Crystal Growth* 310 (15) 3690 – 3697 (2008).
- ¹⁷⁵ J. A. Gavira, J. Ma. Garca-Ruiz – “Effects of a Magnetic Field on Lysozyme Crystal Nucleation and Growth in a Diffusive Environment”, *Crystal Growth & Design* 9 (6) 2610 – 2615 (2009).
- ¹⁷⁶ M. D. Sikirić, H. Furedi-Milhofer – “The Influence of Surface Active Molecules on the Crystallization of Biominerals in Solution”, *Advances in Colloid and Interface Science* 128 – 130, 135 – 158 (2006).
- ¹⁷⁷ A. S. Posner, F. Betts, N. C. Blumenthal – “Formation and structure of synthetic and bone hydroxyapatite”, *Progress in Crystal Growth and Characterization of Materials* 3, 49 – 64 (1980).
- ¹⁷⁸ Lj. Brecević, V. Hlady, H. Furedi-Milhofer – “Influence of Gelatin on the Precipitation of Amorphous Calcium Phosphate”, *Colloids and Surfaces* 28, 301 - 313 (1987).
- ¹⁷⁹ I. V. Melikhov, S. Lazić, Ž. Vuković – “The Effect of Dissolved Impurity on Calcium Phosphate Nucleation in Supersaturated Medium”, *Journal of Colloid and Interface Science* 127 (2) 317 – 327 (1989).
- ¹⁸⁰ S. Lazić – “Microcrystalline Hydroxyapatite Formation from Alkaline Solutions”, *Journal of Crystal Growth* 147 (1-2) 147 – 154 (1995).
- ¹⁸¹ V. Privman, D. V. Goia, J. Park, E Matijević – “Mechanism of Formation of Monodispersed Colloids by Aggregation of Nanosize Precursors”, *Journal of Colloid and Interface Science* 213, 36 – 45 (1999).
- ¹⁸² V. Privman – “Growth of Nanosize and Colloid Particles by Controlled Addition of Singlets”, *Materials Research Society Symposium Proceeding* 703, 577 – 585 (2002).
- ¹⁸³ V. Privman – “Mechanisms of Diffusional Nucleation of Nanocrystals and Their Self-Assembly into Uniform Colloids”, *Annals of the New York Academy of Sciences* 1161, 508 – 526 (2009).
- ¹⁸⁴ E Matijević – “Nanosize Precursors as Building Blocks for Monodispersed Colloids”, *Colloid Journal* 69 (1) 29 – 38 (2007).
- ¹⁸⁵ M. Oh, C. A. Mirkin – “Chemically Tailorable Colloidal Particles from Infinite Coordination Polymers”, *Nature* 438, 651 – 654 (2005).

-
- ¹⁸⁶ D. T. Robb, I. Halaciuga, V. Privman, D. V. Goia – “Computational Model for the Formation of Uniform Silver Spheres by Aggregation of Nanosize Precursors”, *Journal of Chemical Physics* 129, 184705 (2008).
- ¹⁸⁷ K.-S. Cho, D. V. Talapin, W. Gaschler, C. B. Murray – “Designing PbSe Nanowires and Nanorings through Oriented Attachment of Nanoparticles”, *Journal of the American Chemical Society* 127 (19) 7140 – 7147 (2005).
- ¹⁸⁸ P. Alivisatos – “Chemical Transformations in Nanocrystals”, Presented at the Materials Research Society Spring Meeting, San Francisco, CA (2009).
- ¹⁸⁹ V. Jokanović, I. Nikčević, B. Dačić, D. Uskoković – “Synthesis of Nanostructured Carbonated Calcium Hydroxyapatite by Ultrasonic Spray Pyrolysis”, *Journal of Ceramic Processing Research* 5 (2) 157 – 162 (2004).
- ¹⁹⁰ L. C. Chow, L. Sun – “Properties of Nanostructured Hydroxyapatite Prepared by a Spray Drying Technique”, *Journal of the Research of the National Institute of Standards and Technology* 109 (6) 543 – 551 (2004).
- ¹⁹¹ C. Liu, Y. Huang, W. Shen, J. Cui – “Kinetics of Hydroxyapatite Precipitation at pH 10 to 11”, *Biomaterials* 22, 301 – 306 (2001).
- ¹⁹² S. Lazić, J. Katanić-Popović, S. Zec, N. Miljević – “Properties of Hydroxyapatite Crystallized from High Temperature Alkaline Solutions”, *Journal of Crystal Growth* 165, 124 – 128 (1996).
- ¹⁹³ S. Lazić, S. Zec, N. Miljević, S. Milonjić – “The Effect of temperature on the Properties of Hydroxyapatite Precipitated from Calcium Hydroxide and Phosphoric Acid”, *Thermochimica Acta* 374 (1) 13 – 22 (2001).
- ¹⁹⁴ M. J. Larsen – “Ion Products and Solubility of Calcium Phosphates”, Royal Dental College, Denmark (2001).
- ¹⁹⁵ W. Kibalczyk, A. Zielenkiewicz, W. Zielenkiewicz – “Calorimetric investigations of calcium phosphate precipitation in relation to solution composition and temperature”, *Thermochimica Acta* 131, 47 – 55 (1988).
- ¹⁹⁶ J. D. Termine, E. D. Eanes – “Comparative chemistry of amorphous and apatitic calcium phosphate preparation”, *Calcified Tissue Research* 10, 171–197 (1972).
- ¹⁹⁷ J. L. Meyer, C. C. Weatherall – “Amorphous to Crystalline Calcium Phosphate Transformation at Elevated pH”, *Journal of Colloid and Interface Science* 89 (1) 257 – 267 (1982).
- ¹⁹⁸ A. F. Marshall, K. R. Lawless – “TEM Study of the Central Dark Line in Enamel Crystallites”, *Journal of Dental Research* 60 (10) 1773 – 1782 (1981).
- ¹⁹⁹ M. E. Fernandez, J. Reyes-Gasga – “A Model for Human Tooth Enamel Central Dark Line Its HRTEM Images Contrast Transfer Function Analysis”, *Microscopy and Microanalysis* 9 (Suppl 2), 1290 – 1291 (2003).
- ²⁰⁰ J. Reyes-Gasga, C. M. Alcantara-Rodriguez – “Structural Analysis of the Human Tooth Enamel by Transmission Electron Microscopy”, In: *Hydroxyapatite and Related Materials*, edited by P. W. Brown & B. Constantz, pp. 295 – 304, CRC Press, Boca Raton, FL (1994).
- ²⁰¹ W. E. Brown, M. Mathew, L. C. Chow, Roles of OCP in Surface Chemistry of Apatites, In: *Adsorption on and Surface Chemistry of Apatites*, edited by Dwarika N. Misra, Springer, Berlin (1984).
- ²⁰² K. Chaari, S. Baklouti, J. Bouaziz, K. Bouzouita – “Optimizing the Dispersion of a Fluoroapatite Suspension Using Penta Sodium Triphosphate”, *Annales de Chimie – Science Des Materiaux* 29 (4) 1 – 13 (2004).
- ²⁰³ G. Yin, Z. Liu, J. Zhan, F. Ding, N. Yuan – “Impacts of the surface charge property on protein adsorption on hydroxyapatite”, *Chemical Engineering Journal* 87 (2) 181 – 186 (2002).

-
- ²⁰⁴ I. O. Smith, M. J. Baumann, L. R. McCabe – “Electrostatic Interactions as a Predictor for Osteoblast Attachment to Biomaterials”, *Journal of Biomedical Materials Research Part A* 70A, 436 – 441 (2004).
- ²⁰⁵ T. P. Feenstra, P. L. de Bruyn – “The Ostwald rule of stages in precipitation from highly supersaturated solutions: a model and its application to the formation of the nonstoichiometric amorphous calcium phosphate precursor phase”, *Journal of Colloid and Interface Science* 84 (1) 66 – 72 (1981).
- ²⁰⁶ A. L. Boskey, A. S. Posner – “Conversion of Amorphous Calcium Phosphate to Microcrystalline Hydroxyapatite. A pH-Dependent, Solution-Mediated, Solid-Solid Conversion”, *The Journal of Physical Chemistry* 77 (19) 2313 – 2317 (1973).
- ²⁰⁷ R. I. Martin, P. W. Brown – “Aqueous Formation of Hydroxyapatite”, *Journal of Biomedical Materials Research* 35 (33), 299-308 (1997).
- ²⁰⁸ W. Meier – “Nanostructure Synthesis Using Surfactants and Copolymers”, *Current Opinion in Colloid & Interface Science* 4, 6 – 14 (1999).
- ²⁰⁹ Y. E. Greish, P. W. Brown – “Phase Evolution during the Formation of Stoichiometric Hydroxyapatite at 37.4 °C”, *Journal of Biomedical Materials Research B: Applied Biomaterials* 67B, 632–637 (2003).
- ²¹⁰ K. S. tenHuisen, P. W. Brown – “Variations in solution chemistry during calcium-deficient and stoichiometric hydroxyapatite formation from $\text{CaHPO}_4 \times 2\text{H}_2\text{O}$ and $\text{Ca}_4(\text{PO}_4)_2\text{O}$ ”, *Journal of Biomedical Materials Research* 36, 233–241 (1997).
- ²¹¹ V. Uskoković, M. Drogenik – “Synthesis of Nanocrystalline Nickel-Zinc Ferrites via a Microemulsion Route”, *Materials Science Forum* 453 – 4, 225 – 230 (2004).
- ²¹² V. Uskoković, M. Drogenik – “A Mechanism for the Formation of Nanostructured NiZn Ferrites via a Microemulsion-Assisted Precipitation Method”, *Colloids and Surfaces A: Physicochemical and Engineering Aspects* 266, 168 – 174 (2005).
- ²¹³ H. Robbins - “The Preparation of Mn-Zn Ferrites by Co-Precipitation”, In: *Proceedings of the 3rd International Conference on Ferrites*, pp. 7-10, edited by H. Watanabe, S. Iida and M. Sugimoto, Reidel, Tokyo (1982).
- ²¹⁴ D. G. Grier – “When Like-Charges Attract: Interactions and Dynamics in Charge-Stabilized Colloidal Suspensions”, *Journal of Physics: Condensed Matter* 12, A85-A94 (2000).
- ²¹⁵ D. G. Grier – “Colloids: A Surprisingly Attractive Couple”, *Nature* 393 (6686) 621 (1998).
- ²¹⁶ W. Kunz, P. Lo Nostro, B. W. Ninham – “The Present State of Affairs with Hofmeister Effects”, *Current Opinion in Colloid and Interface Science* 9, 1 – 18 (2004).
- ²¹⁷ S. A. Edwards, D. R. M. Williams – “Hofmeister Effects in Colloid Science and Biology Explained by Dispersion Forces: Analytic Results for the Double Layer Interaction”, *Current Opinion in Colloid and Interface Science* 9, 139 – 144 (2004).
- ²¹⁸ B. Ninham – “Self-Assembly – Some Thoughts”, in *Self-Assembly*, edited by B. H. Robinson, IOS Press, Amsterdam (2003).
- ²¹⁹ H. Cölfen, S. Mann – “Higher-Order Organization by Mesoscale Self-Assembly and Transformation of Hybrid Nanostructures”, *Angewandte Chemie International Edition: English* 42, 2350 – 2365 (2003).
- ²²⁰ V. Uskoković – “Trends in Practical Colloid Science”, Nova Science Publishers, Hauppauge, NY (2009).

-
- ²²¹ P. N. De Aza, F. Guitian, A. Merlos, E. Lora-Tamayo, S. De Aza – “Bioceramics – Simulated Body Fluid Interfaces: pH and its Influence of Hydroxyapatite Formation”, *Journal of Materials Science: Materials in Medicine* 7, 399 – 402 (1996).
- ²²² L. T. Canham, C. L. Reeves, A. Loni, M. R. Houlton, J. P. Newey, A. J. Simons, T. I. Cox – “Calcium Phosphate Nucleation on Porous Silicon: Factors Influencing Kinetics in Acellular Simulated Body Fluids”, *Thin Solid Films* 297, 304 – 307 (1997).
- ²²³ T. Kokubo, H. Takadama – “How Useful is SBF in Predicting in vivo Bone Bioactivity?”, *Biomaterials* 27, 2907 – 2915 (2006).
- ²²⁴ I. V. Melikhov, S. Lazić, Ž. Vuković – “The Effect of Dissolved Impurity on Calcium Phosphate Nucleation in Supersaturated Medium”, *Journal of Colloid and Interface Science* 127 (2) 317 – 327 (1989).
- ²²⁵ J. A. Budz, M. LoRe, G. H. Nancollas – “Hydroxyapatite and Carbonated Apatite as Models for the Dissolution Behavior of Human Dental Enamel”, *Advances in Dental Research* 1 (2) 314 – 321 (1987).
- ²²⁶ L. Wang, X. Guan, C. Du, J. Moradian-Oldak, G. H. Nancollas – “Amelogenin Promotes the Formation of Elongated Apatite Microstructures in a Controlled Crystallization System”, *Journal of Physical Chemistry C* 111 (17) 6398 – 6404 (2007).
- ²²⁷ B. J. Tarasevich, C. J. Howard, J. L. Larson, M. L. Snead, J. P. Simmer, M. Paine, W. J. Shaw – “The Nucleation and Growth of Calcium Phosphate by Amelogenin”, *Journal of Crystal Growth* 304 (2) 407 – 415 (2007)
- ²²⁸ W. Wu, G. H. Nancollas – “Kinetics of Nucleation and Crystal growth of Hydroxyapatite and Fluoroapatite on Titanium Oxide Surfaces”, *Colloids and Surfaces B: Biointerfaces* 10, 87 – 94 (1997).
- ²²⁹ S. Koutsopoulos, E. Dalas – “Hydroxyapatite Crystallization in the Presence of Serine, Tyrosine and Hydroxyproline Amino Acids with Polar Side Groups”, *Journal of Crystal Growth* 216, 443 – 449 (2000).
- ²³⁰ R. S. Wagner, W. C. Ellis – “Vapor-Liquid-Solid Mechanism of Single Crystal Growth”, *Applied Physics Letters* 4 (5) 89 – 90 (1964).
- ²³¹ V. A. Sivakov, R. Scholz, F. Syrowatka, F. Falk, U. Gosele, S. H. Christiansen – “Silicon nanowire oxidation: the influence of sidewall structure and gold distribution”, *Nanotechnology* 20, 405607 (8pp) (2009).
- ²³² Zhang, J. W.; Nancollas, G. H. Mechanisms of Growth and Dissolution of Sparingly Soluble Salts. In: *Mineral-Water Interface Geochemistry*, M.F. Hochella, Jr. and A.F. White, Eds., *Reviews in Mineralogy*, 1990, 23, 365 – 396.
- ²³³ J. Moradian-Oldak – “Amelogenins: Assembly, Processing and Control of Crystal Morphology”, *Matrix Biology* 20, 293 – 305 (2001).
- ²³⁴ J. J. De Yoreo, P. G. Vekilov – “Principles of Crystal Nucleation and Growth”, *Reviews in Mineralogy and Geochemistry* 54 (1) 57 – 93 (2003).
- ²³⁵ V. Uskoković – “Isn't Self-Assembly a Misnomer? Multi-Disciplinary Arguments in Favor of Co-Assembly”, *Advances in Colloid and Interface Science* 141 (1-2) 37 - 47 (2008).
- ²³⁶ B. Mutafschiev – “The Atomistic Nature of Crystal Growth”, Springer, Berlin (2001).
- ²³⁷ A Saenz, M. L. Montero, G. Mondragon, V. Rodriguez-Lugo, V. M. Castano – “Effect of pH on the Precipitation of Hydroxyapatite on Silica Gels”, *Materials Research Innovation* 7, 68 – 73 (2003).

-
- ²³⁸ S. Hayakawa, Y. Li, K. Tsuru, A. Osaka, E. Fujii, K. Kawabata – “Preparation of Nanometer-Scale Rod Array of Hydroxyapatite Crystal”, *Acta Biomaterialia* 5, 2152 – 2160 (2009).
- ²³⁹ K. Tomoda, H. Ariizumi, T. Nakaji, K. Makino – “Hydroxyapatite Particles as Drug Carriers for Proteins”, *Colloids and Surfaces B: Biointerfaces* 76 (1) 226 – 235 (2010).
- ²⁴⁰ S. Koutsopoulos – “Synthesis and Characterization of Hydroxyapatite Crystals: A Review Study on the Analytical Methods”, *International Journal of Biomedical Materials Research* 62, 600 – 612 (2002).
- ²⁴¹ A. Barth, C. Zscherp – “What vibrations tell us about proteins” *Quarterly Review in Biophysics* 35 (4) 369 – 430 (2002).
- ²⁴² H. Tsuda, J. Arends – “Raman Spectra of Human Dental Calculus”, *Journal of Dental Research* 72, 1609 – 1613 (1993).
- ²⁴³ H. Tsuda, J. Arends – “Raman Spectroscopy in Dental Research: A Short Review of Recent Studies”, *Advances in Dental Research* 11, 539 – 547 (1997).
- ²⁴⁴ G. Penel, G. Leroy, C. Rey, E. Bres – “Micro Raman Spectral Study of the PO₄ and CO₃ Vibrational Modes in Synthetic and Biological Apatites”, *Calcified Tissue International* 63, 475 – 481 (1998).
- ²⁴⁵ A. Awonusi, D. M. Morris, M. J. Tecklenburg – “Carbonate Assignment and Calibration in the Raman Spectrum of Apatite”, *Calcified Tissue International* 81, 46 – 52 (2007).
- ²⁴⁶ I. R. Gibson, W. Bonfield – “Novel Synthesis and Characterization of an AB-Type Carbonate-Substituted Hydroxyapatite”, *Journal of Biomedical Materials Research* 59, 697 – 708 (2002).
- ²⁴⁷ R. Z. LeGeros, O. R. Trautz, E. Klein, J. P. LeGeros – “Two Types of Carbonate Substitution in the Apatite Structure”, *Experientia* 25, 5 – 7 (1969).
- ²⁴⁸ C. Rey, V. Renugopalakrishnan, B. Collins, M. V. Glimcher – “Fourier Transform Infrared Spectroscopic Study of the Carbonate Ions in Bone Mineral During Aging”, *Calcified Tissue International* 49, 251 – 258 (1991).
- ²⁴⁹ S. Marković, Lj. Veselinović, M. Lukić, Lj. Karanović, I. Bračko, N. Ignjatović, D. Uskoković – “Synthetical bone-like and biological hydroxyapatites: a comparative study of crystal structure and morphology”, *Biomedical Materials* 6 (2011) 045005.
- ²⁵⁰ B. Wopenka, J. D. Pasteris – “A Mineralogical Perspective on the Apatite in Bone”, *Materials Science and Engineering C* 25, 131 – 143 (2005).
- ²⁵¹ E. Schroder, T. Jonsson, L. Poole – “Hydroxyapatite chromatography: altering the phosphate-dependent elution profile of protein as a function of pH”, *Analytical Biochemistry* 313, 176 – 178 (2003).
- ²⁵² R. Giovannini, R. Freitag – “Comparison of Different Types of Ceramic Hydroxyapatite for the Chromatographic Separation of Plasmid DNA and a Recombinant Anti-Rhesus D Antibody”, *Bioseparation* 9 (6) 359 – 368 (2000).
- ²⁵³ S. Fujii, M. Okada, T. Furuzono – “Hydroxyapatite nanoparticles as stimulus-responsive particulate emulsifiers and building block for porous materials”, *Journal of Colloid and Interface Science* 315 (1) 287 – 296 (2007).
- ²⁵⁴ C. Phandugath – “Casein Micelle Structure: A Concise Review”, *Songklanakarinn Journal of Science and Technology* 27 (1) 201 – 212 (2005).
- ²⁵⁵ L. Sas, C. Tang, Z. Rengel – “Suitability of hydroxyapatite and iron phosphate as P sources for *Lupinus albus* grown in nutrient solution”, *Plant and Soil* 235 (2) 159 – 166 (2001).

-
- ²⁵⁶ J. Reichert, J. G. P. Binner – “An evaluation of hydroxyapatite-based filters for removal of heavy metal ions from aqueous solutions”, *Journal of Materials Science* 31 (5) 1231 – 1241 (2004).
- ²⁵⁷ V. Uskoković – “Nanotechnologies: What We Do Not Know”, *Technology in Society* 29 (1) 43 – 61 (2007).
- ²⁵⁸ L. Yang, B. W. Sheldon, T. J. Webber – “Nanophase Ceramics for Improved Drug Delivery”, *American Ceramic Society Bulletin* 89 (2) 24 – 31 (2010).
- ²⁵⁹ R. Duncan – “Nanomedicine Gets Clinical”, *Nano Today* 16 – 17 (August 2005).
- ²⁶⁰ R. Bawa – “Nanoparticles-based Therapeutics in Humans”, *Nanotechnology Law & Business* 135 - 155 (Summer 2008).
- ²⁶¹ S. D. Caruthers, S. A. Wickline, G. M Lanza – “Nanotechnological applications in medicine”, *Current Opinion in Biotechnology* 18:26–30 (2007).
- ²⁶² J.-Y Zhang, Z.-G. Shen, J. Zhong, T.-T. Hu, J.-F. Chen, Z.-Q. Ma, J. Yun – “Preparation of amorphous cefuroxime axetil nanoparticles by controlled nanoprecipitation method without surfactants”, *International Journal of Pharmaceutics* 323 (1-2) 153 – 160 (2006).
- ²⁶³ M. C. Branco, J. P. Schneider – “Self-assembling materials for therapeutic delivery”, *Acta Biomaterialia* 5, 817–831 (2009).
- ²⁶⁴ R. Korenstein – “Targeted drug delivery by nanoparticles and theranostics”, Presentation retrieved from www.tyndall.ie/n21/SRPA%20on%20targeted%20drug%20delivery%20and%20theranostics%20Cork%20Meeting (2009).
- ²⁶⁵ S. Bisht, G. Bhakta, S. Mitra, A. Maitra – “DNA loaded calcium phosphate nanoparticles: highly efficient non-viral vector for gene delivery”, *International Journal of Pharmaceutics* 288, 157–168 (2005).
- ²⁶⁶ M. Zhang, K. Kataoka – “Nano-Structured Composites Based on Calcium Phosphate for Cellular Delivery of Therapeutic and Diagnostic Agents”, *Nano Today* 4, 508 – 517 (2009).
- ²⁶⁷ M. Epple, K. Ganesan, R. Heumann, J. Klesing, A. Kovtun, S. Neumann, V. Sokolova – “Application of Calcium Phosphate Nanoparticles in Biomedicine”, *Journal of Materials Chemistry* 20, 18 – 23 (2010).
- ²⁶⁸ C. M. Wiethoff, C. R. Middaugh – “Barriers to Nonviral Gene Delivery”, *Journal of Pharmaceutical Science* 92 (2) 203 – 217 (2003).
- ²⁶⁹ B. Roszek, W. H. de Jong, R. E. Geertsma – “Nanotechnology in medical applications: state-of-the-art in materials and devices”, *RIVM report* 265001001 (2005).
- ²⁷⁰ Q. Zhang, D. Zhao, X.-Z. Zhang, S.-X. Cheng, R.-X. Zhuo – “Calcium Phosphate/DNA Co-Precipitates Encapsulated Fast-Degrading Polymer Films for Substrate-Mediated Gene Delivery”, *International Journal of Biomedical Material Research Part B: Applied Biomaterials* 91B, 172 – 180 (2009).
- ²⁷¹ A. Watson, D. Latchman – »Gene Delivery into Neuronal Cells by Calcium Phosphate-Mediated Transfection«, *Methods: A Companion to Methods in Enzymology* 10, 289 – 291 (1996).
- ²⁷² E. H. Chowdhury, T. Akaike – »Fibronectin-Coated Nano-Precipitates of Calcium-Magnesium Phosphate for Integrin-Targeted Gene Delivery«, *Journal of Controlled Release* 116 (2) e68 – e69 (2006).
- ²⁷³ M. Okazaki, Y. Yoshida, S. Yamaguchi, M. Kaneno, J. C. Elliott – »Affinity Binding Phenomena of DNA onto Apatite Crystals«, *Biomaterials* 22 (18) 2459 – 2464 (2001).

-
- ²⁷⁴ D. Olton, J. Li, M. E. Wilson, T. Rogers, J. Close, L. Huang, P. N. Kumta, C. Sfeir – “Nanostructured Calcium Phosphates (NanoCaPs) for Non-Viral Gene Delivery: Influence of the Synthesis Parameters on Transfection Efficiency», *Biomaterials* 28 (6) 1267 – 1279 (2007).
- ²⁷⁵ A. H. Faraji, P. Wipf – “Nanoparticles in Cellular Drug Delivery”, *Bioorganic & Medicinal Chemistry* 17, 2950 – 2963 (2009).
- ²⁷⁶ M. Kester, Y. Heakal, T. Fox, A. Sharma, G. P. Robertson, T. T. Morgan, Erhan I. Altinoglu, A. Tabakovic, M. R. Parette, S. M. Rouse, V. Ruiz-Velasco, J. H. Adair – “Calcium Phosphate Nanocomposite Particles for In Vitro Imaging and Encapsulated Chemotherapeutic Drug Delivery to Cancer Cells”, *Nano Letters* 8 (12) 4116 – 4121 (2008).
- ²⁷⁷ N. Singh, B. Manshian, G. J. S. Jenkins, S. M. Griffiths, P. M. Williams, T. G. G. Maffei, C. J. Wright, S. H. Doak – “NanoGenotoxicology: The DNA Damaging Potential of Engineered Nanomaterials”, *Biomaterials* 30, 3891 – 3914 (2009).
- ²⁷⁸ H. Cölfen, S. Mann – “Higher-Order Organization by Mesoscale Self-Assembly and Transformation of Hybrid Nanostructures”, *Angewandte Chemie International Edition: English* 42, 2350 – 2365 (2003).
- ²⁷⁹ S. V. Dorozhkin – “A Hierarchical Structure for Apatite Crystals”, *Journal of Materials Science: Materials in Medicine* 18, 363 – 366 (2007).
- ²⁸⁰ G. K. Lim, J. Wang, S. C. Ng, L. M. Gan – “Processing of fine hydroxyapatite powders via an inverse microemulsion route”, *Materials Letters* 28, 431 – 436 (1996).
- ²⁸¹ V. Ball, J.-M. Planeix, O. Flix, J. Hemmerl, P. Schaaf, M. W. Hosseini, J. C. Voegel – “Molecular Tectonics: Abiotic Control of Hydroxyapatite Crystals Morphology”, *Crystal Growth & Design* 2 (6) 489-492 (2002).
- ²⁸² M. Jevtić, M. Mitrić, S. Škapin, B. Jančar, N. Ignjatović, D. Uskoković, “Crystal structure of hydroxyapatite nano-rods synthesized by sonochemical homogenous precipitation”, *Crystal Growth and Design* 8 (7) 2217 – 2222 (2008).
- ²⁸³ C. E. Fowler, M. Li, S. Mann, H. C. Margolis – “Influence of surfactant assembly on the formation of calcium phosphate materials—A model for dental enamel formation”, *Journal of Materials Chemistry* 15, 3317-3325 (2005).
- ²⁸⁴ P. Wang, C. Li, H. Gong, X. Jiang, H. Wang, K. Li – “Effects of synthesis conditions on the morphology of hydroxyapatite nanoparticles produced by wet chemical process”, *Powder Technology* 203, 315 – 321 (2010).
- ²⁸⁵ V. Uskoković – “Insights into Morphological Nature of Precipitation of Cholesterol”, *Steroids* 73, 356 – 369 (2008).
- ²⁸⁶ Kathleen E. Fischer, Benjamin J. Aleman, Sarah L. Tao, R. Hugh Daniels, Esther M. Li, Mark D. Bunker, Ganesh Nagaraj, Parminder Singh, Alex Zettl, Tejal A. Desai – “Biomimetic Nanowire Coatings for Next Generation Adhesive Drug Delivery Systems”, *Nano Letters* 9 (2) 716-720 (2009).
- ²⁸⁷ P. Ducheyne, S. Radin, L. King – “The Effect of Calcium Phosphate Ceramic Composition and Structure on in vitro Behavior. I. Dissolution”, *Journal of Biomedical Materials Research* 27, 25 – 34 (1993).
- ²⁸⁸ S. Yamada, D. Heymann, J. M. Boulter, G. Daculsi – “Osteoclastic Resorption of Calcium Phosphate Ceramic with Different Hydroxyapatite/ β -Tricalcium Phosphate Ratios”, *Biomaterials* 18, 1037 – 1041 (1997).
- ²⁸⁹ A. J. Melville, L. M. Rodriguez-Lorenzo, J. S. Forsythe – “Effects of Calcination Temperature on the Drug Delivery Behaviour of Ibuprofen from Hydroxyapatite Powders”, *Journal of Materials Science – Materials in Medicine* 19 (3) 1187 – 1195 (2008).

-
- ²⁹⁰ S. Raynaud, E. Champion, D. Bernache-Assollant – “Characterization of Hydroxyapatite-Tricalcium Phosphate Bioceramics Issued from Ca-Deficient Hydroxyapatite Powders: Influence of Ca/P Ratio”, *Phosphorus Research Bulletin* 10, 214 – 218 (1999).
- ²⁹¹ S. Raynaud, E. Champion, D. Bernache-Assollant, P. Thomas – “Calcium Phosphate Apatites with Variable Ca/P Atomic Ratio I. Synthesis, Characterisation and Thermal Stability of Powders”, *Biomaterials* 23, 1065 – 1072 (2002).
- ²⁹² W. Gamble – “Atherosclerosis: the carbonic anhydrase, carbon dioxide, calcium concerted theory”, *Journal of Theoretical Biology* 239 (1) 16 – 21 (2006).
- ²⁹³ V. Uskoković – “Surface Charge Effects Involved in the Control of Stability of Sols Comprising Uniform Cholesterol Particles”, *Materials and Manufacturing Processes* 23 (6) 620 – 623 (2008).
- ²⁹⁴ B. M. Craven – “Crystal Structure of Cholesterol Monohydrate”, *Nature* 260, 727 – 9 (1976).
- ²⁹⁵ S. V. Dorozhkin, M. Epple – “Biological and Medical Significance of Calcium Phosphates”, *Angewandte Chemie International Edition* 41 (17) 3130 – 3146 (2002).
- ²⁹⁶ B. Sumer, J. Gao – “Theranostic nanomedicine for cancer”, *Nanomedicine* 3 (2) 137 – 140 (2008).
- ²⁹⁷ A. R. Boccaccini, J. J. Blaker – “Bioactive Composite Materials for Tissue Engineering Scaffolds”, *Expert Reviews in Medical Devices* 2 (3) 303 – 317 (2005).
- ²⁹⁸ D. A. Wahl, E. Sachlos, C. Liu, J. T. Czernuszka – “Controlling the Processing of Collagen-Hydroxyapatite Scaffolds for Bone Tissue Engineering”, *Journal of Materials Science: Materials in Medicine* 18 (2) 201 – 209 (2007).
- ²⁹⁹ S. Best – “Nanostructured Apatites: The Next Generation of Bioactive Materials”, Presented at the 10th YUCOMAT Conference in Herceg-Novi, Montenegro (2008).
- ³⁰⁰ W.-Q. Yan, T. Nakamura, M. Kobayashi, H.-M. Kim, F. Miyaji, T. Kokubo – “Bonding of Chemically Treated Titanium Implants to Bone”, *Journal of Biomedical Materials Research Part A* 37 (2) 267 – 275 (1998).
- ³⁰¹ F.-Z. Cui, T.-F. Jiang – “Tissue-Engineered Bone”, in: Wiley Encyclopedia of Biomedical Engineering, John Wiley & Sons, New York, NY (2009).
- ³⁰² S. M. Best, A. E. Porter, E. S. Thian, J. Huang – “Bioceramics: Past, Present and for the Future”, *Journal of the European Ceramic Society* 28 (7) 1319 – 1327 (2008).
- ³⁰³ N. Ignjatović, C. Liu, J. Czernuszka, D. Uskoković - “Micro and nano/injectable composite biomaterials containing calcium phosphate coated with poly(dl-lactide-co-glycolide)”, *Acta Biomaterialia* 3 (6) 927-935 (2007).
- ³⁰⁴ G. Daculsi, A. P. Uzel, P. Weiss, E. Goyenvalle, E. Aguado – “Developments in Injectable Multiphasic Biomaterials. The Performance of Microporous Biphasic Calcium Phosphate Granules and Hydrogels”, *Journal of Materials Science: Materials in Medicine* 21, 855 – 861 (2010).
- ³⁰⁵ H. Doty, W. Haggard, H. Courtney, J. Bumgardner – “Effectiveness of a Dual Drug Delivery Calcium Sulfate, Chitosan-Calcium Phosphate Bone Scaffold”, Society for Biomaterials 2010 Annual Meeting & Exposition, Seattle, WA (2010), presentation #677.
- ³⁰⁶ C. Combes, C. Rey – “Amorphous Calcium Phosphates: Synthesis, Properties and Uses in Biomaterials”, *Acta Biomaterialia* 6, 3362 – 3378 (2010).

-
- ³⁰⁷ J. Huang, Y. W. Lin, X. W. Fu, S. M. Best, R. A. Brooks, N. Rushton, W. Bonfield – “Development of Nano-Sized Hydroxyapatite Reinforced Composites for Tissue Engineering Scaffolds”, *Journal of Materials Science: Materials in Medicine* 18 (11) 2151 – 2157 (2007).
- ³⁰⁸ X. Zheng, S. Zhou, Y. Xiao, X. Yu, B. Feng – “In Situ Preparation and Characterization of a Novel Gelatin/Poly(D, L-Lactide)/Hydroxyapatite Nanocomposite”, *Journal of Biomedical Materials Research Part B: Applied Biomaterials* 91B, 181 – 190 (2009).
- ³⁰⁹ N. Ignjatović, S. Tomić, M. Dakić, M. Miljković, M. Plavšić and D. Uskoković - ”Synthesis and Properties of Hydroxyapatite/poly-L-lactide Composite Biomaterials”, *Biomaterials* 20, 809-816 (1999).
- ³¹⁰ N. Ignjatović, P. Ninkov, V. Kojić, M. Bokurov, V. Srdić, D. Krnojelac, S. Selaković, D. Uskoković - “Cytotoxicity and fibroblast properties during in vitro test of biphasic calcium phosphate/poly-dl-lactide-co-glycolide biocomposites and different phosphate materials”, *Microscopy Research and Technique* 69 (12) 976-982 (2006).
- ³¹¹ M. Vukomanović, M. Mitrić, S.D. Škapin, E. Žagar, J. Plavec, N. Ignjatović, D. Uskoković – “Influence of ultrasonic processing on the macromolecular properties of poly (D,L-lactide -co-glycolide) alone and in its biocomposite with hydroxyapatite”, *Ultrasonics Sonochemistry* 17 (5) 902 – 908 (2010).
- ³¹² M. Stevanović, T. Maksin, J. Petković, M. Filipič, D. Uskoković, "An innovative, quick and convenient labeling method for the investigation of pharmacological behavior and the metabolism of poly(DL-lactide-co-glycolide) nanospheres", *Nanotechnology* 20 (33) 335102 (2009).
- ³¹³ V. Uskoković, N. Ignjatović, N. Petranović – “Synthesis and Characterization of Hydroxyapatite-Collagen Biocomposite Materials”, *Materials Science Forum* 413, 269 – 274 (2003).
- ³¹⁴ J. D. Kretlow, S. Young, L. Klouda, M. Wong, A. G. Mikos – “Injectable Biomaterials for Regenerating Complex Craniofacial Tissues”, *Advanced Materials* 21, 1 – 26 (2009).
- ³¹⁵ T. Miyai, A. Ito, G. Tamazawa, T. Matsuno, Y. Sogo, C. Nakamura, A. Yamazaki, T. Satoh – “Antibiotic-loaded poly-ε-caprolactone and porous β-tricalcium phosphate composite for treating osteomyelitis” *Biomaterials* 29, 350–358 (2008).
- ³¹⁶ V. C. A. Martins, G. Goissis, A. C. Ribeiro, E. Marcantonio Jr, M. R. Bet – “The controlled release of antibiotic by hydroxyapatite: anionic collagen composites”, *Artificial Organs* 22, 215-221 (1998).
- ³¹⁷ M. A. Rauchmann, T. A. Wichelhaus, V. Stirnal, E. Dingeldein, L. Zichner, R. Schnettler, V. Alt – “Nanocrystalline hydroxyapatite and calcium sulphate as biodegradable composite carrier material for local delivery of antibiotics in bone infections”, *Biomaterials* 26, 2677-2684 (2005).
- ³¹⁸ A. El-Kamel, M. Baddour – “Gatifloxacin biodegradable implant for treatment of experimental osteomyelitis: in vitro and in vivo evaluation”, *Drug Delivery* 14:349-356 (2007).
- ³¹⁹ N. L. Ignjatović, P. Ninkov, R. Sabetrsekh, D. P. Uskoković – “A Novel Nano Drug Delivery System based on Tigecycline-Loaded Calcium Phosphate Coated with Poly-DL-Lactide-co-Glycolide”, *Journal of Materials Science: Materials in Medicine* 21 (1) 231 – 239 (2010).
- ³²⁰ J. Hylands – “Tigecycline: a new antibiotic”, *Intensive and Critical Care Nursing* 24, 260 – 263 (2008).
- ³²¹ S. K. Nandi, B. Kundu, S. K. Ghosh, T. K. Mandal, S. Datta, D. K. De, D. Basu – “Cefuroxime-impregnated calcium phosphates as an implantable delivery system in experimental osteomyelitis”, *Ceramics International* 35, 1367–1376 (2009).

-
- ³²² Q. G. Xu, J. I. Czernuszka – “Controlled release of Amoxicillin from Hydroxyapatite-Coated Poly(Lactic-Co-Glycolic Acid) Microspheres”, *Journal of Controlled Release* 127 (2) 146 – 153 (2008).
- ³²³ Q. G. Xu, Y. Tanaka, J. T. Czernuszka – “Encapsulation and Release of a Hydrophobic Drug from Hydroxyapatite Coated Liposomes”, *Biomaterials* 28 (16) 2687 – 2694 (2007).
- ³²⁴ S. P. Victor, T. S. S. Kumar – “Tailoring Calcium-Deficient Hydroxyapatite Nanocarriers for Enhanced Release of Antibiotics”, *Journal of Biomedical Nanotechnology* 4 (2) 203 – 209 (2008).
- ³²⁵ M. Stevanović, A. Radulović, B. Jordović, D. Uskoković, “Poly(DL-lactide-co-glycolide) Nanospheres for the Sustained Release of Folic Acid”, *Journal of Biomedical Nanotechnology* 4 (3) 349-358 (2008).
- ³²⁶ M. M. Stevanović, B. Jordović, D. P. Uskoković, “Preparation and characterization of poly(D,L-lactide-co-glycolide) microparticles containing ascorbic acid”, *Journal of Biomedicine and Biotechnology*, Article ID 84965 (2007).
- ³²⁷ M. Stevanović, J. Savić, B. Jordović, D. Uskoković, "Fabrication, in vitro degradation and the release behaviours of poly(DL-lactide-co-glycolide) nanospheres containing ascorbic acid", *Colloids and Surfaces B: Biointerfaces* 59, 215-223 (2007).
- ³²⁸ M. Stevanović, D. Uskoković – “Influence of Different Degradation Medium on Release of Ascorbic Acid from Poly(D,L-Lactide-co-Glycolide) Nano- and Microspheres”, *Russian Journal of Physical Chemistry A* 83 (9) 1457 – 1460 (2009).
- ³²⁹ B. J. Nehilla, M. Bergkvist, K. C. Papat, T. A. Desai – “Purified and surfactant-free coenzyme Q10-loaded biodegradable nanoparticles”, *International Journal of Pharmaceutics* 348, 107–114 (2008).
- ³³⁰ Y. Cai, H. Pan, X. Xu, Q. Hu, L. Li and R. Tang – “Ultrasonic Controlled Morphology Transformation of Hollow Calcium Phosphate Nanospheres: A Smart and Biocompatible Drug Release System”, *Chemistry of Materials* 19 (13) 3081 – 3083 (2007).
- ³³¹ M. Y. Ma, Y. J. Zhu, L. Li, S. W. Cao – “Nanostructured Porous Hollow Ellipsoidal Capsules of Hydroxyapatite and Calcium Silicate: Preparation and Application in Drug Delivery”, *Journal of Materials Chemistry* 18 (23) 2722 – 2727 (2008).
- ³³² A. A. Akasha, I. Sotiriadou, M. X. Doss, M. Halbach, J. Winkler, J. J.S. Baunach, A. Katsen-Glob, H. Zimmermann, Y. Chood, J. Hescheler, A. Sachinidis – “Entrapment of Embryonic Stem Cells-Derived Cardiomyocytes in Macroporous Biodegradable Microspheres: Preparation and Characterization”, *Cell Physiology and Biochemistry* 22:665-672 (2008).
- ³³³ S. Tielens, H. Declercq, T. Gorski, E. Lippens, E. Schacht, M. Cornelissen – “Gelatin-Based Microcarriers as Embryonic Stem Cell Delivery System in Bone Tissue Engineering: An in-Vitro Study”, *Biomacromolecules* 8, 825-832 (2007).
- ³³⁴ R. S. Ashton, A. Banerjee, S. Punyani, D. V. Schaffer, R. S. Kane – “Scaffolds based on degradable alginate hydrogels and poly(lactide-co-glycolide) microspheres for stem cell culture”, *Biomaterials* 28, 5518–5525 (2007).
- ³³⁵ S. Liao, C. K. Chan, S. Ramakrishna – “Stem Cells and Biomimetic Materials Strategies for Tissue Engineering”, *Materials Science and Engineering C* 28, 1189 – 1202 (2008).
- ³³⁶ Y. Yanga, F. M.V. Rossib, E. E. Putninsa – “Ex vivo expansion of rat bone marrow mesenchymal stromal cells on microcarrier beads in spin culture”, *Biomaterials* 28, 3110–3120 (2007).

-
- ³³⁷ M. Matsusaki, K. Kadowaki, K. Tateishi, C. Higuchi, W. Ando, D. A. Hart, Y. Tanaka, Y. Take, M. Akashi, H. Yoshikawa, N. Nakamura – “Scaffold-free tissue-engineered construct-hydroxyapatite composites generated by an alternate soaking process: potential for repair of bone defects”, *Tissue Engineering A* 15 (1) 55 – 63 (2009).
- ³³⁸ R. Gonzales-McQuire, J.-Y. Chane-Ching, E. Vignaud, A. Lebugle, S. Mann – “Synthesis and Characterization of Amino Acid-Functionalized Hydroxyapatite Nanorods”, *Journal of Materials Chemistry* 14, 2277 – 2281 (2004).
- ³³⁹ K. Wormuth – »Superparamagnetic Latex via Inverse Emulsion Polymerization«, *Journal of Colloid and Interface Science* 241, 366 – 377 (2001).
- ³⁴⁰ R. M. El-Backly, A. G. Massoud, A. M. El-Badry, R. A. Sherif, M. K. Marei – “Regeneration of dentine/pulp-like tissue using a dental pulp stem cell/poly(lactic-co-glycolic) acid scaffold construct in New Zealand white rabbits”, *Australian Endodontic Journal* 34: 52–67 (2008).
- ³⁴¹ J. Chevalier, L. Gremillard – “Ceramics for medical applications: A picture for the next 20 years”, *Journal of the European Ceramic Society* 29, 1245–1255 (2009).
- ³⁴² T. Kokubo, T. Matsushita, H. Takadama, T. Kizuki – “Development of bioactive materials based on surface chemistry”, *Journal of the European Ceramic Society* 29, 1245–1255 (2009).
- ³⁴³ M. Teller, U. Gopp, H. G. Neumann, K. D. Kuhn – Release of Gentamicin from Bone Regenerative Materials: An in vitro Study”, *Journal of Biomedical Materials Research Part B: Applied Biomaterials* 81B, 23 – 29 (2007).
- ³⁴⁴ F. Grasset, N. Labhsetwar, D. Li, D. C. Park, N. Saito, H. Haneda, O. Cador, T. Roisnel, S. Mornet, E. Duguet, J. Portier, J. Etourneau – »Synthesis and Magnetic Characterization of Zinc Ferrite Nanoparticles with Different Environments: Powder, Colloidal Solution, and Zinc Ferrite-Silica Core-Shell Nanoparticles«, *Langmuir* 18 (21) 8209 – 8216 (2002).
- ³⁴⁵ T. Tago, T. Hatsuta, K. Miyajima, M. Kishida, S. Tashiro, K. Wakabayashi – »Novel Synthesis of Silica-Coated Ferrite Nanoparticles Prepared Using Water-in-Oil Microemulsion”, *Journal of American Ceramic Society* 85 (9), 2188 – 94 (2002).
- ³⁴⁶ F. J. Arriagada, K. Osseo-Asare – »Controlled Hydrolysis of Tetraethoxysilane in a Nonionic Water-in-Oil Microemulsion: A Statistical Model of Silica Nucleation«, *Colloids and Surfaces A: Physicochemical and Engineering Aspects* 154, 311 – 326 (1999).
- ³⁴⁷ V. Uskoković, A. Košak, M. Drogenik – “Silica-Coated Lanthanum-Strontium Manganites for Hyperthermia Treatments”, *Materials Letters* 60 (21-22) 2620 – 2622 (2006).
- ³⁴⁸ V. Uskoković, E. Matijević – “Uniform Particles of Pure and Silica Coated Cholesterol”, *Journal of Colloid and Interface Science* 315 (2) 500 – 511 (2007).
- ³⁴⁹ K. Park, S. Lee, E. Kang, K. Kim, K. Choi, I. C. Kwon – “New Generation of Multifunctional Nanoparticles for Cancer Imaging and Therapy”, *Advanced Functional Materials* 19, 1 – 14 (2009).
- ³⁵⁰ L. Jongpaiboonkit, T. Franklin-Ford, W. L. Murphy – “Mineral-Coated Polymer Microspheres for Controlled Protein Binding and Release”, *Advanced Materials* 21, 1 – 4 (2009).
- ³⁵¹ M. Vukomanović, S. Škapin, B. Jančar, T. Maksin, N. Ignjatović, V. Uskoković, D. Uskoković – “Poly(d,l-lactide-co-glycolide)/Hydroxyapatite Core-Shell Nanospheres. Part 1: A Multifunctional System for Controlled Drug Delivery”, *Colloids and Surfaces B: Biointerfaces* (in press, 2010).
- ³⁵² S. Fischer, C. Foerg, H. P. Merkle, B. Gander – “Chitosan Coated PLGA-Microspheres - A Modular System for Targeted Drug Delivery”, *European Cells and Materials* 7, Suppl. 2, 11 – 12 (2004).

-
- ³⁵³ M. M. Stevanović, D. P. Uskoković – “Poly(lactide-co-glycolide)-based micro and nanoparticles for the controlled drug delivery of vitamins”, *Current Nanoscience*, 5 (1) 1 - 14 (2009).
- ³⁵⁴ H. T. Ong, J. S. C. Loo, F. Y. C. Boey, S. J. Russell, J. Ma, K. W. Peng – “Exploiting the high-affinity phosphonate-hydroxyapatite nanoparticle interaction for delivery of radiation and drugs”, *Journal of Nanoparticle Research* 10 (1) 141 – 150 (2008).
- ³⁵⁵ T. T. Morgan, H. S. Muddana, E. I. Altinoglu, S. M. Rouse, A. Tabakovic, T. Tabouillot, T. J. Russin, S. S. Shanmugavelandy, P. J. Butler, P. C. Eklund, J. K. Yun, M. Kester, J. H. Adair – “Encapsulation of Organic Molecules in Calcium Phosphate Nanocomposite Particles for Intracellular Imaging and Drug Delivery”, *Nano Letters* 8 (12) 4108 – 4115 (2008).
- ³⁵⁶ P. P. Yang, Z. W. Quan, C. X. Li, X. J. Kang, H. Z. Lian, J. Lin – “Bioactive, luminescent and mesoporous europium-doped hydroxyapatite as a drug carrier”, *Biomaterials* 29 (32) 4341 – 4347 (2008).
- ³⁵⁷ O. A. Graeve, R. Kanakala, A. Madadi, B. C. Williams, K. C. Glass – “Luminescence Variations in Hydroxyapatites Doped with Eu²⁺ and Eu³⁺ Ions”, *Biomaterials* 31, 4259 – 4267 (2010).
- ³⁵⁸ S. P. Mondejar, A. Kovtun and M. Epple – “Lanthanide-doped calcium phosphate nanoparticles with high internal crystallinity and with a shell of DNA as fluorescent probes in cell experiments”, *Journal of Materials Chemistry* 17, 4153 – 4159 (2007).
- ³⁵⁹ A. Doat, M. Fanjul, F. Pelle, E. Hollande, A. Lebugle – “Europium-Doped Bioapatite: A New Photostable Biological probe, Internalizable by Human Cells”, *Biomaterials* 24, 3365– 3371 (2003).
- ³⁶⁰ V. Sokolova, A. Kovtun, R. Heumann, M. Epple – “Tracking the pathway of calcium phosphate/DNA nanoparticles during cell transfection by incorporation of red-fluorescing tetramethylrhodamine isothiocyanate–bovine serum albumin into these nanoparticles”, *Journal of Biological Inorganic Chemistry* 12, 174 – 179 (2007).
- ³⁶¹ B. J. Nehilla, P. G. Allen, T. A. Desai – “Surfactant-free, drug-quantum-dot coloaded poly(lactide-co-glycolide) nanoparticles: towards multifunctional nanoparticles”, *ACS Nano* 2 (3) 528 – 544 (2008).
- ³⁶² Y. Guo, D. L. Shi, J. Lian, Z. Y. Dong, W. Wang, H. S. Cho, G. K. Liu, L. M. Wang, R. C. Ewing – “Quantum Dot Conjugated Hydroxylapatite nanoparticles for In Vivo Imaging”, *Nanotechnology* 19 (17) 175102 (2008).
- ³⁶³ V. Sokolova, A. Kovtun, R. Heumann, M. Epple – “ Tracking the pathway of calcium phosphate/DNA nanoparticles during cell transfection by incorporation of red-fluorescing tetramethylrhodamine isothiocyanate–bovine serum albumin into these nanoparticles”, *Journal of Biological Inorganic Chemistry* 12, 174 – 179 (2007).
- ³⁶⁴ R. Z. LeGeros – “Biological and Synthetic Apatites”, In: *Hydroxyapatite and Related Materials*, edited by P. W. Brown & B. Constantz, pp. 3 – 28, CRC Press, Boca Raton, FL (1994).
- ³⁶⁵ S. S. Banerjee, M. S. Tarafder, A. Bandyopadhyay, S. Bose – “Tricalcium Phosphate Based Resorbable Ceramics: Influence of MgO and SrO Addition on Mechanical Properties and Biocompatibility”, Society for Biomaterials 2010 Annual Meeting & Exposition, Seattle, WA (2010), presentation #435.
- ³⁶⁶ M. Bohner – “Silicon-Substituted Calcium Phosphates – A Critical View”, *Biomaterials* 30 (32) 6403 – 6406 (2009).
- ³⁶⁷ I. D. Xynos, A. J. Edgar, L. D. K. Buttery, L. L. Hench, J. M. Polak – “Gene-expression profiling of human osteoblasts following treatment with the ionic products of Bioglass (R) 45S5 dissolution”, *Journal of Biomedical Materials Research* 55 (2) 151-157 (2001).
- ³⁶⁸ L. L. Hench – “Genetic Design of Bioactive Glasses”, *Journal of the European Ceramic Society* 29 (7) 1257 – 1265 (2009).

-
- ³⁶⁹ L. L. Hench, J. M. Polak – “Third-Generation Biomedical Materials”, *Science* 295 (5557) 1014 – 1017 (2002).
- ³⁷⁰ Lj. Veselinović, Lj. Karanović, Z. Stojanović, I. Bračko, S. Marković, N. Ignjatović, D. Uskoković – “Crystal Structure of Cobalt-Substituted Calcium Hydroxyapatite Nanopowders Prepared by Hydrothermal Processing”, *Journal of Applied Crystallography* 43, 320 – 327 (2010).
- ³⁷¹ Z. Stojanović, Lj. Veselinović, S. Marković, N. Ignjatović, D. Uskoković – “Hydrothermal Synthesis of Nanosize Pure and Cobalt-Exchanged Hydroxyapatite”, *Materials and Manufacturing Processes* 24 (10-11) 1096 – 1103 (2009).
- ³⁷² H.-C. Wu, T.-W. Wang, J.-S. Sun, W.-H. Wang, F.-H. Lin – “A Novel Biomagnetic Nanoparticle Based on Hydroxyapatite”, *Nanotechnology* 18, 165601 – 165610 (2007).
- ³⁷³ H. R. Low, N. Phonthammachai, A. Maignan, G. A. Stewart, T. J. Bastow, L. L. Ma, T. J. White – “The Crystal Chemistry of Ferric Oxyhydroxyapatite”, *Inorganic Chemistry* 47 (24) 11774 – 11782 (2008).
- ³⁷⁴ X. Liu, M. D. Kaminski, H. Chen, M. Torno, L. Taylor, A. J. Rosengart - “Synthesis and Characterization of Highly-Magnetic Biodegradable Poly(D,L-lactide-co-glycolide) Nanospheres”, *Journal of Controlled Release* 119, 52–58 (2007).
- ³⁷⁵ M. S. Tarafder, A. Bandyopadhyay, S. Bose – “Osteoblast Interactions on Electrically Polarized Biphasic Calcium Phosphate”, Society for Biomaterials 2010 Annual Meeting & Exposition, Seattle, WA (2010), presentation #234.
- ³⁷⁶ N. Bock, A. Riminucci, C. Dionigi, A. Russo, A. Tampieri, E. Landi, V. A. Goranov – “A Novel Route in Bone Tissue Engineering: Magnetic Biomimetic Scaffolds”, *Acta Biomaterialia* 6 (3) 786 – 796 (2010).
- ³⁷⁷ M. Misono, W. K. Hall – “Oxidation–Reduction Properties of Copper- and Nickel-Substituted Hydroxyapatites”, *Journal of Physical Chemistry* 77, 791–800 (1973).
- ³⁷⁸ M. Wakamura, K. Kandori, T. Ishikawa – “Surface structure and composition of calcium hydroxyapatites substituted with Al(III), La(III) and Fe(III) ions”, *Colloids and Surfaces A* 164 297–305 (2000).
- ³⁷⁹ V. Uskoković, A. Košak, M. Drogenik – “Preparation of Silica-Coated Lanthanum-Strontium Manganite Particles with Designable Curie Point, for Application in Hyperthermia Treatments”, *International Journal of Applied Ceramic Technology* 3 (2) 134 – 143 (2006).
- ³⁸⁰ C.-H. Hou, S.-M. Hou, Y.-S. Hsueh, J. Lin, H.-C. Wu, F.-H. Lin – “The in vivo performance of biomagnetic hydroxyapatite nanoparticles in cancer hyperthermia therapy”, *Biomaterials* 30, 3956 – 3960 (2009).
- ³⁸¹ V. Uskoković, M. Drogenik, I. Ban – “The Characterization of Nanosized Nickel-Zinc Ferrites Synthesized within Reverse Micelles of CTAB/1-Hexanol/Water Microemulsion”, *Journal of Magnetism and Magnetic Materials* 284, 294 – 302 (2004).
- ³⁸² H.-C. Wu, T.-W. Wang, M. C. Bohn, F.-H. Lin, M. Spector – “Novel Magnetic Hydroxyapatite Nanoparticles as Non-Viral Vectors for the Glial Cell Line-Derived Neurotrophic Factor Gene”, *Advanced Functional Materials* 20, 67 – 77 (2010).
- ³⁸³ H. Chen, M. D. Kaminski, X. Liu, C. J. Mertz, Y. Xie, M. D. Torno, A. J. Rosengart – “A novel human detoxification system based on nanoscale bioengineering and magnetic separation techniques”, *Medical Hypotheses* 68, 1071–1079 (2007).
- ³⁸⁴ G. A. Silva – “Nanotechnology approaches to crossing the blood-brain barrier and drug delivery to the CNS”, *BMC Neuroscience* 10 (9) Suppl 3:S4 (2008).

-
- ³⁸⁵ R. Bawa – “NanoBiotech 2008: Exploring Global Advances in Nanomedicine ”, *Nanomedicine* 5 (1) 5 - 7 (2009).
- ³⁸⁶ L. N. Patel, J. L. Zaro, W. C. Shen – “Cell penetrating peptides: intracellular pathways and pharmaceutical perspectives”, *Pharmaceutical Research* 24, 1977 – 1992 (2007).
- ³⁸⁷ S. E. Gratton, P. A. Ropp, P. D. Pohlhaus, J. C. Luft, V. J. Madden, M. E. Napier – “The effect of particle design on cellular internalization pathways”, *Proceedings of the National Academy of Sciences* 105, 11613 – 11618 (2008).
- ³⁸⁸ B. D. Chithrani, W. C. Chan – “Elucidating the mechanism of cellular uptake and removal of protein-coated gold nanoparticles of different sizes and shapes”, *Nano Letters* 7, 1542 – 1550 (2007).
- ³⁸⁹ A. S. Lubbe, C. Bergemann, J. Brock, D. G. McClure – “Physiological Aspects in Magnetic Drug-Targeting”, *Journal of Magnetism and Magnetic Materials* 194, 149–155 (1999).
- ³⁹⁰ Q. A. Pankhurst, J. Connolly, S. K. Jones, J. Dobson – “Applications of Magnetic Nanoparticles in Biomedicine” *Journal of Physics D* 36 R167–R181 (2003).
- ³⁹¹ A. S. Mistry, A. G. Mikos - “Tissue Engineering Strategies for Bone Regeneration”, *Advances in Biochemical Engineering/Biotechnology* 94: 1–22 (2005).
- ³⁹² O. Harush-Frenkel, N. Debotton, S. Benita, Y. Altschuler – “Targeting of nanoparticles to the clathrin-mediated endocytic pathway”, *Biochemical and Biophysical Research Communication* 353, 26 – 32 (2007).
- ³⁹³ J. Klesing, A. Wiehe, B. Gitter, S. Grafe, M. Eppe – “Positively Charged Calcium Phosphate/Polymer Nanoparticles for Photodynamic Therapy”, *Journal of Materials Science: Materials in Medicine* 21, 887 – 892 (2010).
- ³⁹⁴ M. Mrksich – “Using self-assembled monolayers to model the extracellular matrix”, *Acta Biomaterialia* 5, 832–841 (2009).
- ³⁹⁵ S. Tang, A. Jin, Y. Wang – “Preparation of plastic nano-hydroxyapatite/poly (3-hydroxybutyrate-hydroxyvalerate)-polyethylene glycol gentamicin drug delivery system”, *Zhongguo Xiu Fu Chong Jian Wai Ke Za Zhi* 20 (7) 758 – 761 (2006).
- ³⁹⁶ M. L. Ho, Y. C. Fu, G. J. Wang, H. T. Chen, J. K. Chang, T. H. Tsai, C. K. Wang – “Controlled release carrier of BSA made by W/O/W emulsion method containing PLGA and hydroxyapatite”, *Journal of Controlled Release* 128 (2) 142 – 148 (2008).
- ³⁹⁷ O. C. Wilson Jr., J. R. Hull – “Surface Modification of Nanophase Hydroxyapatite with Chitosan”, *Materials Science and Engineering C* 28 (3) 434 – 437 (2008).
- ³⁹⁸ G. Zhou, Y. Li, L. Zhang, Y. Zuo, J. A. Jansen – “Preparation and characterization of nano-hydroxyapatite/chitosan/konjac glucomannan composite”, *Journal of Biomedical Materials Research Part A* 83A (4) 931 – 939 (2007).
- ³⁹⁹ M. L. Tan, P. F. Choong, C. R. Dass – “DNAzyme Delivery Systems: Getting past First Base”, *Expert Opinion on Drug Delivery* 6 (2) 127 – 138 (2009).
- ⁴⁰⁰ C. R. Dass, P. F. M. Choong – “The use of chitosan formulations in cancer therapy”, *Journal of Microencapsulation* 25 (4) 275 — 279 (2003).

-
- ⁴⁰¹ B. T. Reves, J. D. Bumgardner, J. A. Cole, Y. Yang, W. O. Haggard – “Lyophilization to improve drug delivery for chitosan-calcium phosphate bone scaffold construct: a preliminary investigation”, *Journal of Biomedical Materials Research B* 90 (1) 1 – 10 (2009).
- ⁴⁰² J. Adair – “Nano Delivery of Cancer Therapeutics & siRNA”, Presentation retrieved from nanotechinstitute.org/wp-content/cornwall_keystone-nano-10-25.ppt (2009).
- ⁴⁰³ U. Mukesh, V. Kulkarni, R. Tushar, R. S. R. Murthy – “Methotrexate Loaded Self Stabilized Calcium Phosphate Nanoparticles: A Novel Inorganic Carrier for Intracellular Drug Delivery”, *Journal of Biomedical Nanotechnology* 5 (1) 99 – 105 (2009).
- ⁴⁰⁴ X. Cheng, L. Kuhn – “Chemotherapy Drug Delivery from Calcium Phosphate Nanoparticles”, *International Journal of Nanomedicine* 2 (4) 667 - 674 (2007).
- ⁴⁰⁵ J.-M. Oh, S.-J. Choi, G.-E. Lee, S.-H. Han, J.-H. Choy – “Inorganic Drug-Delivery Nanovehicle Conjugated with Cancer-Cell-Specific Ligand”, *Advanced Functional Materials* 19, 1 – 8 (2009).
- ⁴⁰⁶ K. Baba, H. E. Pudavar, I. Roy, T. Y. Ohulchanskyy, Y. Chen, R. K. Pandey, P. N. Prasad – “New method for delivering a hydrophobic drug for photodynamic therapy using pure nanocrystal form of the drug”, *Molecular Pharmacy* 4 (2) 289 – 297 (2007).
- ⁴⁰⁷ A. L. Koh, C. M. Shachaf, S. Elchuri, G. P. Nolan, R. Sinclair – “Electron microscopy localization and characterization of functionalized composite organic-inorganic SERS nanoparticles on leukemia cells”, *Ultramicroscopy* 109, 111 – 121 (2008).
- ⁴⁰⁸ S.-H. Cheng, C.-H. Lee, C.-S. Yang, F.-G. Tseng, C.-Y. Mou and L.-W. Lo – “Mesoporous silica nanoparticles functionalized with an oxygen-sensing probe for cell photodynamic therapy: potential cancer theranostics”, *Journal of Materials Chemistry* 19 (9) 1252 – 1257 (2009).
- ⁴⁰⁹ V. V. Sokolova, I. Radtke, R. Heumann, M. Epple – “Effective transfection of cells with multi-shell calcium phosphate-DNA nanoparticles”, *Biomaterials* 27 (16) 3147 – 3153 (2006).
- ⁴¹⁰ A. L. Mazin – “Hydroxyapatite Thin-Layer Chromatography of Nucleic Acid”, *Molecular Biology* 11 (3) 477 – 498 (1977).
- ⁴¹¹ P. Gagnon – “Improved antibody aggregate removal by hydroxyapatite chromatography in the presence of polyethylene glycol”, *Journal of Immunological Methods* 336 (2) 222 – 228 (2008).
- ⁴¹² C. Fargues, M. Bailly, G. Grevillot – “Adsorption of BSA and Hemoglobin on Hydroxyapatite Support: Equilibria and Multicomponent Dynamic Adsorption”, *Adsorption – Journal of the International Adsorption Society* 4 (1) 5 – 16 (1998).
- ⁴¹³ V. Sokolova, M. Epple – “Inorganic Nanoparticles as Carriers of Nucleic Acids into Cells”, *Angewandte Chemie International Edition* 47, 1382 – 1395 (2008).
- ⁴¹⁴ D. N. Nguyen, J. J. Green, J. M. Chan, R. Langer, D. G. Anderson – “Polymeric Materials for Gene Delivery and DNA Vaccination” *Advanced Materials* 21, 847–867 (2009).
- ⁴¹⁵ F. L. Graham, A. J. van der Eb – “A new technique for the assay of infectivity of human adenovirus 5 DNA”, *Virology* 63, 456–467 (1973).
- ⁴¹⁶ C. R. Dass, P. F. M. Choong – “The use of Chitosan Formulations in Cancer Therapy”, *Journal of Microencapsulation* 25(4): 275–279 (2008).

-
- ⁴¹⁷ S. Zhu, K. Zhou, B. Huang, S. Huang, F. Liu, Y. Li, Z. Xue, Z. Long – “Hydroxyapatite Nanoparticles: A Novel Material of Gene Carrier”, *Sheng Wu Yi Xue Gong Cheng Xue Za Zhi* 22 (5) 980 – 984 (2005).
- ⁴¹⁸ V. Sokolova, O. Prymak, W. Meyer-Zaika, H. Cölfen, H. Rehage, A. Shukla, M. Epple – “Synthesis and characterization of DNA-functionalized calcium phosphate nanoparticles”, *Mat.-wiss. u. Werkstofftech.* 37, 6 (2006).
- ⁴¹⁹ J. Schwiertz, W. Meyer-Zaika, L. Ruiz-Gonzalez, J. M. Gonzalez-Calbet, M. Vallet-Regi, Matthias Epple – “Calcium phosphate nanoparticles as templates for nanocapsules prepared by the layer-by-layer technique“, *Journal of Materials Chemistry* 18 (32) 152 (2008).
- ⁴²⁰ X. Bai, K. More, C. M. Rouleau, A. Rabiei – “Functionally Graded Hydroxyapatite Coatings Doped with Antibacterial Components”, *Acta Biomaterialia* 6 (6) 2264 – 2273 (2010).
- ⁴²¹ B. Gopalakrishnan, J. Wolff – “siRNA and DNA Transfer to Cultured Cells”, *Methods in Molecular Biology* 480, 31 – 52 (2009).
- ⁴²² M. Zhang, A. Ishii, N. Nishiyama, S. Matsumoto, T. Ishii, Y. Yamasaki, K. Kataoka – “PEGylated Calcium Phosphate Nanocomposites as Smart Environment-Sensitive Carriers for siRNA Delivery”, *Advanced Materials* 21, 1 – 6 (2009).
- ⁴²³ D. S. Seferos, A. E. Prigodich, D. A. Giljohann, P. C. Patel, C. A. Mirkin – “Polyvalent DNA Nanoparticle Conjugates Stabilize Nucleic Acids”, *Nano Letters* 9 (1) 308 – 311 (2009).
- ⁴²⁴ J. Ren, P. Zhao, T. Ren, S. Gu, K. Pan – “Poly (D,L-lactide)/nano-hydroxyapatite composite scaffolds for bone tissue engineering and biocompatibility evaluation”, *Journal of Materials Science: Materials in Medicine* 19 (3) 1075 – 1082 (2008).
- ⁴²⁵ M. Wang, Y. Li, J. Wu, F. Xu, Y. Zuo, J. A. Jansen – “In vitro and in vivo study to the biocompatibility and biodegradation of hydroxyapatite/poly(vinyl alcohol)/gelatin composite”, *Journal of Biomedical Materials Part A* 85 (2) 418 – 426 (2008).
- ⁴²⁶ S. Najman, V. Savić, Lj. Djordjević, N. Ignjatović, D. Uskoković - "Biological evaluation of hydroxyapatite/poly-L-lactide (HAp/PLLA) composite biomaterials with poly-L-lactide of different molecular weights intraperitoneally implanted into mice", *Bio-Medical Materials and Engineering* 14, 61-70 (2004).
- ⁴²⁷ N. Ignjatović, V. Savić, S. Najman, M. Plavšić, D. Uskoković - “Study of HAp/PLLA Composite as a Substitute for Bone Powder, using FT-IR Spectroscopy”, *Biomaterials* 22, 571-575 (2001).
- ⁴²⁸ R. F. Service – “Nanomaterials Show Signs of Toxicity”, *Science* 300: 243 (2003).
- ⁴²⁹ G. Oberdörster, E. Oberdörster, J. Oberdörster – “Nanotoxicology: an emerging discipline evolving from studies of ultrafine particles”, *Environmental Health Perspectives* 113 (7): 823–839 (2005).
- ⁴³⁰ F. Stellacci – “On the Role of Interfacial Energy in Bio-interfaces”, Presentation at the Materials Research Society Conference, San Francisco, CA, April 2010.
- ⁴³¹ K. Park, S. Lee, E. Kang, K. Kim, K. Choi, I. C. Kwan – “New Generation of Multifunctional Nanoparticles for Cancer Imaging and Therapy”, *Advanced Functional Materials* 19, 1 – 14 (2009).
- ⁴³² D. Shi, H. S. Cho, Y. Chen, H. Xu, H. Gu, J. Lian, W. Wang, H. Liu, C. Huth, L. Wang, R. C. Ewing, S. Budko, G. M. Pauletti, Z. Dong – “Fluorescent Polystyrene-Fe₃O₄ Composite Nanospheres for In Vivo Imaging and Hyperthermia”, *Advanced Materials* 21, 1 – 4 (2009).

⁴³³ J.-H. Lee, K. Lee, S. H. Moon, Y. Lee, T. G. Park, J. Cheon – “All-in-One Target-Cell-Specific Magnetic Nanoparticles for Simultaneous Molecular Imaging and siRNA Delivery”, *Angewandte Chemie International Edition: English* 48, 4174 – 4179 (2009).

⁴³⁴ A. Ashokan, D. Menon, S. Nair, M. Koyakutty – “A Molecular Receptor Targeted, Hydroxyapatite Nanocrystal Based Multi-Modal Contrast Agent”, *Biomaterials* 31 (9) 2606 – 1616 (2010).

⁴³⁵ J. Kim, S. Park, J. E. Lee, S. M. Jin, J. H. Lee, I. S. Lee, I. Yang, J. S. Kim, S. K. Kim, M. H. Cho, T. Hyeon – “Designed Fabrication of Multifunctional Magnetic Gold Nanoshells and Their Application to Magnetic Resonance Imaging and Photothermal Therapy”, *Angewandte Chemie International Edition* 45, 7754 – 7758 (2006).